Focal Adhesion Kinase Regulates Smooth Muscle Cell Recruitment to the Developing Vasculature


Objective—The investment of newly formed endothelial cell tubes with differentiated smooth muscle cells (SMC) is critical for appropriate vessel formation, but the underlying mechanisms remain unknown. We previously showed that depletion of focal adhesion kinase (FAK) in the nkx2.5 expression domain led to aberrant outflow tract (OFT) morphogenesis and strove herein to determine the cell types and mechanisms involved.

Methods and Results—We crossed fakloxP targeted mice with available Cre drivers to deplete FAK in OFT SMC (FAKWT and FAKΔ) or coronary SMC (FAKcSMC). In each case, depletion of FAK led to defective vasculogenesis that was incompatible with postnatal life. Immunohistochemical analysis of the mutant vascular structures revealed that FAK was not required for progenitor cell proliferation, survival, or differentiation into SMC but was necessary for subsequent SMC recruitment to developing vasculature. Using a novel FAK-null SMC culture model, we found that depletion of FAK did not influence SMC growth or survival, but blocked directional SMC motility and invasion toward the potent endothelial-derived chemokine, PDGFB. FAK depletion resulted in unstable lamellipodial protrusions due to defective spatial-temporal activation of the small GTPase, Rac-1, and lack of Rac1-dependent recruitment of cortactin (an actin stabilizing protein) to the leading edge. Moreover, FAK null SMC exhibited a significant reduction in platelet-derived growth factor-stimulated extracellular matrix degradation.

Conclusion—FAK drives PDGFB-stimulated SMC chemotaxis/invasion and is essential for SMC to appropriately populate the aorticopulmonary septum and the coronary vascular plexus. (Arterioscler Thromb Vasc Biol. 2011;31:00-00.)

Key Words: biology developmental ■ extracellular matrix ■ morphogenesis ■ vascular biology

The investment of newly formed endothelial cell tubes with differentiated smooth muscle cells (SMC) is a very important process during vessel formation and requires intricate regulation of SMC specification, motility, growth, and differentiation. Failure of SMC recruitment to and migration along developing vessels can lead to vascular instability and regression, an event that is likely due in part to the ability of these cells to secrete and organize extracellular matrix-containing basement membranes and elastin. Moreover, mature medial SMC express high levels of contractile genes (i.e., myosin heavy chain, α-smooth muscle actin (α-SMA), SM22 among others) and their presence is essential for maintaining vessel tone and for providing dynamic control of blood pressure (see review).

Although the precise signaling pathways have yet to be identified, a number of major extrinsic factors have been shown to regulate SMC recruitment and function. TGF-β, which promotes SMC differentiation,3–4 and platelet-derived growth factor (PDGF), which promotes SMC growth and motility, are both important extrinsic regulators of SMC phenotype and genetic ablation of receptors for these genes resulted in defective outflow tract (OFT) or coronary vascular morphogenesis.5–7 Extensive evidence indicates that extracellular matrix signaling is also an important regulator of SMC growth and differentiation as deletion of either fibronectin, the α5 integrin FN receptor, or focal adhesion kinase (FAK) (the kinase that mediates α5-dependent signaling) each results in extraembryonic and embryonic vessel defects leading to lethality in the mouse from E8.5 to E10.8–10

Although a direct role for FAK in vascular smooth muscle growth and development has yet to be examined, our laboratory recently showed that depletion of FAK from nkx2.5-expressing precursors led to perinatal lethality resulting from a profound subaortic ventricular septal defect accompanied (in some neonates) by malalignment of the OFT including double-outlet right ventricle and persistent truncus arteriosus.
Figure 1. Focal adhesion kinase (FAK) deletion impairs recruitment of cardiac neural crest-derived smooth muscle cells (SMC) to the developing aorta and pulmonary artery. A, H&E staining of genetic control and FAK<sup>−−</sup> mice at postnatal day 0 (P0). FAK<sup>−−</sup> outflow tract (OFT) shows persistent truncus arteriosus (PTA) and ventricular septal defect (VSD). LA indicates left atrium; RA, right atrium; Ao, aorta; PT, pulmonary trunk; IVS, intraventricular septum. B, C, SM-22 immunostaining (brown) of embryonic day 14 (E14; B) or P0 (C) genetic control and FAK<sup>−−</sup> OFT revealed PTA without defect in SMC differentiation. D, Proximal OFT of genetic control and FAK<sup>−−</sup> hearts at E 12 reveals colocalization of α-SMA (red) with the cardiac neural crest lineage marker CRABP1 (green), indicating that the SMC covering the aorta and pulmonary artery were derived from cardiac neural crest cells. X-gal staining demarcates wnt-1–derived cardiac neural crest in the Rosa26R<sup>Rosa26</sup>–positive embryos. Nuclei (blue) were stained with DAPI. Note the lack of SMC coverage
(PTA).

Thus, conditional inactivation of FAK in nkx2.5-expressing cells phenocopies the most common congenital heart disease that is also a subset of abnormalities associated with Tetralogy of Fallot and the DiGeorge Syndrome.

However, whether these defects arose from the inability of nkx2.5 precursor cells to differentiate into SMC or from a specific defect in nkx2.5-derived SMC remained unclear.

The aim of the present study was to identify the critical FAK-dependent functions during vasculogenesis. Because both wnt-1- and nkx2.5-expressing cells contribute to development of conotruncal septal SMC that divide the OFT into the aorta and pulmonary artery,

we explored the effects of FAK deletion in both origins using available Cre lines. We show FAK deletion (by homologous recombination) in either embryological origin results in OFT defects including PTA. Moreover, targeted deletion of FAK in these SMC precursors did not affect progenitor cell proliferation, survival, or differentiation but resulted in lack of appropriate SMC coverage of the developing vasculature. Similarly, deletion of FAK in a third SMC precursor origin, the wnt1-expressing epicardial cells that provide SMC that line coronary vessels, did not affect SMC specification but led to aberrant recruitment of SMC to the developing coronary vasculature. Collectively, these studies indicate that FAK is essential for a SMC autonomous function during vascular remodeling. Our mechanistic studies in FAK null SMC, revealed that FAK does not influence SMC growth or survival but functions to promote directional SMC chemotaxis/invasion toward the potent endothelial-derived chemokine, PDGF.

