Fibroblast Growth Factor-2 Is Required for Vasa Vasorum Plexus Stability in Hypercholesterolemic Mice


Objective—Vasa vasorum are angiogenic in advanced stages of human atherosclerosis and hypercholesterolemic mouse models. Fibroblast growth factor-2 (FGF-2) is the predominant angiogenic growth factor in the adventitia and plaque of hypercholesterolemic low-density lipoprotein receptor–deficient/apolipoprotein B100/100 mice (DKO). FGF-2 seems to play a role in the formation of a distinct vasa vasorum network. This study examined the vasa vasorum structure and its relationship to FGF-2.

Methods and Results—DKO mice treated with saline, antiangiogenic recombinant plasminogen activator inhibitor-123, or soluble FGF receptor 1 were perfused with fluorescein-labeled Lycopersicon esculentum lectin. Confocal images of FGF-2–probed descending aorta adventitia show that angiogenic vasa vasorum form a plexus-like network in saline-treated DKO similar to the FGF-2 pattern of distribution. Mice treated with recombinant plasminogen activator inhibitor-123, or soluble FGF receptor 1 lack a plexus; FGF-2 and vasa vasorum density and area are significantly reduced. A perlecan/FGF-2 complex is critical for plexus stability. Excess plasmin produced in recombinant plasminogen activator inhibitor-123–treated DKO mice degrades perlecan and destabilizes the plexus. Plasmin activity and plaque size measured in DKO and DKO/plasminogen activator inhibitor-123 mice demonstrate that elevated plasmin activity contributes to reduced plaque size.

Conclusion—An FGF-2/perlecan complex is required for vasa vasorum plexus stability. Elevated plasmin activity plays a significant inhibitory role in vasa vasorum plexus and plaque development. (Arterioscler Thromb Vasc Biol. 2012;32:00-00.)

Key Words: adventitia • angiogenesis • atherosclerosis • fibroblast growth factor-2 • vasa vasorum

Vasa vasorum are a network of microvasculature that originate primarily in the adventitia of large arteries. They supply oxygen and nutrients to the outer layers of the arterial wall which are beyond the limit of diffusion from the luminal surface. Vasa vasorum become angiogenic during atherosclerosis in humans and in mouse models of atherosclerosis. It is thought that they may contribute to the atherosclerotic disease process by facilitating plaque growth and serving as a conduit for inflammatory cell infiltration into the plaque, however, there is no direct evidence.

Kwon et al* used micro-computerized tomography to identify 2 types of vasa vasorum in pig coronary arteries, which they named first and second order. The first order originates from large vessels and runs longitudinally between the adventitia and media of the main vessel. The second order consists of small vessels that branch from the first order and form circumferential arches around the vessel wall. Pigs with normal hearts have significantly greater first order vasa vasorum density and mean diameter compared with second order, whereas hypercholesterolemic pigs have more second-order vasa vasorum. The second-order vessels form a dense plexus in the adventitia, which is not found in healthy animals. Others, who used micro-computerized tomography to analyze vasa vasorum branching patterns in nondiseased pigs, show a dichotomous tree structure similar to the vasculature of systemic circulation. However, the branching lacks uniformity in that some continue to branch, whereas others discontinue the process. These data and additional studies indicate that vasa vasorum in healthy pigs behave like end arteries. The combined studies clearly suggest that the adventitial vasa vasorum can acquire an altered branching pattern in response to the disease process.

Fibroblast growth factor-2 (FGF-2) belongs to the FGF family of growth factors. FGFs play a critical role in early embryonic development and a key role in adult neovascularization, inflammation, wound healing and tumor growth. They stimulate their activities primarily through binding interactions with FGF receptor (FGFR) 1 and FGFR2. FGF-2
can have other binding partners, to include syndecan-4,13 perlecan,14,15 betaglycan,16 fibrin, and fibrinogen.17,18 These interactions can influence FGF-2 activity and angiogenesis, protect FGF-2 from degradation, and prolong its effects on endothelial cells (ECs).19

