Novel CREB3L3 Nonsense Mutation in a Family With Dominant Hypertriglyceridemia

Angelo B. Cefalu,* Rossella Spina,* Davide Noto,* Vincenza Valenti, Valeria Ingrassia, Antonina Giammanco, Maria D. Panno, Antonina Ganci, Carlo M. Barbagallo, Maurizio R. Averna

Objective—Cyclic AMP responsive element–binding protein 3–like 3 (CREB3L3) is a novel candidate gene for dominant hypertriglyceridemia. To date, only 4 kindred with dominant hypertriglyceridemia have been found to be carriers of 2 nonsense mutations in CREB3L3 gene (245fs and W46X). We investigated a family in which hypertriglyceridemia displayed an autosomal dominant pattern of inheritance.

Approach and Results—The proband was a 49-year-old woman with high plasma triglycerides (≤1300 mg/dL; 14.68 mmol/L). Her father had a history of moderate hypertriglyceridemia, and her 51-year-old brother had triglycerides levels as high as 1600 mg/dL (18.06 mmol/L). To identify the causal mutation in this family, we analyzed the candidate genes of recessive and dominant forms of primary hypertriglyceridemia by direct sequencing. The sequencing of CREB3L3 gene led to the discovery of a novel minute frame shift mutation in exon 3 of CREB3L3 gene, predicted to result in the formation of a truncated protein devoid of function (c.359delG–p.K120fsX20). Heterozygosity for the c.359delG mutation resulted in a severe phenotype in the proband, and her brother with a late in life expression and a good response to diet and a hypotriglyceridemic treatment. The same mutation was detected in a 13-year-old daughter who to date is normotriglyceridemic.

Conclusions—We have identified a novel pathogenic mutation in CREB3L3 gene in a family with dominant hypertriglyceridemia with a variable pattern of penetrance. (Arterioscler Thromb Vasc Biol. 2015;35:00-00.
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Key Words: codon, nonsense ▪ heterozygote ▪ hypertriglyceridemia ▪ mutation ▪ triglycerides
to date, 4 kindred with dominant hypertriglyceridemia associated with CREB3L3 heterozygous mutations (245fs and W46X) have been described. Hypertriglyceridemia because of CREB3L3 pathogenic variants might represent a rare example of monogenetic dominant hypertriglyceridemia. Here, we report a family harboring a novel mutation of CREB3L3 gene, which shows a dominant pattern of inheritance and a variable expression of the hypertriglyceridemia trait (Table 1).

Materials and Methods
Materials and Methods are available in the online-only Data Supplement.

Results
Phenotype
DNA samples were available from 6 subjects across 2 generations. Lipid profiles of the family members are presented in Figure 1. The proband (subject II-1) and her brother (II-3) showed moderate hypertriglyceridemia, whereas her sister (II-4) and her daughters (III-1 and III-2) showed a normal lipid profile (Figure 1). Because the triglyceride concentrations in the proband and her brother were >400 mg/dL (>4.52 mmol/L) and we have not performed direct low-density lipoprotein cholesterol measurement, the levels of low-density lipoprotein cholesterol are not provided for these subjects.

The proband’s response to treatment, including diet and various regimens of lipid-lowering medication, during the period between 2009 and 2015 is shown in Figure 2.

During this period, plasma triglyceride levels showed large fluctuations ranging from 150 to 1300 mg/dL (1.69–14.67 mmol/L). These differences were mainly attributed to the poor compliance to low-fat diet and hypolipidemic treatment with ω-3 fatty acids (EPA+DHA, 3 g/d) and fenofibrate (200 mg/d). On the occasion of our fist evaluation, triglyceride plasma levels were 500 mg/dL (5.64 mmol/L); because of the reluctance of the patient to be treated with fibrates, a treatment with 4 g of ω-3 fatty acids supplementation was prescribed. Under ω-3 fatty acids therapy, she showed an improved lipid profile with triglyceride plasma levels as low as 213 mg/dL after 4 months of treatment (2.40 mmol/L, 43% reduction). For personal reason between February and July 2014, she discontinued the ω-3 fatty acids supplementation with a subsequent relapse of hypertriglyceridemia (plasma triglyceride, 537 mg/dL–6.06 mmol/L). She restarted the treatment and the triglyceride plasma levels decreased to 151 mg/dL (1.7 mmol/L; Figure 2).