**Methods**

### Experimental Animals

fak<sup>−/−</sup> mice were bred to existing Cre lines including cardiac neural crest (CNC)-specific wnt-1<sup>Cre</sup> mice obtained from Andrew McMahon; primary and secondary heart field-specific nkx2.5<sup>Cre</sup> mice obtained from Robert Schwartz; and epicardial-specific wnt1<sup>Cre</sup> mice obtained from John Burch. The resulting fak<sup>−/−</sup>/Cre<sup>−/−</sup> mice were bred with fak<sup>lox/lox</sup> Rosa26<sup>lacZ/wt</sup> mice to obtain fak<sup>lox/lox</sup>/Cre<sup>−/−</sup>/Rosa26<sup>lacZ/wt</sup> (experimental) or genetic control mice (fak<sup>lox/lox</sup>/Cre<sup>−/−</sup>/Rosa26<sup>lacZ/wt</sup> without Cre and fak<sup>lox/lox</sup>/wt <sup>−/−</sup> control). Mice were housed in the University of North Carolina Animal Care Facility, which is accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International, and all experimental procedures were approved by the University of North Carolina Animal Care and Use Committee.

### Statistical Analysis

All quantitative data represent at least 3 separate experiments and are presented as mean±SEM. Means were compared by 2-tailed Student t test. P<0.05 was considered statistically significant.


### Results

FAK Deletion Impairs Recruitment of nkx2.5- and wnt-1-Derived SMC to the Developing Aorta and Pulmonary Artery

We previously showed that conditional deletion of FAK in nkx2.5-expressing cells (FAK<sup>−/−</sup>) resulted in malalignment of the OFT and incomplete aortopulmonary septum formation in a percentage of neonates. Because nkx2.5-derived SMC contribute to the formation of the aortic root and septum, we hypothesized that this phenotype may have resulted from a SMC defect in FAK-depleted cells. To further examine FAK’s role in OFT development, we used the wnt-1<sup>Cre</sup> line to deplete FAK from the CNC-derived SMC that also make a major contribution to this process. These well characterized mice generate high-efficiency recombination around E9.5 in the CNC-derived cells populating the truncus arteriosus and pharyngeal arch arteries (3, 4, and 6) and in the CNC-derived cells invading the conotruncal cushions and the aortopulmonary septum by E11.5. Although fak<sup>lox/−</sup>/wnt-1<sup>Cre/+</sup> Rosa26<sup>lacZ/wt</sup> mice (herein referred to as FAK<sup>wt</sup>) were present at the appropriate Mendelian ratio at P0 (Supplemental Table I, available at http://atvb.ahajournals.org), all FAK<sup>wt</sup> mice died within a few days after birth. Gross morphology of the cardiac OFT revealed aberrant septation and branching of the major outflow vessels in all FAK<sup>wt</sup> mice with the majority (95%; N=42/44) exhibiting PTA (Figure 1A). Thus, our data indicate that FAK activity in nkx2.5- and wnt-1-derived cells is required for appropriate septation of the truncus arteriosus.

Because the aortic arch defects observed in CNC-targeted Notch<sup>−/−</sup> mice were associated with impaired SMC differentiation within the truncus, we tested whether FAK depletion affected this process in our models. Expression of α-SMA in the pharyngeal arch region at E11 was similar between control and FAK<sup>wt</sup> mice (Supplemental Figure IA, available at http://atvb.ahajournals.org). The unseptated ascending OFT in FAK<sup>wt</sup> mice contained layers of SM-22-positive medial SMC that were comparable to littermate controls from E14 onwards (Figure 1B and 1C). SMC layers and differentiation in the carotid arteries also appeared to be normal in FAK<sup>wt</sup> mice (Supplemental Figure IB). These mice were bred to contain the Rosa26<sup>lacZ</sup> allele to track progenitors of the Cre-expressing cells, and the neural crest origin of carotids was confirmed by β-galactosidase positive staining (Figure S1C). In addition, as shown in Supplemental Figure ID, SMC investment of coronary SMC (that were not targeted for FAK deletion) was also similar between genetic control and FAK<sup>wt</sup> hearts, indicating that global changes in flow or hemodynamic properties do not account for the SMC investment phenotypes observed in the FAK<sup>wt</sup> OFTs.

Aortopulmonary septation occurs between E11.5 and E12.5 when a wedge of CNC-derived SMC partitions the aortic sac into distinct aortic and pulmonary channels. To more closely evaluate this process in FAK<sup>wt</sup> mice, serial sections were costained for α-SMA and CNC lineage markers (CRABP1 or expression of the Rosa26<sup>lacZ</sup> allele). As shown in Figure 1D, the SMC lining the pulmonary artery and aorta at the level of the proximal OFT are mainly derived from CNC in control and FAK<sup>wt</sup> mice. However, a selective lack of CNC (and CNC-derived SMC) was apparent in the aortic wall juxtaposed to pulmonary artery in the FAK<sup>wt</sup> mice (Figure 1D; highlighted in box). FAK deficient CNC.
cells populated the conotruncal region dorsal to the PTA and properly differentiated into SMC (Figure 1E). However, in FAK\textsuperscript{wnt} mice the SMC in the dorsal conus were more centrally located (Figure 1E; arrow) and did not surround the developing vessels as completely in control mice.

We previously showed that FAK deficiency did not alter proliferation or survival within the affected OFTs of FAK\textsuperscript{nk} mice.\textsuperscript{11} Similarly, there was no significant difference in levels of cell death in the OFT of E13.5 FAK\textsuperscript{wnt} mice in comparison to controls (27.8 ± 4.3 versus 29.3 ± 7.4 per 0.5 mm\textsuperscript{2}, respectively; N = 5/5). The numbers of BrdU-labeled cells in the affected OFT region of FAK\textsuperscript{wnt} embryos also did not differ significantly from those in the genetic controls (45.8 ± 3.7 versus 43.6 ± 4.8 per 0.5 mm\textsuperscript{2}, respectively; N = 4/4). In support of a lack of major effect on the proliferation and survival of CNC-derived cells, the more distal CNC-derived structures such as the aforementioned carotid arteries (Supplemental Figure IB and IC) and other neural crest-derived structures including the thymic lobes, thyroid, tongue, and ear appeared to be unaffected by FAK deletion (data not shown).

Close evaluation of the cranial cartilage and bones did reveal a modest mandibular truncation in 3/6 FAK\textsuperscript{wnt} neonates, but the crown-rump length and maxilla formation were normal (Supplemental Figure IE). Collectively, these studies reveal that CNC functions, population of the developing OFT, and differentiation of \( wnt-1 \) lineages into SMC occur normally in FAK\textsuperscript{wnt} mutants. However, in these mice and those in which FAK was depleted in \( n k x 2.5 \)-derived cells, FAK null SMC fail to appropriately invest the inner walls of the developing aorta and pulmonary trunk, indicating that FAK may be necessary for directional SMC movement within the conotruncus.