Heparin-binding growth factors have a high affinity for heparan sulfate proteoglycan, particularly perlecan. In the case of FGF-2, the interaction with perlecan increases FGF-2 binding affinity for FGFRs and is required for FGFR phosphorylation and signal transduction.14,20,21

A truncated plasminogen activator inhibitor-1 (PAI-1) isoform, recombinant PAI (rPAI)-1, has significant antiangiogenic activity in vitro.22–24 It inhibits angiogenic vasa vasorum and promotes plaque regression in a low-density lipoprotein receptor-deficient (LDLR−/−) apolipoprotein B(001/001) (Apob(001/001)) (DKO) mouse model of atherosclerosis.4,5 FGF-2 is the predominant angiogenic growth factor associated with adventitial vessels in hypercholesterolemic DKO mice.4 FGF-2 seems to play a role in the formation of a distinct vasa vasorum network which is not present in rPAI-1−/−-treated DKO.4,5 The objective of this study was to further examine the unique vasa vasorum structure and its relationship to FGF-2.

Methods

Mouse Strain, Diet, and Treatment

LDLR−/−Apob(001/001) mice (B6; 129S-Apobtm2Sgy Ldlrtm1Her/J) (DKO) were purchased from Jackson Laboratory (stock 003000). The LDLR−/−Apob(001/001)/PAI−/− mouse strain (DKO/PAI−/−) was produced by crossing B6; 129S-Apobtm2Sgy Ldlrtm1Her/J with B6;129S2-Serpine1tm1Mlg/J (Jackson Laboratory, stock 002507). They were backcrossed for a minimum of 7 generations. Mice were fed either Paigen’s diet (PD) without cholate (Research Diets, New Brunswick, NJ) or normal chow diet (CH) for 20 weeks and then received either rPAI−/− (5.4 µg/kg per day) or saline treatment for the last 6 weeks of the diet, as previously described.1 An adenoviral soluble FGFR1 (sFGFR1) construct (1×109 pfu), produced by Dr Simons laboratory25 (see online-only Data Supplement), was delivered by intraperitoneal injection into DKO mice at 14 weeks of age. Mice were perfused and euthanized 10 days after delivery. (See the online-only Data Supplement), was delivered by intraperitoneal injection into DKO mice at 14 weeks of age. Mice were perfused and euthanized 10 days after delivery. (See the online-only Data Supplement, for details regarding mouse models, rationale for use, treatment regimen, and experimental application.) Animal care and procedures were performed in accordance with the guidelines of the Animal Care and Use Committee and procedures outlined in the Guide for the Care and Use of Laboratory Animals (National Institutes of Health publication No. 86-23, 1985). All procedures were approved by the Dartmouth College Institutional Animal Care and Use Committee.

Perfusion

Mice were injected with 50 µL of heparin, followed by an injection of 0.1 mL ketamine/30 g of weight. They were euthanized and perfused with PBS, followed by 3.5% paraformaldehyde under 110 to 120 mm Hg pressure. Alternatively, mice were perfused as follows: (1) PBS, pH 7.2; (2) PBS containing 1% BSA and 1% fluorescein-labeled Lycopersicon esculentum lectin (fluorescein isothiocyanate [FITC]-lectin) (Vector Laboratories, Burlingame, CA); (3) PBS containing 1% BSA; and (4) PBS containing calcium, magnesium, 1% BSA, 2% glutaraldehyde, and 1% paraformaldehyde.

Confocal Imaging of Vasa Vasorum and FGF-2 in Descending Aorta Whole Mounts

Mice were perfused with FITC-lectin. The descending aorta (DA) to the iliac bifurcation was surgically removed and probed for FGF-2 (Sigma, St. Louis, MO). DA whole mounts were examined by confocal microscopy at ×20 and ×63 magnification, and z-stacks were collected as previously described.4,5 N=21 for DKO treatment groups and 8 for DKO/PAI-1−/− treatment groups.