9ω-3 fatty acids therapy was also prescribed to subject II-3 (proband’s brother), but no follow-up data are available.

Genetic Analysis
There were 6 individuals in the family who had both plasma lipids phenotype and DNA available for genotyping. Of these 6 individuals, the proband (II-1) and her brother (II-3) were affected on clinical grounds (triglyceride >175 mg/dL). No functionally relevant mutations in the LPL, APOC2, APOA5, GPIHBP1, and LMF1 genes were detected.

These results and the clinical suspicion of a dominant form of hypertriglyceridemia prompted us to analyze by direct sequencing the whole CREB3L3 gene.

The proband was found to be heterozygous for a novel frameshift mutation in exon 3 of CREB3L3 gene, predicted to result in the formation of a truncated protein devoid of function (c.359delG–p.K120fsX20). The presence of the mutation in exon 3 (Figure 3) was confirmed in 3 independent polymerase chain reaction amplifications and direct sequencing. The proband’s hypertriglyceridemic brother (II-3) and one of her normotriglyceridemic daughter (III-2) were found to be carriers of the same mutation in the heterozygous state (Figure 1).

Even if the c.359delG is predicted to generate a premature stop codon and it is expected to be a rare and deleterious variant, we screened this mutation by direct sequencing in 30 unrelated healthy Italian normolipidemic individuals; the mutation was not found in any of these subjects. Because this analysis was performed in the same ethnic background population of the proband, we cannot draw any certain conclusion about the frequency of the mutation in other ethnic groups.

More, the variant was not reported in the Exome Variant Server repository of the NHLBI Exome Sequencing Project (http://evs.gs.washington.edu/EVS/), the Exome Aggregation Consortium Website (ExAC, http://exac.broadinstitute.org), and the 1000 Genomes web catalog (http://www.1000genomes.org), and we did not detect any variant

Table 1. Primers Used for Sequencing the CREB3L3 Gene

<table>
<thead>
<tr>
<th>Exon</th>
<th>Primer</th>
<th>Sequence</th>
<th>T, °C</th>
<th>Size, bp</th>
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<td>CTGTCACCACTCAGCAGCT</td>
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</tr>
<tr>
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<td>R</td>
<td>CATTGGACCTTTCCTTGG</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>F</td>
<td>GCACTGACACTCTAGCCGGA</td>
<td>60</td>
<td>348</td>
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<tr>
<td></td>
<td>R</td>
<td>GCACGTGAGTGGACGAGA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>F</td>
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<td></td>
</tr>
<tr>
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</tr>
<tr>
<td></td>
<td>R</td>
<td>CCAGGCTACGTGGCAAGAAA</td>
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<tr>
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<td>F</td>
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<td></td>
<td>R</td>
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<tr>
<td>7</td>
<td>F</td>
<td>GGCTTGGGTTTCTTCTG</td>
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<tr>
<td></td>
<td>R</td>
<td>CATCAGAGGGCTCTTCTCT</td>
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<tr>
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<td>AAAGGACGTGGTGAGAGCA</td>
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<td>9</td>
<td>F</td>
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<tr>
<td>10</td>
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<td>480</td>
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<tr>
<td></td>
<td>R</td>
<td>GTCTGGGCTCCACTGCC</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

F indicates forward; and R, reverse.

Nonstandard Abbreviations and Acronyms

CREB3L3  cyclic AMP responsive element–binding protein 3–like 3
CREBH  CRE-binding protein H
LPL  lipoprotein lipase
Cefalù et al. CREB3L3 Mutation in Hypertriglyceridemia

by the analysis of whole CREB3L3 gene in other 7 unrelated patients with moderate hypertriglyceridemia, suggesting that the c.359delG is a rare mutation.

In order to try to explain the different lipid phenotype in mutation carriers, we investigated the possible genetic modulating effect on triglyceride levels by APOE genotypes and APOC3 variants. All investigated subjects were carriers of the E2/E3 genotype, but the proband who was E2/E4; direct sequencing of the whole APOC3 gene did not reveal any variant.