**FAK Deletion Impairs Recruitment of Epicardial-Derived SMC to the Coronary Plexus**

To provide additional in vivo evidence for our hypothesis, we tested the effects of FAK deletion on the recruitment of epicardial-derived SMC to the intramyocardial endothelial plexus, a process also dependent on directional motility.\textsuperscript{7} To this end, we intercrossed our FAK\textsuperscript{lox/lox}/\textit{Rosa26R\textsuperscript{LacZ/LacZ}} mice to an available \textit{Wilm’s Tumor-1} \textit{Cre} line (\textit{wt-1}, hereafter referred to as FAK\textsuperscript{cSMC} so as to not be confused with FAK\textsuperscript{wnt}) that induces strong and uniform recombination in the proepicardium by E9.5 and is restricted to the epicardium from E10.5 onward.\textsuperscript{17,20} Previous studies revealed that \textit{wt1}-derived cells give rise to the SMC that invest the coronary arteries and veins,\textsuperscript{7,20} and we found similar results on analysis of \( \beta\)-galactosidase expression in P0 \textit{wt1\textsuperscript{Cre/Rosa26RLacZ}} hearts (Supplemental Figure II, available at http://atvb.ahajournals.org). We observed Mendelian distribution of FAK\textsuperscript{cSMC} at P0 (Supplemental Table I), but all FAK\textsuperscript{cSMC} mice died within a few days after birth by still to be determined mechanisms.

Importantly, immunohistochemical assessment of \( \alpha\)-SM\A positive cells in P0 hearts, revealed a striking and consistent lack of SMC coverage of the coronary endothelial plexus (Figure 2). All large diameter vessels in FAK\textsuperscript{cSMC} hearts exhibited a thin noncontinuous layer of SMC in comparison to comparable genetic control vessels (Figure 2B, arrowheads), although most of the smaller arterioles exhibited a complete lack of associated SMC (Figure 2B, arrows).
Quantification of SMC numbers revealed significantly fewer SMC associated with both arteries and arterioles in FAKcSMC hearts relative to genetic controls (Figure 2C,D). In contrast, significantly higher numbers of α-SMA positive cells were observed in the subepicardial space in FAKcSMC, compared to genetic control hearts (Figure 2A, arrows, Figure 2E), indicating that the FAK null cells efficiently delaminated from the epicardium and differentiated into SMC, but failed to migrate into the myocardium. Importantly, a reduction in platelet/endothelial cell adhesion molecule labeled coronary vasculature was not observed in the FAKcSMC hearts (Figure 3), indicating appropriate formation of the primary plexus. This conclusion is supported by the finding that these hearts did not exhibit signs of hypoplastic growth or noncompaction that are associated with defects in endothelial plexus function (Supplemental Figure IIIA and IIIB, available at http://atvb.ahajournals.org).21 As well, no defects in the formation or SMC investment in carotid arteries (not targeted for FAK depletion) were observed in the FAKcSMC neonates, indicating a lack of effect on global hemodynamics (Supplemental Figure IIIC).

We next performed additional studies to confirm that FAK activity was not required for the proliferation or survival of SMC progenitors, or their differentiation into coronary SMC. As shown in Supplemental Figure IVB (available at http://atvb.ahajournals.org), there was no statistical difference in the rates of cell proliferation or apoptosis in β-gal labeled wt-1-derived cells within FAKcSMC hearts compared to genetic controls at either E15.5 or P0. Interestingly, by E15.5, many β-gal labeled cells in genetic control hearts had moved into the subepicardial zone, whereas most of the β-gal labeled cells in E15.5 FAKcSMC hearts remained associated with the epicardium (Supplemental Figure IV/A), corroborating our previous findings in the P0 hearts that suggested impaired directional motility of the FAK-depleted SMC (Figure 2B and 2E). We also analyzed SMC differentiation marker gene expression in quail proepicardial organ explants treated with adenoviruses that express GFP or the FAK inhibitor, GFP-FRNK. As shown in Supplemental Figure V (available at http://atvb.ahajournals.org), expression of FRNK attenuated the outward movement of the epicardial explants but did not affect the induction of SMC marker gene expression in these cultures. These studies strongly suggest that the SMC investment defect observed in FAKcSMC coronary vessels did not result from a block in the proliferation, survival, or differentiation of wt-1 progenitors but rather from a lack of SMC recruitment to the endothelial plexus.

FAK Deletion Impairs SMC Chemotaxis but not Proliferation or Survival

To confirm that FAK plays a significant role in the regulation of SMC chemotaxis, we established a conditional FAK null SMC culture model using cells isolated from faklox/flox mice. As shown in Figure 4A, treatment of faklox/flox SMC with Cre adenovirus (but not LacZ) resulted in a significant reduction of FAK protein. In excellent agreement with our in vivo studies, FAK deletion had no effect on SMC differentiation marker gene expression (Figure 4A) and, importantly, did not alter the expression of FRNK or the FAK homologue, Pyk2 (Supplemental Figure VIA, available at http://atvb.ahajourn...
FAK Increases Lamellipodial Stability Through Rac1-Dependent Recruitment of Cortactin

Because PDGF-dependent motility involves rapid remodeling of the plasma membrane and underlying actin cytoskeleton, we next explored whether FAK-depleted SMC exhibited a biomechanical defect in one of these processes. To this end, we performed time-lapse imaging of Cre- and LacZ-treated fak<sup>flox/flox</sup> SMC following PDGF treatment. In control cells, PDGF stimulated extensive membrane ruffling as early as 2.5 minutes that culminated in a formation of a dominant, stable leading edge lamellipodium by 15 to 20 minutes (Figure 5A, left). In contrast, PDGF treated FAK null SMC remained unpolarized and rarely formed a distinctive leading edge (Figure 5A, right). Kymographic analysis of lamellipodia dynamics indicated that this difference was mainly due to decreased lamellipodia persistence in FAK null SMC (Figure 5B-5D) and that this defect culminated in a significant decrease in leading edge displacement (Figure 5E) and cell speed in 2-D (Figure 5F).

PDGF-stimulated motility is dependent on Rac1, a small GTPase that controls cell protrusion and lamellipodial dynamics. To test whether FAK is necessary for Rac1 activation, we first analyzed total Rac1 activation in response to PDGF using the standard GST-PBD pull-down assay. As shown in Supplemental Figure VIG, PDGF robustly and transiently activated Rac1 with peak activity by 2.5 minutes and, somewhat surprisingly, this response was not affected by depletion of FAK. Thus, global activation of Rac1 is FAK-independent. Because recent studies indicate that PDGF induces leading edge localized activation of Rac1 via induction of new integrin-extracellular matrix connections, we explored the idea that FAK might be necessary for restricting localized Rac1 activity to the leading edge. Immunofluorescent staining with a Rac1 antibody revealed that PDGF treatment of wildtype and FAK-deficient SMC led to rapid and equivalent accumulation of Rac1 at dorsal ruffles that peaked at 2.5 minutes (Figure 5G top, 5H). By 15 minutes following PDGF-treatment, restricted enrichment of Rac1 at the leading edge was observed in wildtype SMC, but this response was perturbed in FAK null SMC, which exhibited random distribution of membrane-associated Rac1 (Figure 5G bottom, 5I).