Quantification of Confocal Images

Confocal z-stack slices were aligned in volumetric images. The resolution of the reconstructed volumetric data was increased by tri-linear interpolation for a detailed geometrical representation in 3-dimension. Microview software (GE Healthcare, Piscataway, NJ) was used to quantify the volume and area of FGF-2 and lectin. Quantitative values were obtained from the program generated isosurface. The 2 images were overlaid. N=8/group. Vessel diameter was measured in reconstructed confocal images using Microview software. N=20/group.

Detection of FGF-2, Perlecan, and CD31 in the Adventitia of Descending Aortas

Thirty micron (µm) frozen sections of the DA from rPAI−/−, or saline treated PD DKO mice were probed for FGF-2, perlecan (Millipore, Billerica, MA) and CD31 (BD Biosciences, San Jose, CA) as previously described.4,5 N=3/group.

Detection of EC Death in the Vasa Vasorum

Propidium iodide (20 mg/kg) (Sigma) was delivered by retro orbital injection in 4 increments during a period of 15 minutes. Mice were perfused and euthanatized 20 minutes after the final injection. N=3/group.

Plasmin Activity Measurement

Plasmin activity, in 165 µg of plasma protein (sodium citrated collected), was measured in a Chromozym PL assay (Roche, Indianapolis, IN), as previously described.7 N=6/group.

Plaque Measurements

Image J software (National Institutes of Health) was used to measure plaque area in sequential 30 µm cross sections of the DA. Plaque volume was calculated by multiplying the plaque length times the average measured area. N=9/group.

Statistical Analysis

Statistical analysis was performed with a 2-tailed indirect Student t test, 1-way ANOVA with a post hoc least significant difference test with or without repeated measures or with a χ2 test, as appropriate, using the SPSS 12.0.1 statistical software package.

Results

Vasa Vasorum Form a Plexus-Like Structure in Hypercholesterolemic Mice

DKO mice, fed CH diet for 12 weeks and thereafter by 20 weeks of PD, were treated with either saline or rPAI-1−/− during the last 6 weeks of PD.4 Mice were perfused with FITC-lectin and confocal z-stack images of DA whole mounts were acquired (Figure 1). Chow-fed mice have detectable adventitial vessels, but they lack a branching pattern (Figure 1A and 1D). The vasa vasorum in PD-fed, saline-treated mice form a distinct, dense network (Figure 1B) of interconnecting vessels (Figure 1E). The plexus-like network appears to have been present in rPAI−/−-treated PD mice (Figure 1C), but smaller vessels have regressed/collapsed (Figure 1F). The data show that the vasa vasorum acquire a different structure in response to the diet-induced disease process and many of the vessels become unstable with rPAI-1−/− treatment.
FGF-2 Forms a Distinct Pattern in the Adventitia

Our previous studies indicate that FGF-2 is the predominant angiogenic growth factor in the DA adventitia of PD-fed mice.4 Further examination of FGF-2 and vascular endothelial growth factor protein expression in PD-fed mice perfused with FITC-lectin show that vascular endothelial growth factor is mostly undetectable in the adventitia and in rare cases is found randomly distributed in the extracellular matrix/basement membrane (Figure II in the online-only Data Supplement). However, FGF-2 is abundantly expressed and forms a distinct pattern in the adventitia of saline-treated PD mice (Figure III in the online-only Data Supplement). The vessel density, diameter, and pattern of distribution are variable within the same adventitia (Figure IIIB in the online-only Data Supplement). Those differences appear to be dependent upon FGF-2 distribution (Figure IIIC in the online-only Data Supplement, right side versus left side).

FGF-2 Is Required for Vasa Vasorum Plexus Network

To determine whether FGF-2 is required for maintenance and stability of the vasa vasorum plexus, DA whole mounts from saline-treated, FITC-lectin perfused mice were compared with rPAI-123– or sFGFR1-treated mice. The rPAI-123 protein causes vasa vasorum collapse through a mechanism that increases plasmin activity; therefore it was used as a means of investigating FGF-2 distribution in relationship to loss of vasa vasorum. However, sFGFR1 is a decoy for FGFs and provides a means of demonstrating how removal of FGF affects vasa vasorum stability. Mice that were CH-fed for 32 weeks were used as controls.