Discussion

In this study, we report a novel mutation of CREB3L3 gene, the c.359delG, in 3 related members (mother, daughter, and...

Figure 1. Pedigree of the nuclear family with hypertriglyceridemia. The proband is indicated by an arrow. Half filled symbols denote affected subjects, carriers of the mutation c.359delG of the CREB3L3 gene; empty symbols, noncarriers subjects; and ?, data unavailable. For subjects II-1 and II-3 low-density lipoprotein (LDL) cholesterol levels are not provided because the triglyceride (TG) concentrations were >400 mg/dL (>4.52 mmol/L) and direct LDL cholesterol measurement was not performed. HDL indicates high-density lipoprotein; and WT, wild type.

Figure 2. Proband’s plasma triglycerides levels and response to treatment. *ω-3 fatty acids (EPA+DHA) 4 g/d.
mother’s brother) of a family with a different and variable pattern of penetrance. Heterozygosity for the c.359delG mutation resulted in a severe hypertriglyceridemia phenotype in the proband and her brother with a late in life expression and a good response to diet and hypotriglyceridemic treatments based on fenofibrate and ω-3 fatty acids. The same mutation was detected in the daughter, 13 years old, who to date is normotriglyceridemic.

CREB-H was first identified by Abel et al. CREB-H is a liver-specific bZIP transcription factor that is selectively expressed in the liver and in the small intestine. CREB-H binds to the CRE consensus (TGACGTCA) and box-B–like elements present in the promoters of liver-expressed genes. CREB-H regulated genes are involved in acute phase response, iron absorption, and gluconeogenesis. In the hepatic lipid metabolism, notably in lipogenesis and triglyceride metabolism. The first evidence came from primary mouse hepatocytes incubated with fatty acids. In this experimental setting, fatty acids induced CREB-H mRNA hepatic expression, and this effect was attributed to the involvement of peroxisome proliferator–activated receptor α. Moreover CREB3L3−/− knockout mice show a 3-fold higher plasma triglyceride levels compared with wild-type mice suggesting a direct involvement of CREB-H in triglyceride metabolism. Additional experiments in CREB3L3−/− mice demonstrated that CREB-H does not affect hepatic very low-density lipoprotein secretion rate and that LPL activity was reduced indicating that hypertriglyceridemia in these mice is a consequence of an impaired triglyceride clearance from plasma. The infusion of wild-type animals plasma into CREB-H–deficient mice contributed to ameliorate the hypertriglyceridemia phenotype, suggesting that CREB-H effects on triglyceride clearance were mediated by circulating cofactors for LPL. In fact CREB-H controls the expression of genes involved in triglyceride metabolism, including APOC2, APOA4, and APOA5, and the reduced expression of these known cofactors for LPL in CREB3L3−/− mice could give reasons for the hypertriglyceridemia phenotype. Moreover, CREB-H via peroxisome proliferator–activated receptor α induces FGF21, a liver expressed hormone, which increases insulin sensitivity and reduces plasma triglyceride levels. In early large genome-wide association studies CREB3L3 gene did