To confirm a role for FAK in localized Rac1 activation, we next examined the spatiotemporal distribution of cortactin, an actin binding protein that is recruited to the cell periphery in a Rac1-dependent fashion. Immunofluorescent staining showed that recruitment of cortactin to leading edge-lamellipodia (Figure 5J bottom, 5L), but not dorsal ruffles (Figure 5J top, 5K) was significantly reduced in FAK null SMC, indicating that FAK is critical for this aspect of PDGF-stimulated cytoskeletal remodeling. These data are consistent with the finding that active FAK accumulates at the leading edge of cells following PDGF treatment (Supplemental Figure VIC). Importantly, either blocking Rac1 activity with the pharmacological inhibitor NSC23766 (10 μmol/L, Calbiochem) or treatment with cortactin siRNAs mirrored the effects of depleting FAK on PDGF-stimulated SMC membrane dynamics and chemotaxis (Supplemental Figure VII, available at http://atvb.ahajournals.org). In sum,
these studies indicate that FAK plays a critical role in coordinating and stabilizing the protein complexes that are required for Rac1-dependent leading edge protrusions, and hence, directional SMC motility toward the potent endothelial-derived chemokine, PDGF.

Figure 6. Focal adhesion kinase (FAK) is necessary for smooth muscle cell (SMC)-mediated extracellular matrix degradation. A. LacZ- or Cre-infected fak^flx/flx smc were plated on Oregon Green 488 gelatin/ fibronectin matrix in serum-free medium and treated with vehicle or platelet-derived growth factor (PDGF-BB) for 90 minutes. Cells were fixed and costained with anti-FAK antibody and phalloidin. B. Cells were scored for the presence of degradation puncta (black spots) and data are presented as puncta per cell normalized to values for the LacZ-infected SMC following PDGF treatment. Data represent mean±SEM of at least 200 cells from 3 independent experiments. P<0.05. C. LacZ- or Cre-infected fak^flx/flx smc were plated on DQ-gelatin-coated 96-well plate and treated with PDGF-BB for 90 minutes. Fluorescence intensity was monitored at Ex/Em 495/515 nm. Data represent mean±SEM of 3 independent experiments. P<0.05. D. GFP and LacZ or Cre coinfected fak^flx/flx smc were cells treated as above were plated on matrigel-coated inserts (10 µg/mL; Bio-Cat) in serum-free media using either PDGF-BB (20 ng/mL) as the chemoattractant. Invading cells were counted by direct fluorescence at 10x magnification. Data represent mean±SEM of 3 independent experiments. **P<0.001.

FAK Is Necessary for SMC-Mediated Extracellular Matrix Degradation

SMC are known to have a high basal invasive potential, a function mediated by podosomes, sites of dynamic actin polymerization activity thought to be the topological equivalent of lamellipodia formed during 3D movement. Interestingly, podosomes are initiated at the inner-face of focal adhesion complexes and require cortactin and its associated Arp 2/3 complex for extension into the underlying matrix and for the recruitment and secretion of matrix-degrading proteases. To determine whether the chemotactic defect of FAK null SMC in the Boyden chamber assays resulted from impaired directional movement alone or to a combination of impaired motility and invasion, we assessed the capacity of wt and FAK null SMC to degrade extracellular matrix. To this end, we cultured LacZ- or Cre adenovirus pretreated fak^flx/flx smc on fluorescent matrix (Oregon green 488 gelatin/fibronectin mixture) and treated them with or without PDGF for 90 minutes. As shown in Figure 6A, PDGF induced focal areas of matrix degradation (cell associated dark spots) in wt SMC, but this response was significantly

Figure 5. Focal adhesion kinase (FAK) regulates Rac1-dependent cortactin recruitment and lamellipodial stability. LacZ- or Cre-infected fak^flx/flx smc were treated with 20 ng/mL platelet-derived growth factor (PDGF)-BB and immediately imaged at 5-second intervals for 60 minutes. A. Representative images of cells 20 minutes following PDGF treatment. Cell polarization as shown by formation of leading edge (arrowhead) and trailing edge (arrow) was observed in control but not in FAK-deficient smooth muscle cells. B. Representative kymographs of LacZ and Cre-infected cells 0 to 20 minutes following PDGF-BB treatment. Ascending slopes represent protrusion (arrows) and descending slopes represent retraction (arrowheads). C–E. Kymographic analysis of lamellipodial dynamics following PDGF treatment. Calculated protrusion rate (D), lamellipodial persistence (E), and leading edge displacement (F) from 8 to 10 cells from 4 independent experiments (mean±SEM; *P<0.05). F. Average 2D cell speed within 4 hours following PDGF treatment. Results are mean±SEM of 30 to 60 cells from 4 independent experiments. *P<0.05. G–I. LacZ or Cre infected fak^flx/flx cells stained for Rac1 showing dorsal ruffle localization in both control (G, top left) and FAK-deficient (G, top right) cells at 2.5 minutes following PDGF treatment. At 15 minutes, leading edge localization of Rac1 (arrows) was observed in control cells, whereas FAK-deficient cells showed unorganized Rac1 localization (arrowheads). H, I. Quantification of cells exhibiting Rac1-labeled dorsal ruffles 2.5 minutes post-PDGF treatment (H) and Rac1-labeled dominant leading edge lamellipodia 15 minutes post-PDGF treatment (I). Results are mean±SEM of 150 to 200 cells from 3 independent experiments. ***P<0.001. J–L. Smooth muscle cells treated as above were stained with anti-cortactin antibodies and processed as described in G–I. Similar dorsal ruffle but not leading edge recruitment of cortactin was observed in FAK-deficient cells. Results are mean±SEM of 200 to 250 cells from 4 independent experiments. **P<0.01. Scale bar=10 µm.
diminished in FAK-deficient SMC. We next evaluated the ability of FAK-containing and FAK null SMC to degrade DQ-gelatin, which fluoresces on degradation allowing quantification of protease activity via fluorescent spectrophotometry. As shown in Figure 6C, significantly lower levels of gelatin degradation occurred in FAK-depleted SMC compared to control SMC, confirming that SMC matrix protease activity is FAK-dependent. In addition, we found that invasion of FAK null SMC into matrigel was significantly impaired in comparison to wt SMC (Figure 6D). These studies indicate that FAK is necessary for both PDGF-stimulated chemotaxis and invasive cell migration; and may explain, at least in part, the selective requirement for FAK in coronary and OFT morphogenesis, as these processes each require SMC to cross tissue/extracellular matrix boundaries.