FGF-2 in CH-fed mice is present in a diffuse pattern, lectin perfused vessels are few in number and do not form a network (Figure 2A). FGF-2 distribution is more defined in the T0 group, vessels are expanded and some form a pattern that is parallel to FGF-2 (Figure 2B). The likeness between the FGF-2 pattern of distribution and the vascular network pattern becomes even more distinct by 20 weeks of PD (Figure 2C). FGF-2 is significantly reduced after 6 weeks of rPAI-123 treatment and most vessels collapse/regress. The remaining vessels are aligned with FGF-2 and appear to have a larger diameter (Figure 2D). Similarly, FGF-2 and associated vessels in PD mice treated with sFGFR1 have regressed significantly (Figure 2E). These data demonstrate that FGF-2 is required for the vasa vasorum to maintain a stable plexus-like structure.

Quantitative Validation of Observed Differences Among Treatment Groups

The observed differences in FGF-2 and vasa vasorum among the treatment groups were verified in measurements of FGF-2 and lectin volume (Figure 3A), area (Figure 3B), and vessel diameter (Figure 3D) in confocal z-stack images reconstructed in Microview software. The measurements show that lectin volume is 2-fold greater in the PD-fed saline versus the PD-fed, rPAI-123 treatment group (saline, 0.04±0.007; rPAI-123, 0.02±0.004mm³; P<0.03). FGF-2 volume is 2.4-fold greater in the saline-treated mice (saline, 0.059±0.02; rPAI-123, 0.025±0.01mm³; P=0.05). Similarly, lectin area is 2.3-fold greater in PD-fed saline compared with rPAI-123–treated mice (saline, 4±0.2; rPAI-123, 1.75±0.4mm²; P<0.001). FGF-2 area is 3.3-fold more in the mice treated with saline (saline, 4.9±0.5; rPAI-123, 1.48±0.14mm²; P<0.001). The volume and area measured in sFGFR1-treated mice are comparable to rPAI-123.

Confocal images show that the vasa vasorum in PD-fed DKO, saline-treated mice have a vascular tree with vessel branches of varied diameter (Figure 3C). Measurements of vessel diameter in all treatment groups fall into 3 categories, <5, 5, or >5 micron (µm). Those that comprise the plexus...
are 5 μm, whereas the main vessels from which the plexus branches are >5 μm. The numbers of vessels in the ≤5 or >5 μm ranges are not significantly different among all groups. However, the rPAI-123- and sFGFR1-treated mice have significantly fewer vessels with a 5 μm diameter compared with the T0 and saline-treated groups (T0, 10±2; saline, 10±1; rPAI-123, 5±1; sFGFR1, 4±2; P=0.008). These differences are consistent with the confocal images, which show that smaller vessels are the ones that collapse in response to rPAI-123 and sFGFR1 treatments.

Potential FGF-2 Binding Partners

We considered that FGF-2 may require a binding partner to define its distinct pattern of distribution. Perlecan was a potential partner because it is known to bind FGF-2.26,27 Additionally, rPAI-1 stimulates increase in plasmin and matrix metalloproteinase-3 activities degrades perlecan, fibrinogen, and nidogen,5 all key components of the extracellular matrix/basement membrane. Degradation leads to loss of the supportive scaffold needed for angiogenic vessel stability and results in vessel collapse.3 Cross sections of the DA from saline and rPAI-123-treated, PD-fed DKO mice were probed for perlecan, FGF-2, and EC marker CD31 to visualize the distribution of perlecan and FGF-2 in relation to vessels in the adventitia. Colocalization of perlecan and FGF-2 was detected along with adventitial vessels in the saline-treated group (Figure 4A). However, mice treated with rPAI-123 show complete loss of perlecan, loss of most well-defined vessels and loss of a distinct FGF-2 pattern (Figure 4B). These data indicate that an FGF-2/perlecan complex is necessary for the distinct FGF-2 pattern of distribution. In the absence of the complex, the vessels collapse/regress. Propidium iodide was injected retro orbitally into mice from both treatment groups to determine whether loss of the complex results in EC death. After perfusion, DA whole mounts were probed for CD31. The saline-treated mice have an intact vasa vasorum plexus (Figure 4C), whereas ECs in the vasa vasorum of rPAI-123- treated mice are mostly dead (Figure 4D).