**Table 2. TG Levels in Carriers of CREB3L3 Mutations**

<table>
<thead>
<tr>
<th>Pedigree Generation</th>
<th>Sex (M/F)</th>
<th>Age, y</th>
<th>TG Levels, mg/dL–mmol/L</th>
<th>BMI, kg/m²</th>
<th>Mutation</th>
<th>Reference</th>
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</thead>
<tbody>
<tr>
<td>1 Proband (II-5)</td>
<td>M</td>
<td>62</td>
<td>4513–51</td>
<td>30.3</td>
<td>245fs</td>
<td>7</td>
</tr>
<tr>
<td>II-2</td>
<td>F</td>
<td>67</td>
<td>189–2.1</td>
<td>27.5</td>
<td>245fs</td>
<td>7</td>
</tr>
<tr>
<td>II-3</td>
<td>M</td>
<td>65</td>
<td>235–2.6</td>
<td>27.5</td>
<td>245fs</td>
<td>7</td>
</tr>
<tr>
<td>II-4</td>
<td>F</td>
<td>64</td>
<td>238–2.7</td>
<td>27</td>
<td>245fs</td>
<td>7</td>
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<tr>
<td>II-6</td>
<td>M</td>
<td>58</td>
<td>280–3.2</td>
<td>25.1</td>
<td>245fs</td>
<td>7</td>
</tr>
<tr>
<td>II-7</td>
<td>F</td>
<td>56</td>
<td>102–1.1</td>
<td>23.6</td>
<td>245fs</td>
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</tr>
<tr>
<td>2 Proband (I-1)</td>
<td>M</td>
<td>42</td>
<td>2433–27.5</td>
<td>36</td>
<td>245fs</td>
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<tr>
<td>I-2</td>
<td>M</td>
<td>15</td>
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<td>3 Proband (I-1)</td>
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<td>245fs</td>
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<tr>
<td>4 Proband (I-1)</td>
<td>M</td>
<td>43</td>
<td>583–6.6</td>
<td>29.7</td>
<td>W46X</td>
<td>7</td>
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<tr>
<td>II-1</td>
<td>M</td>
<td>91</td>
<td>131–1.5</td>
<td>21.8</td>
<td>W46X</td>
<td>7</td>
</tr>
<tr>
<td>5 Proband (II-1)</td>
<td>F</td>
<td>49</td>
<td>537–6.1</td>
<td>22.5</td>
<td>K120fsX20</td>
<td>This report</td>
</tr>
<tr>
<td>II-3</td>
<td>M</td>
<td>51</td>
<td>553–6.2</td>
<td>25</td>
<td>K120fsX20</td>
<td>This report</td>
</tr>
<tr>
<td>II-2</td>
<td>F</td>
<td>13</td>
<td>74–0.8</td>
<td>20</td>
<td>K120fsX20</td>
<td>This report</td>
</tr>
</tbody>
</table>

F indicates female; M, male; and TG, triglyceride.
not emerge as a locus associated with the triglyceride trait. When this gene was resequenced in a cohort of 449 unrelated hypertriglyceridemia subjects, 12 individuals were identified as heterozygous carriers of nonsynonymous or insertional CREB3L3 gene mutations. The analysis of the 4 unrelated nonsense mutation carriers allowed the identification of 7 additional carriers; the triglyceride mean plasma levels of the 11 carriers was significantly higher than those of the noncarriers family members (9.67 versus 1.66 mmol/L, 856 versus 147 mg/dL). Interestingly among the 11 heterozygotes for CREB3L3 nonsense mutations, 3 carriers were normotriglyceridemic, 4 had a mild-to-moderate hypertriglyceridemia, and only 2 had a severe hypertriglyceridemia (Table 2). These latter observations might suggest that other factors could contribute to the penetrance of the CREB3L3 genetic defect.

Recently, Johansen et al resequenced CREB3L3 in 413 hypertriglyceridemia subjects and 241 nondiabetic control and found an enrichment of CREB3L3 rare noncoding, synonymous, and nonsynonymous variants. Three of them (W46X, E240K, and K245Efs374X) were definitely damaging. Unfortunately, it was not reported the severity degree of the hypertriglyceridemia-associated phenotype. The family we described represents the fifth kindred in which a heterozygous frame shift mutation (c.359delG→p.K120fsX20) of CREB3L3 is associated with a severe hypertriglyceridemia phenotype.

Overall 7 rare variants have been reported in CREB3L3 gene so far. Among these, only 2 lead to premature stop codon (W46X and 245fs) and both have been shown to alter the CREB-H protein function (Table 3) and to segregate in pedigrees with hypertriglyceridemia in families. The nonsense mutation we describe is expected to lead to a CREB-H protein with an intermediate size compared with the 2 previously described mutations and although we did not perform functional studies it can be speculated a deleterious effect on function.