Discussion

Our studies using 3 independent Cre lines that induce recombination in SM precursor cells revealed a SMC autonomous role for FAK in vascular development. Depletion of FAK in wnt1- and nks2.5-derived cells (ie, FAK


observed normal migration and growth of CNC in FAK


embryos as well as normal differentiation of neural crest cells into SMC within the aorticopulmonary septum. These findings coupled with our mechanistic studies in FAK null SMC suggest that precise regulation of SMC phenotype within the walls of the aortic sac is necessary for subsequent formation of the aorticopulmonary septum. Because PDGF and now FAK have been implicated in the promotion of SMC chemotaxis, it is tempting to speculate that the final septation event may be due to coordinated movement of CNC-derived SMC to the conus. Mis-localization of these cells could therefore lead to impaired coverage of the OFT vessels and impaired force generation within the conus that could impact the dynamic morphogenetic movements during septogenesis.

Because formation of the OFT is extremely complex and involves multiple cell types with interconnected functions, we chose a separate in vivo model (coronary vasculogenesis) to confirm a role for FAK in SMC chemotaxis. The epicardium contains progenitor cells, the major source of SMC that will eventually line the coronary vessels. Elegant lineage tracing studies have shown that SMC precursors are specified in the proepicardium before spreading of this epithelial tissue around the heart.35–37 In response to signals from the myocardium, these specified cells undergo an epithelial to mesenchyme transition, delaminate from the epicardium, and move into the subepicardial space. Epicardial-derived mesenchymal cells will eventually be induced to migrate into the myocardium and to differentiate into definitive cell types; predominantly the SMC that cover the primary endothelial plexus but also some endothelial and myocardial cells.20 The endothelial primary plexus originates from the atrial-ventricular groove at the base of the heart and gradually extends to the apex between E11.5-E13.5 in response to prior wave-like secretion of trophic factors from the myocardium (VEGFA and B) and epicardium (sonic hedgehog)21, an event that is necessary to support midgestational growth. Because the investment of the coronary plexus with mature SMC occurs in a similar (albeit delayed) spatial-temporal pattern, it has been postulated that endothelial-derived factors initiate the recruitment of epicardial-derived SMC. Although the spatial-temporal coupling of epicardial-derived mesenchymal cell differentiation and migration has not been well characterized, the possibility that SMC differentiation may occur before chemotaxis/invasion toward the endothelial plexus is supported by the recent finding that cardiac-restricted depletion of the chemoattractant, thymosin β-4 resulted in robust α-SMA stained cells that aberrantly lined the epicardium (at E14.5 in mouse) but failed to invade the myocardium.38

The following findings from our studies support the hypothesis that activation of FAK in SMC by endothelial-derived signals regulates coronary SMC recruitment to (and along) the developing arterioles: (1) Defects were observed in SMC coverage of arterioles in FAK


hearts but not in the formation or function of the endothelial plexus (as assessed by platelet/endothelial cell adhesion molecule staining and appropriate midgestational heart growth); (2) no significant difference was observed in either the rates of proliferation or apoptosis in FAK-null precursor cells, and (3) significantly higher numbers of α-SMA-positive cells were apparent in the
subepicardial space (as opposed to along the endothelial plexus) of the conditional mutants at P0, indicating that FAK was not essential for epicardial-mesenchymal transition but was necessary for movement into the myocardium. These findings coupled with our in vitro evidence that FAK is necessary for chemotaxis/invasion toward the major endothelial-derived chemo-attractant, PDGFβBB, indicate that lack of directional motility is likely causal for the SMC investment phenotypes observed in the SMC-selective FAK knock-outs. However, it is formally possible that defects in other parameters not evaluated herein could contribute to the defects observed.

Our mechanistic studies revealed that depletion of FAK markedly impaired PDGF-stimulated formation of a stable leading edge lamellipodium, the hallmark of polarized movement. This defect was accompanied by reduced leading edge recruitment of cortactin, a branch filament stabilizer that is essential for lamellipodial persistence and polarized motility. Moreover, cortactin knock-down was shown to affect lamellipodial activity and directional chemotaxis in exactly the same way as depletion of FAK, suggesting that the functions of these two proteins are interrelated. Interestingly, we found that formation of circular dorsal ruffles in response to PDGF does not require Rac1 activation, cortactin, or FAK. Although the function of these transient dorsal ruffles is not completely understood, there is considerable support for the idea that they are formed as a prelude to leading edge lamella and are important for recruitment and recycling of membrane and actin polymerizing/depolymerizing agents to the presumptive leading edge. Indeed, dorsal ruffles contain many of the same actin regulating components as leading edge lamella including (among others) ARP2/3, WASP1/2, WAVE, dynamin, and coflin. We surmise that FAK regulates leading edge and not dorsal ruffle formation because FAK is essential for the spatial redistribution of active Rac1 from these complexes to the leading edge.

Because FAK is one of the first proteins recruited to nascent focal complexes, we surmise that FAK initiates recruitment of active Rac1 to these newly formed sites via multifunctional protein complex formation. FAK associates with a number of adapter molecules through well-defined protein interaction sites that could serve as a Rac1 binding platforms. FAK-dependent recruitment of Rac1 could proceed through FAK/(CAS or paxillin)/Crk/DOCK 180 complex formation or a paxillin/PIX/Cool complex. Whether FAK-dependent activation of Rac1 at the leading edge is linked to the impaired spatial-temporal leading edge specific redistribution (or phosphorylation) of these or other FAK substrates are important questions for future studies.

There are a few possible explanations for why FAK might be essential for SMC investment of the conotruncus and coronary vasculature, but not for the recruitment of SMC to the distal vasculature (ie, carotid arteries). First, there may be cell type-specific differences in the factors that drive chemotaxis or regional differences in the levels and locale of these so-called motogens. In this regard, it is interesting to note that SMC chemotaxis toward PDGF was exceptionally sensitive to depletion of FAK, although chemotaxis toward serum was only modestly decreased in FAK null SMC (indicating that there are likely FAK-dependent and FAK-independent SMC motogens). The findings that depletion of PDGF receptors from CNC- or proepicardial cells phenocopies the OFT and coronary vessel defects observed in our FAK-depleted models and that PDGFs are expressed in high levels within the conotruncus supports the possibility that PDGF and FAK may work in concert to spatially regulate SMC motility in these regions. The second (not mutually exclusive) possibility is that a SMC invasive phenotype (that requires FAK activity) is required for SMC to cross tissue boundaries during both septal formation and coronary vasculogenesis (but not carotid vasculogenesis). Indeed, it may be possible that SMC coverage of carotid vessels occurs by a sheet-like movement/expansion of SMC along the continuously remodeling SMC-lined pharyngeal arches. In support of the possibility that FAK promotes an invasive phenotype, we showed that FAK is necessary for SMC podosome-mediated matrix degradation, and others have recently reported that FAK is essential for invadopodia-mediated matrix degradation in colon cancer cells. Lack of appropriate cortactin recruitment could also be causal for the impaired invasion observed in the FAK null SMC because cortactin has been shown to be necessary for regulating the localized recruitment and secretion of matrix metalloproteinases from lamellipodial-like membrane protrusions. Future studies will be necessary to more fully understand the requirement for SMC invasion versus chemotaxis during the formation of various vascular beds.