Plasmin Remodels the Angiogenic Vasa Vasorum

The effects of plasmin activity on vasa vasorum structure and stability were further examined by comparing PD-fed DKO with PD-fed DKO/PAI-1−/− mice. DKO/PAI-1−/− mice were selected, because PAI-1 inhibits the conversion of plasminogen to plasmin thus enabling a comparison with elevated plasmin activity in response to rPAI-123.

Reconstructed confocal images of the adventitial vasa vasorum in DA whole mounts show that the saline-treated, PD-fed DKO mice have a structural hierarchy; there is a large main vessel from which smaller vessels branch and they in turn branch to form a plexus (Figure 5B). The vasa vasorum in the DKO, rPAI-123 treatment group have large main vessels, but the plexus is collapsing/regressing (Figure 5C). The adventitial vessels in PD-fed DKO/PAI-1−/− mice treated with saline appear to have been an ordered arterial tree with smaller vessels between the larger branches (Figure 5D). The smaller vessels located between and connecting the main branches of the ordered arterial tree seem to be a plexus undergoing remodeling and regression.
PAI-1−/− mice have large vessels that do not display a vascular degradation/pruning. However, the rPAI-123-treated DKO/PAI-1−/− mice have large vessels that do not display a vascular degradation/pruning. Complete loss of the plexus network results in collapse of the larger vessels. The rPAI-123-treated DKO are able to maintain the structure of the larger branches (Figures 5C and 1C), which suggests that PAI-1 plays an important role in maintaining plexus stability.

**Increased Plasmin Activity Corresponds With Plaque Reduction**

Because plasmin activity has a significant impact on vasa vasorum stability, its effect on plaque growth was analyzed in DKO and DKO/PAI-1−/− mice. Plasmin activity measured (Figure 6A) in rPAI-123–treated DKO mice is >2-fold its saline counterpart (rPAI-123, 0.2±0.1×10−3 units/mL; versus saline, 0.1±0.01×10−3 units/mL; P<0.05). The activity in PD-fed, rPAI-123–treated DKO/PAI-1−/− mice is elevated 6-fold compared with rPAI-123–treated DKO (DKO/PAI-1−/−, rPAI-123, 1.2±0.06×10−3 units/mL; P<0.001). The plaque volume (Figure 6B) in PD-fed DKO, saline-treated mice is >3.8-fold that of the rPAI-1−/−–treated counterpart (DKO, saline, 1.5±2.0 versus DKO, rPAI-1−/−, 0.4±0.2 mm3; P<0.003) and >7.5-fold DKO/PAI-1−/−, rPAI-123–treated mice (0.2±0.1 mm3; P<0.001). These data clearly indicate that elevated plasmin activity has a significant impact on plaque growth (Figure 6C–6F).

**Discussion**

This study shows, for the first time, the detailed structure of the vasa vasorum in murine atherosclerosis. The data further show that the vasa vasorum acquire a plexus-like structure in LDLR−/−/ApoB100/100 mice that are fed a high-fat, high-cholesterol diet; the plexus contains the newly formed microvasculature. Our previous studies show that the vasa vasorum acquire an unusual pattern in hypercholesterolemic, saline-fed LDLR−/−/ApoB100/100 mice and that FGF-2 seems to be associated with the vasa vasorum. This study advances our initial observations by showing that FGF-2 is required for the vasa vasorum to maintain stability of the plexus network. Moreover, we show that an FGF-2/perlecan complex is critical for specifying FGF-2 spatial distribution. Degradation of the larger vessels. The rPAI-123–treated DKO are able to maintain the structure of the larger branches (Figures 5C and 1C), which suggests that PAI-1 plays an important role in maintaining plexus stability.
perlecan destabilizes the complex causing FGF-2 to lose its defined pattern of distribution and the vasa vasorum plexus destabilizes/regresses.