Our proband was healthy, and the only relevant clinical feature was the presence of fatty liver detected by ultrasonography. This finding is interesting because of the absence of overweight and insulin resistance. In CREB3L3 knock-out mice fed an atherogenic diet, Zhang et al demonstrated the development of hepatic steatosis underlining the role of CREB-H in the regulation of hepatic lipogenesis, lipolysis, and fatty acid oxidation. The proband’s daughter (subject III-2) is carrier of the mutation but her triglyceride levels were normal. The analysis of the 5 families (including the present) raises the issue of the penetrance of the heterozygous loss-of-function mutations of CREB3L3 gene. Because the majority of the heterozygotes have a mild-to-moderate hypertriglyceridemia, why do only few heterozygotes develop a severe hypertriglyceridemia form? To answer this question, we can rely only on the few published data. The 2 severe hypertriglyceridemia subjects belonging to 2 unrelated kindred harboring the CREB3L3 W46X kindred, the proband was overweight but the hypertriglyceridemia form was mild-to-moderate. Among the other 8 sporadic heterozygotes, carriers of CREB3L3 nonsense mutations, 5 showed a mild-to-moderate form of hypertriglyceridemia (n.2 G105R, n.1 P166L, n.1 D182N, and n.1 E240K) and 3 (n.1 P166L, n.1 V180M, n.1 E240K) a severe form of hypertriglyceridemia. It is worth to note that only 1 P166L heterozygote was diabetic. Overall, based on these sparse clinical information, it is difficult to draw any conclusion about the role as precipitating factors of overweight, obesity, and diabetic mellitus. An accurate follow-up of the identified heterozygotes would significantly contribute to a better understanding of the mechanisms that underlie the clinical expression of the CREB3L3 genetic defect.

The proband of our kindred presented a severe form of hypertriglyceridemia. The response to treatment with fenofibrate (200 mg/d) and ω-3 fatty acids (3 g/d) was excellent, and interestingly when fenofibrate was stopped a mild-to-moderate hypertriglyceridemia developed. This response to the treatment suggests that fenofibrate, as previously demonstrated in vitro, could stimulate the expression of CREB-H via the peroxisome proliferator–activated receptor α pathway in humans.

In conclusion, we identified a novel nonsense mutation of CREB3L3 gene, the c.359delG→p.K120fsX20, which was associated with a severe hypertriglyceridemia phenotype and showed a variable penetrance. No functional variants were identified in the other major known hypertriglyceridemia-associated genes. Moreover, we investigated the possible modulating action of APOE common genotypes and APOC3 gene variants on triglyceride plasma levels. No APOC3 gene variants were identified in all family members; thus, we ruled out an effect on triglyceride levels in mutation carriers. Subjects carriers of the APOE2 allele show higher triglyceride

<table>
<thead>
<tr>
<th>Status</th>
<th>Variation in cDNA</th>
<th>Effect on CREB-H Protein (Amino Acids)</th>
<th>Mutation Type</th>
<th>Functional Impairment Evaluation In Vitro</th>
<th>Reference</th>
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</thead>
<tbody>
<tr>
<td>He</td>
<td>c.359delG</td>
<td>K120fsX20</td>
<td>Small deletion</td>
<td>No</td>
<td>This report</td>
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<tr>
<td>He</td>
<td>c.138G&gt;A</td>
<td>W46X</td>
<td>Nonsense</td>
<td>Yes. Precluded translation of the DNA binding and bZIP domains, and failure to transactivate reporter genes</td>
<td>7</td>
</tr>
<tr>
<td>He</td>
<td>c.733insG</td>
<td>K245E→374X</td>
<td>Complex mutation which consists of a G insertion in the first nucleotide of codon 245 together with a A&gt;T point mutation 7 nucleotides downstream</td>
<td>Yes. Precluded translation of the DNA binding and bZIP domains, and failure to transactivate reporter genes</td>
<td>7</td>
</tr>
<tr>
<td>He</td>
<td>c.834G→A</td>
<td>E240K</td>
<td>Missense</td>
<td>Yes. Precluded translation of the DNA binding and bZIP domains, and failure to transactivate reporter genes</td>
<td>7</td>
</tr>
</tbody>
</table>

CREB-H indicates cyclic AMP-responsive element–binding protein H.
levels compared with the more common APOE3 allele carriers. The 3 mutation carriers (subjects II-1, II-3, and III-2) of this family are carriers of the APOE2 allele and in addition 2 of them (II-3 and III-2) share the same genotype (E2/E3), indicating that the variability of triglyceride plasma levels is not related to APOE genotype.