In conclusion, our studies indicate that FAK plays a SMC autonomous role in aortic/pulmonary septum and coronary vessel formation. Our studies in cultured SMC indicate that FAK activity is critical for SMC chemotaxis/invasion toward PDGF, a potent chemoattractant elaborated from the endothelium. Thus, we surmise that defective PDGF-dependent SMC recruitment leads to improper morphogenesis of FAK null vessels. We found that FAK functions to induce directional SMC motility by regulating the spatial and temporal locale of Rac1-dependent processes, including leading edge recruitment of the actin modifier, cortactin, which is necessary to stabilize lamellipodia and to form productive podosomes. It will be of future interest to evaluate the extent to which FAK may play a role in the pathogenesis of microaneurysms and other vascular diseases associated with defective recruitment of PDGF receptor β positive progenitors.

Sources of Funding
This work was supported in part by grants from National Institutes of Health, National Heart, Lung, and Blood Institute (HL-081844 and HL-071054) and the American Heart Association (0355776U) to Joan M. Taylor, National Institutes of Health, National Heart, Lung, and Blood Institute (HL-070953) and American Heart Association (0555476U) to Christopher P. Mack and National Institutes of Health HL-19242 and HL-93594 to Mark W. Majesky. Liisa J. Sundberg-Smith was supported by an American Heart pre-doctoral fellowship (0515329U).

Disclosures
No conflict of interest to declare.
References


Focal Adhesion Kinase Regulates Smooth Muscle Cell Recruitment to the Developing Vasculature


Arterioscler Thromb Vasc Biol. published online July 14, 2011;
Arteriosclerosis, Thrombosis, and Vascular Biology is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
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Print ISSN: 1079-5642. Online ISSN: 1524-4636

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SUPPLEMENTAL INFORMATION

Supplemental Information includes an expanded Methods section and additional data including:
seven figures and legends, one table, and reference citations.
Supplemental Methods

Antibodies, Reagents, and Constructs

The Pyk2 and phospho-specific ERK1/2 antibodies were purchased from Cell Signaling. Anti-vinculin, SMα-actin, SMγ-actin, and α-tubulin antibodies were purchased from Sigma. The N-term specific anti-FAK, C-term specific anti-FAK, cortactin, Rac1, ERK2, and anti-phospho-PDGF receptor β antibodies were purchased from Upstate. Paxillin antibody was purchased from BD Transduction Laboratories and anti-Cyclin D1 was purchased from Santa Cruz. The anti-phospho Y397FAK antibody was purchased from BioSource and the Texas-Red phalloidin was purchased from Molecular Probes. SM-22 antibody was a generous gift from Mario Gimona and SM-MHC antibody was obtained from U. Groeschel-Stewart. PDGFBB and TGF-β were purchased from Calbiochem. Ad5CMV Cre adenovirus was purchased from the University of Iowa Gene Transfer Vector Core and Ad5CMV LacZ adenovirus was purchased the University of North Carolina-Chapel Hill Viral Core. Both viruses were expanded using Puresyn Adenopure adenovirus purification kit according to manufacturer protocol. Flag-FAK variants were a generous gift from Dr. Tom Parsons (University of Virginia).

Primary SMC culture and treatment

Aortic SMC were isolated from 4-8 week old male fak^{floxed} mice as previously described. In brief, thoracic aortas were stripped of the endothelial and adventitial layers. Primary SMC were then isolated by trypsin and collagenase digestion. Only the cell preparations at least 85% pure are utilized for further experiments. Cells were maintained in DMEM: F12 (1:1) supplemented with 10% FBS and 1% penicillin-streptomycin and were used between passages 5 and 18. After infection with Cre or LacZ adenoviruses for 72 hr, cells were serum starved for 24 hr and treated with PDGF-BB (20ng/ml) for indicated times. For cortactin silencing experiments, cortactin specific short-interfering RNA (siRNA) oligos were obtained from Invitrogen with the following sequences:
GCUCUUCCAGCCCAACUATT[dT][dT] and Stealtho
GGAGAAGCAUGAGCUCAGAAAGAU[dT][dT]. SMC maintained in growth conditions were transfected with 150 nM control or a mixture of the two cortactin-specific siRNAs according to manufacturer's specifications using Dharmafect 1.

**Transwell migration assay**

SMC were plated on either 10 μg/ml fibronectin-coated or matrigel-coated transwell membranes (8μm; Bio-Coat) with the lower chamber containing serum-free media, 10% serum, or PDGF-BB (20 ng/ml, Calbiochem). After incubation at 37°C for 7 h, the upper surface of the membrane was scraped gently to remove non-migrating cells. The remaining cells were stained with 1% crystal violet and counted. Data represent the total number of cells in four separate fields for each condition.

**Western Blotting**

Western blots were performed using specific primary antibodies at a 1/1000 dilution (see Supplemental Methods for reagent details). Blots were washed in TBS-T followed by incubation with horseradish peroxidase conjugated secondary antibody at a 1/2000 dilution. Signals were visualized by using chemiluminescence reagents (Amersham).

**Immunostaining**

Tissue sections (10 μm) were deparaffinized, rehydrated, and antigen-retrieved in 10 mmol/L citrate buffer or 0.1% trypsin. After blocking in 10% goat serum, slides were incubated with primary antibodies at 1/200 dilution at 4°C overnight. Signals were detected using either biotinylated-, fluorescent or HRP linked secondary antibodies coupled with ABC detection kit (Vector) or diaminobenzidine.

Cells were fixed in 4% paraformaldehyde, permeabilized with 0.5% Triton X-100, and incubated with primary antibodies for 1 hr (1/250 for Rac1, all others 1/1000). Signals were detected using fluorescent-conjugated secondary antibodies. Filamentous actin and nuclei were visualized by staining.
with Alexa 488-conjugated phalloidin or DAPI. To quantify polarized Rac1 and cortactin labeling, lines were drawn through the nucleus to divide the cell into quartiles. Membrane localized protein in 1-2 quartiles was considered polarized (if the lamella was continuous), whereas labeling in 3-4 quartiles was considered random. All cells were scored in a blinded fashion.