It remains to be determined whether FGF-2 provides a pattern for the vasa vasorum to form a plexus-like network or whether the vasa vasorum form a plexus, produce and release FGF-2 into the matrix. The images of DA whole mounts in chow-fed mice suggest that FGF-2 is present in a similar, but more diffuse pattern than the PD-fed mice. This occurs in the absence of an expanded vasa vasorum, which strongly suggests that FGF-2 provides the pattern for the vasa vasorum plexus. The more diffuse FGF-2 distribution may be due to differences in extracellular matrix/basement membrane composition or reduced matrix remodeling in non-hypercholesterolemic mice.

Clearly, the vasa vasorum expand and begin to form a plexus when the mice are fed a high-fat, high-cholesterol diet for 14 weeks. Our previous studies show that plaque has developed in the aortic arch and DA of DKO mice at 14 weeks of PD.4 The combined data indicate that development of the

Figure 5. Plasmin remodels the angiogenic vasa vasorum. Atherosclerosis mouse model low-density lipoprotein receptor–deficient (LDLR−/−) apolipoprotein B (ApoB)100/100 (DKO) and plasminogen activator inhibitor (PAI)-1 deficient LDLR−/−ApoB100/100/PAI-1−/− (DKO/PAI-1−/−, plasmin in excess) mice were chow diet (CH)-fed for 32 weeks or CH for 12 weeks and thereafter by Paigen's diet (PD) from 12 to 32 weeks of age and treated with saline or recombinant PAI (rPAI)-1−/−, for the last 6 weeks of PD. Mice were perfused with fluorescein isothiocyanate-labeled lectin to detect plasmin effects on adventitial vessel structure. Descending aorta whole mounts were imaged by confocal microscopy at ×20 magnification. (A) DKO, CH; (B) DKO, PD, saline; (C) DKO, PD, rPAI-1−/−; (D) DKO/PAI-1−/−, PD, saline; (E) DKO/PAI-1−/−, PD, rPAI-1−/−. Yellow arrows in Figure 6D and 6E indicate position of plexus. Scale bar=20 μm. Note the extensive plexus in PD-fed, saline-treated DKO that is collapsing in rPAI-1−/−-treated mice. Plasmin in DKO/PAI-1−/− mice from both treatment groups appears to significantly remodel the plexus. N=8/group. Scale=20 μm.

Figure 6. Increased plasmin activity corresponds with plaque reduction. Hypercholesterolemic low-density lipoprotein receptor–deficient (LDLR−/−) apolipoprotein B (ApoB)100/100 (DKO) and plasminogen activator inhibitor (PAI)-1 deficient LDLR−/−ApoB100/100/PAI-1−/− (DKO/PAI-1−/−, plasmin in excess) mice treated with recombinant PAI (rPAI)-1−/− or saline were examined for (A). Plasmin activity measured in plasma using a Chromozym PL assay. (B) Plaque volume measured in sequential descending aorta (DA) cross sections using Image J software. N=9/group. Representative images of plaque in hematoxylin and eosin-stained DA from each group are shown at ×10 magnification: (C) DKO, saline; (D) DKO, rPAI-1−/−; (E) DKO/PAI-1−/−, saline; (F) DKO/PAI-1−/−, rPAI-1−/−. Note that increased plasmin activity corresponds with reduced plaque size.
Vasa vasorum plexus is part of the disease process. This is in keeping with the work of Kwon et al., who described a second-order vasa vasorum plexus in hypercholesterolemic pigs, but not in healthy animals.