Further studies are warranted to identify putative modulators of the CREB-H partial deficiency. Based on our results and on the other already published observations, searching for CREB3L3 gene mutations should be part of the primary hypertriglyceridemia genetic work-up.

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Disclosures

None.

References

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Material and methods

Case description and pedigree
The proband is a 49-year-old woman. Her parents are apparently unrelated. Her father
died at the age of 48 because of lung cancer and he had a history of moderate
hypertriglyceridemia (referred up to 500 mg/dl). The mother and one older sister are
reported to be nomolipidemic. Her 51 years old brother has been suffered of sporadic
episodes of abdominal pain and discomfort and in one occasion his triglycerides levels
were as high as 1600 mg/dl. The clinical history of the two 24 and 13 years old daughters is
reported as unremarkable but no data on lipid profile were known.
Severe hypertriglyceridemia (about 1000 mg/dl) was first noted in the occasion of a routine
laboratory check up. A diet restriction to about 15\% of fat intake (<7\% of saturated fat) and
treatment with fibrates and ω-3 fatty acids were prescribed but the adherence to
prescriptions was poor resulting in fluctuating plasma triglycerides levels (referred from
150 mg/dl and 1300 mg/dl).
On December 2013 the patient was referred for the first time to our Lipid Clinic. The
physical examination was unremarkable. Mild fatty liver was detected upon ultrasound
examination.
Routine laboratory tests, with the exception of plasma lipids, were within the reference
range. Plasma triglyceride (TG), total cholesterol (TC) and HDL-C levels were respectively
500 mg/dl (5.64 mmol/l), 218 mg/dl (5.64 mmol/l), and 33 mg/dl (0.77 mmol/l). At the time
of this clinical evaluation the adherence to low-fat diet was poor and she has discontinued
the hypolipidemic drugs.
Based on this clinical history we decided to start a diagnostic genetic work out.
Informed consent was obtained from all subjects investigated.

Plasma lipid analysis
Blood samples from proband and her relatives (10 mL each) were collected into plain and
containing EDTA (1 mg/mL) tubes to obtain serum, plasma and buffy coat by
centrifugation at 3000 rpm for 15 min. Plasma TC, TG and HDL-C were measured using
standard enzymatic–colorimetric procedures (Roc
de, Basel Switzerland) on a
COBAS MIRA plus auto-analyzer (Roche Diagnostics, Basel Switzerland). LDL-C was
calculated by the Friedewald formula.

Analysis of candidate genes
Genomic DNAs from studied subjects were extracted from whole blood using the Wizard
DNA Purification System (Promega Italia, Italy). Candidate genes of recessive (LPL,
APOC2, APOA5, GPIHBP1and LMF1 genes) and dominant (CREB3L3) forms of
hypertriglyceridemia were analyzed.
All exons, the promoter region the intron/exon boundaries of LPL, APOC2, APOA5,
GPIHBP1and LMF1 genes were amplified by PCR from genomic DNA using appropriate
primers as previously described (1-2). Resequencing of the coding regions and consensus
splice sites of APOC3 gene in all investigated subjects was performed using appropriate
primers as previously described (3).
The coding regions including the intron/exon boundaries (reference sequence
NM_032607) of CREB3L3 gene were amplified using the primers listed in Table 1.
PCR fragments were purified with a commercial kit (Wizard PCR Preps–DNA Purification
System; Promega Italia, Italy) then sequenced directly in both directions using BigDye
Terminator Cycle sequencing kit 3.1 in a 3500 Genetic Analyzer (Life Technologies Italia,
Italy) and the results were analyzed with the SeqScape software version 3 (Life
Technologies Italia, Italy).
The proband and other available family members were also genotyped for ε2, ε3, and ε4 polymorphisms in APOE gene by a commercially available preformulated TaqMan assay (Life Technologies) in a ViiA™ 7 Real-Time PCR System (Life Technologies).

References