**Kymographic Analysis**

Kymography was performed by using an Olympus IX70 inverted microscope that is equipped with a programmable Delta T motorized x,y,z stage and is encased in Plexiglas housing to control the internal environment (37°C, 5% CO2 and a relative humidity of 60%). Images were acquired at 40x magnification every 5 seconds (for kymography) or every 2 min (for random motility) by an Optronix DEI 750 CCD camera using OpenLab software (Improvision). Kymographic measurements were taken using the Multiple Kymograph plugin for ImageJ and analyzed as described previously.

**Rac GTPase Activity Assays**

Cells were lysed in Buffer A (50 mM Tris pH 7.6, 500 mM NaCl, 0.1% SDS, 0.5% deoxycholate, 1% Triton X-100, 10 mM MgCl2) containing 100 μM leupeptin, 1 mM 4-(2-aminoethyl) benzenesulfonyl fluoride hydrochloride, 20 μg/mL soybean trypsin inhibitor, and 0.05 trypsin inhibitory units/ml aprotinin. Lysates (200-500 μg/condition) were rotated with 30 μg immobilized GST-p21 binding domain (PBD) for 30 minutes at 4°C, then washed three times with Buffer B (50 mM Tris pH 7.6, 150 mM NaCl, 1% Triton X-100, 10 mM MgCl2) containing the protease inhibitors listed above. Complexes were pelleted by centrifugation, boiled in SDS-PAGE buffer, electrophoresed (15% SDS-PAGE), transferred to PVDF membrane and immunoblotted for total Rac1.

**Epicardial Explant Assays**

Proepicardial organs were isolated from HH16-17 quail embryos and placed into explant culture as described previously. After attachment (3-8 hrs post plating), explants were infected with 2.2 x 10^8 pfu of GFP- or GFP-FRNK (FAK related non-kinase) adenoviruses for the
indicated times. Explants were fixed in 4% paraformaldehyde and processed using immunocytochemistry as described above.

**Cell proliferation and apoptosis**

*fak<sup>flox/flox</sup>* SMC (pre-treated with Cre or LacZ adenovirus for 72 hr) were trypsinized and plated onto a 96-well microplate (5 x 10<sup>3</sup> cells/well). Cells were serum starved for 24 hr and treated with PDGFBB (20ng/ml) or EGF (100 ng/ml) for 48 hr. Cells were incubated with the formazen dye, WST-1 (10 µl; Roche) for 4 hr and the absorbance was read at 450 nm as per manufacturer’s instructions. Alternatively, BrdU (Sigma, 30 µg/mL) was administered to SMC grown on chamber slides. After 4 hrs, cells were fixed in 4% paraformaldehyde and stained using a BrdU detection kit (Invitrogen). Caspase activity was determined using the Caspase-Glo kit (Promega).

**Matrix degradation assays**

*fak<sup>flox/flox</sup>* SMC (pre-treated with Cre or LacZ adenovirus for 72 hr) were trypsinized and plated in serum-free medium onto glass chamber slides pre-coated with FN(10 ug/ml) plus 0.2% Oregon Green 488-Gelatin (Invitrogen) as previously described<sup>5</sup>. After 4 hr, cells were treated with vehicle or PDGF-BB for 90 min at 37°C. Cells were stained as described above and fluorescent images of cells were evaluated for patches of degradation. Alternatively, Cre- or LacZ-treated SMC were plated onto 96 well plates (4 x 10<sup>3</sup> cells/well) and incubated with DQ-gelatin (100µg/well; Molecular Probes) for 90 min in the presence of PDGF-BB. Fluorescence was read on a Polarstar plate reader (BMG Lab Technologies, Durham NC) at Ex/Em 495/515 nm.
**Supplemental Table 1.** Viable FAK\(^{\text{Wnt}}\) and FAK\(^{\text{cSMC}}\) offspring (expect 25%).

<table>
<thead>
<tr>
<th>Age</th>
<th>E10.5</th>
<th>E12.5</th>
<th>E13.5</th>
<th>P0</th>
<th>P4</th>
<th>P14</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>FAK(^{\text{Wnt}}) (viable/total)</strong></td>
<td>8/31</td>
<td>14/60</td>
<td>12/50</td>
<td>18/88</td>
<td>n.d.</td>
<td>0/43</td>
</tr>
<tr>
<td>% viable</td>
<td>26%</td>
<td>23%</td>
<td>24%</td>
<td>20%</td>
<td>n.d.</td>
<td>0%</td>
</tr>
<tr>
<td><strong>FAK(^{\text{cSMC}}) (viable/total)</strong></td>
<td>n.d.</td>
<td>n.d.</td>
<td>n.d.</td>
<td>8/42</td>
<td>0/29</td>
<td>n.d.</td>
</tr>
<tr>
<td>% viable</td>
<td>n.d.</td>
<td>n.d.</td>
<td>n.d.</td>
<td>19%</td>
<td>0%</td>
<td>n.d.</td>
</tr>
</tbody>
</table>

E, embryonic day; P, postnatal day; n.d., not determined.
Supplemental References

1. Richardson A, Malik RK, Hildebrand JD, Parsons JT. Inhibition of cell spreading by expression of the C-terminal domain of focal adhesion kinase (FAK) is rescued by coexpression of Src or catalytically inactive FAK: a role for paxillin tyrosine phosphorylation. *Mol Cell Biol.* 1997;17:6906-6914.


Supplemental Figure Legends

Supplemental Figure 1. Conditional deletion of fak in \textit{wnt-1} and \textit{n}\textit{kx2-5} expressing cells leads to PTA that is not associated with defective SM differentiation.  

A. Similar SMC differentiation is observed surrounding the pharyngeal arches (PA) in E11 Control and FAK\textsuperscript{wnt} mice as shown by Ú-SMA IHC (brown).  

B. Cross sections through \textit{wnt}-derived regions of the left common carotid artery of E13 embryos stained with SM-22 antibody reveal similarly intense staining in FAK\textsuperscript{wnt} vessels in comparison to genetic controls.  

C. P0 neonates were fixed and stained with X-gal (blue) to identify \textit{wnt}-derived cells. Red arrows denote septated pulmonary and aortic arteries in genetic control (left) and un-septated OFT in FAK\textsuperscript{wnt} neonate (right). Yellow arrows reveal ß-galactosidase expression throughout the common carotids in the FAK\textsuperscript{wnt} neonate.  

D. Heart sections of P0 genetic control or FAK\textsuperscript{wnt} neonates showed similarly intense SM-22 staining (brown) in coronary arteries (not targeted for FAK deletion).  

E. Transverse sections through the cardiac outflow tracts of P0 genetic control or FAK\textsuperscript{nk} neonates stained with ß-SMA. Data are representative of at least 5 mice that exhibited PTA (approximately 13\% of the FAK\textsuperscript{nk} mice analyzed).  