FGF-2 has a short in vivo half-life of 2 to 4 minutes. It is highly susceptible to denaturation and degradation; its various binding partners provide FGF-2 with protection from degradation, thus prolonging its effects on ECs. Additionally, heparan sulfate proteoglycans can serve as an FGF-2 reservoir enabling higher than normal local concentrations of FGF-2 that prolong its stimulation of ECs. Because FGF-binding partners are at many-fold higher concentrations than FGF-2, it should be found complexed with a binding partner. The relative concentrations of FGF-2-binding partners may change because of physiological or pathophysiological changes to result in different FGF-2 activity. That is consistent with what we find for FGF-2 in the PD-fed DKO model. Its complex with perlecan is critical for maintaining the defined vasa vasorum structures, as demonstrated in saline-treated DKO mice. Loss of perlecan results in dispersal of FGF-2, collapse of the vessels and EC death, as shown in rPAI-123–treated mice.

Evidence of a perlecan/FGF-2 interaction does not rule out the possibility of other binding partners that may contribute to or alter the effect we observe for FGF-2 in PD-fed DKO mice. Changes in binding partners may occur at various stages of vasa vasorum development during the disease process. Others have reported that when an FGF-2-binding partner changes from a bound to soluble form it changes the activity of FGF-2.

Altogether, this study demonstrates that the branching pattern of the adventitial vasa vasorum in DKO mice is similar to the findings of Kwon and Gossl. We demonstrate that the angiogenic microvascularity (5 μm lumen diameter) form a plexus in hypercholesterolemic mice, but not CH-fed mice, which is consistent with Kwon’s description of the second-order vasa vasorum structure in diseased pig coronary arteries. However, it should be noted that micro-computerized tomography images in Kwon’s studies relate more to arterioles with a lumen diameter >50 μm, whereas the adventitial plexus in DKO mice are at the capillary size. The plexus branches form a vascular tree structure. Degradation or pruning of the plexus in DKO/PAI-1−/− mice reveals a tree-like structure that is in keeping with Gossl’s prediction that the vasa vasorum in nondiseased pigs behave like functional end arteries not connected by a plexus.

It would not be surprising if FGF-2 provides guidance of the vasa vasorum into a plexus, because FGFs control guidance and branching patterns in the Drosophila trachea. Additionally, FGFs are known to serve as guidance cues for axons in Xenopus laevis retinal ganglion cells. It would not be unusual to find FGF-2 associated with the vasa vasorum in athereogenic mice as others have shown that it enhances the coupling of intimal hyperplasia and proliferation of vasa vasorum in injured rat arteries.

Our previous studies identified a novel pathway by which rPAI-1, regulates plasmin activity. The elevated plasmin activity degrades key components of the basement membrane/extracellular matrix that are important for angiogenic vessel stability. In this study we further demonstrate that elevated plasmin activity corresponds with a reduction in plaque size. Elevated plasmin activity could potentially influence plaque growth through regression of the vasa vasorum and by degradation of the plaque matrix that contributes to the disease process. The rPAI-1,–stimulated pathway that increases plasmin activity provides a mechanism for reducing plaque size that does not necessitate lipid lowering.

Altogether the data presented in this study demonstrate that the murine vasa vasorum and its supportive matrix constitute a dynamic environment. The vasa vasorum can readily expand or collapse in response to cues from the vessel wall.

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References


Disclosures

None.


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**LDLR \(^{-/-}\) ApoB\(^{100/100}\) (DKO)**

- Wean-12wks
- Chow diet
- 12 wks start PD
- 26 wks
- 28 wks T\(_0\) start treatment
- perfuse
- sFGFR1
- 32 wks
- perfuse
- saline, rPAI-1\(_{123}\)
- 3/group lectin
- 21/group/lectin
- 15/group/PFA
- 1. H&E
- 2. PI
- 3. perlecan/FGF2

**LDLR \(^{-/-}\) ApoB\(^{100/100}\)/PAI-1\(^{-/-}\) (DKO/ PAI-1\(^{-/-}\))**

- Wean-12wks
- Chow diet
- 12 wks start PD
- 26 wks
- 28 wks T\(_0\) start treatment
- perfuse
- saline, rPAI-1\(_{123}\)
- 8/group/lectin
- 9/group/PFA
- 1. H&E
**Supplemental Figure II.** LDLR^{-/-}/ApoB^{100/100} mice (DKO) were fed Paigen's diet without cholate (PD) for 20 weeks. Mice were perfused with FITC-labeled lectin (green). Descending aorta whole mounts were probed for VEGF-A (red) and imaged by confocal microscopy at 40x magnification. Scale bar = 10 μm
Supplemental Figure III. FGF-2 forms a distinct pattern in the adventitia of hypercholesterolemic mice. DKO mice were fed PD for 14 weeks then perfused with FITC-labeled lectin. Whole mounted descending aortas were probed for FGF-2 and imaged by confocal microscopy. (A) FGF-2 probed adventitia; (B) FITC-lectin perfused vessels; (C) Overlap of FGF-2 and lectin. Red vertical line separates right from left side. Note the differences in FGF-2 and vessel patterns of distribution between right vs. left. Scale bar = 10 um
SUPPLEMENT MATERIAL

MOUSE MODELS

1. **B6; 129S-Apob<sup>tm2Sgy</sup> Ldlr<sup>tm1Her</sup>/J strain**
   B6; 129S-Apob<sup>tm2Sgy</sup> Ldlr<sup>tm1Her</sup>/J mouse strain (referred to as DKO)(Jackson Laboratories) was selected, because it is reported to more closely mimic human atherosclerosis than other models. This strain has a mutation in the ApoB48 gene; thus the mouse model expresses only apolipoprotein B-100 (ApoB-100) and is deficient in low density lipoprotein receptor (LDLR). The LDLR plays an important role in cholesterol homeostasis by mediating the clearance of LDL and IDL from plasma through binding interactions with ApoB and ApoE. ApoB undergoes post translational editing to form ApoB-100 and ApoB-48.

2. **B6; 129S-Apob<sup>tm2Sgy</sup> Ldlr<sup>tm1Her</sup>/J x B6.129S2-Serpine<sup>tm1Mlg</sup>/J**
   The B6; 129S-Apob<sup>tm2Sgy</sup> Ldlr<sup>tm1Her</sup>/J mouse strain (DKO) was crossed with B6.129S2-Serpine<sup>tm1Mlg</sup>/J (PAI-1<sup>−/−</sup>) to obtain DKO/PAI-1<sup>−/−</sup>. The PAI-1<sup>−/−</sup> mouse was selected, because PAI-1<sup>−/−</sup> binds tPA to prevent tPA conversion of plasminogen to plasmin, therefore plasmin activity is elevated in PAI-1<sup>−/−</sup> mice.

ADENO SOLUBLE FGFR1 CONSTRUCT

The Adeno-soluble FGFR1 construct was produced in Dr. Simons’ laboratory as described by M. Murakami, et al. The details, as reported in the J. Clinical Investigation, are as follows:

The mouse FGFR1 (flg) A341-733 cDNA clone was a gift from Dr. Mansukhani (New York University). The truncated form of FGFR1 lacking the cytoplasmic part of the receptor (FGFR1DN) encodes the entire extracellular two Ig domains with 32 amino acids of the intracellular region. To design sFGFR constructs, extracellular parts of mouse FGFR1DN cDNA (FGFR1IIC isoform), mouse FGFR3IIB and mouse FGFR3IIC cDNA (gift from Dr. D. Ornitz, Washington University School of Medicine) were amplified by PCR with 5' HindIII and 3' BamHI sites, respectively. After restriction enzyme digestion, PCR products were fused in frame to the hinge-CH₂-CH₃ region of human IgG1 (CDM7B/CD5-Ig expression vector). FGFR1DN and sFGFR cDNAs were subcloned into an adenovirus shuttle vector, and nonreplicative, recombinant adenovirus vectors were generated and propagated to high titer.

ATHEROGENIC DIET

Paigen’s atherogenic diet without cholate (PD) contains 12.5 grams cholesterol, 75 grams cocoa butter and 35 grams coconut oil, which equates to 20, 45 and 35 kcal% protein, carbohydrate and fat, respectively.