F. Alcian blue/alizarin red stain of P0 FAK\textsuperscript{wnt} and genetic control P0 neonate reveals grossly normal skeleton and cranial facial development.

Supplemental Figure 2. \textit{wt}l\textsuperscript{Cre} induces coronary-restricted recombination.  

\textit{wt}l\textsuperscript{Cre} and Rosa26R\textsuperscript{LacZ} reporter mice were intercrossed and double positive P0 transgenic hearts were subjected X-gal staining, embedded, sectioned and imaged. Data are representative of at least 5 hearts.
Supplemental Figure 3. Depletion of FAK does not affect cardiac growth or other major vessels that are not targeted for FAK deletion in the FAK<sup>cSMC</sup> neonates.  A. Transverse sections of H&E stained P0 FAK<sup>cSMC</sup> and genetic control hearts.  Note similar heart size and shape.  Arrows demarcate distended coronary vessels apparent in FAK<sup>cSMC</sup> heart.  B. Quantification of ventricular cross-sectional area showing no significant difference in heart size between P0 FAK<sup>cSMC</sup> and genetic control hearts (mean +/- SE; n=8).  C. Left carotid artery of P0 FAK<sup>cSMC</sup> and genetic control mice revealed similar SMC coverage as measured by α-SMA staining.

Supplemental Figure 4. Depletion of FAK does not affect cell proliferation or apoptosis in the FAK<sup>cSMC</sup> hearts.  A. Confocal images of E15.5 FAK<sup>cSMC</sup> (right) and genetic control hearts (left) co-stained for β-gal (green) and the proliferation marker phosphor-Histone H3 (Ser 10) (top, red) or the apoptotic marker TUNEL (bottom, red). Nuclei are labeled with DAPI (blue). Scale bar = 20 μm. B. Quantification of cell proliferation and apoptosis in E15.5 FAK<sup>cSMC</sup> and genetic control hearts (n = 4 each). Proliferation is presented as a percentage of phosphor-Histone H3 (Ser 10) <sup>+</sup> / β-Gal <sup>+</sup> double-positive cells to β-Gal <sup>+</sup> cells. Apoptosis is presented as a percentage of TUNEL <sup>+</sup> / β-Gal <sup>+</sup> double-positive cells to β-Gal <sup>+</sup> cells. C. Quantification of cell proliferation and apoptosis in P0 FAK<sup>cSMC</sup> and genetic control hearts (n = 4 each).

Supplemental Figure 5. FAK activity is not required for coronary SMC differentiation from pro-epicardial cells.  HH stage 17 quail pro-epicardial explants were incubated with GFP- or GFP-FRNK adenoviruses and cultured in serum-containing medium for 24-96 hours.  A, B) Phase and fluorescent images were collected and area of the explants was assessed. Lines in
panel A show the radii of each colony. Data are presented as mean +/- SE (N=3, n=10-16) in panel B. C, D) Explants were fixed 72 hr following treatment with viruses and stained with anti SMγA antibody.

Supplemental Figure 6. FAK is not required for growth or PDGFBB-dependent mitogenic signaling in SMC. A. fak<sup>ff</sup> SMC maintained in 10% serum were infected with either LacZ or Cre adenovirus for 72 hrs and cell lysates were immunoblotted with indicated antibodies. B. Cells treated as above were fixed and stained with phalloidin and indicated antibodies. Data are representative of at least four experiments. C. fak<sup>ff</sup> SMC were serum starved for 24 hrs prior to treatment with PDGFBB (20 ng/ml) for 15 min and cells were stained with pY397 FAK. Note activation and redistribution of active FAK to naciente leading-edge focal adhesions. D. LacZ- or Cre-pretreated fak<sup>ff</sup> SMC were serum starved for 24 hrs prior to treatment with either PDGFBB (20 ng/ml) or EGF (100 ng/ml) for 48 hrs. Cell growth was assessed by WST-1 activity as described in the Materials and Methods section. Data represent mean +/- SE of three separate experiments E. Cre or LacZ infected fak<sup>ff</sup> SMC were serum starved for 24 hrs prior to treatment with PDGFBB (20 ng/ml) for the times indicated. Cells were lysed, electrophoresed and Western blotting was performed with anti-FAK, anti-active pERK1/2 and anti-ERK antibodies. F. fak<sup>ff</sup> SMC were infected with either Cre or LacZ adenovirus for 72 hrs. Cells were serum starved for 4 hrs prior to treatment with either 10% serum (SM) or PDGFBB (20 ng/ml) for 24 hrs. Cells were lysed, electrophoresed, and Western blotting was performed with anti-FAK, anti-Cyclin D1 and anti-ERK antibodies. G. LacZ or Cre infected fak<sup>ff</sup> SMC were treated with 20 ng/ml PDGF-BB for the times indicated and total Rac1 activity was determined using the GST-PBD precipitation assay.
Supplemental Figure 7. Rac1 and cortactin are required for PDGF-stimulated leading edge formation and chemotaxis. A,B SMC were treated with NSC23766 (10 µM, Calbiochem) for 15 min. prior to treatment with 20 ng/ml PDGF-BB for 2.5 min (A) and 15 min (B). Cells were stained with anti-cortactin antibodies and scored for dorsal ruffle and leading edge formation. Results are mean±SEM of 200-250 cells from 3 independent experiments. *p< .05. C. Cells treated as above were plated on fibronectin-coated inserts (10 µg/ml; Bio-Coat) in serum-free media using PDGF-BB (20 ng/ml) as the chemoattractant (mean±SEM of cells counted in 4 fields from 3 independent experiments; *p< .05). D. SMC were treated with control siRNA (cSi) or cortactin siRNAs (CTNsi; see Methods) for 72 hr and cell extracts were blotted with antibodies directed against cortactin and ERK (loading control). E-G. SMC pre-treated with cSi (not shown) or CTNsi for 72 hr were treated with 20 ng/ml PDGF-BB for 2.5 min (E,F), 15 (E,G), or 30 min (E). Cells were co-stained with pY118-paxillin (pPAX, red), phalloidin (green) and anti-cortactin (blue) antibodies and scored for dorsal ruffle and leading edge formation. Results are mean±SEM of 150-200 cells from 3 independent experiments. *p< .05. H. CSi or CTNsi pre-treated cells were plated on fibronectin-coated inserts (10 µg/ml; Bio-Coat) in serum-free media using PDGF-BB (20 ng/ml) as the chemoattractant (mean±SEM of cells counted in 4 fields from 3 independent experiments; *p< .05).
Cheng et al. Supplemental Figure 1
Cheng et al. Supplemental Figure 2
Cheng et al. Supplemental Figure 3
Cheng et al. Supplemental Figure 4
Cheng et al. Supplemental Figure 6