Inheritance of High Density Lipoprotein and Lipoprotein Lipase and Hepatic Lipase Activity

Timo Kuusi, Y. Antero Kesäniemi, Matti Vuoristo, Tatu A. Miettinen, and Markku Koskenvuo

The role of genetic and environmental factors in the regulation of plasma high density lipoprotein (HDL) was estimated in 17 monozygotic (MZ) and 18 dizygotic (DZ) male twins randomly selected from the Finnish Twin Cohort Study. In addition to HDL cholesterol, we determined the HDL subfractions, HDL₂ and HDL₃, and the major HDL apoproteins (apo) A-I and A-II. The activities of lipoprotein lipase (LPL) and hepatic lipase (HL) were also assayed from postheparin plasma to get information on their possible contribution to the heritability of HDL. Evidence for the genetic component in the regulation of plasma HDL received support from the heritability estimate of 0.34. The different heritability estimates of HDL₂ and HDL₃ (h² of 0.56 and <0, respectively) support the idea that the HDL subfraction distribution might be important in the genetic regulation of plasma HDL level. This also received support from the heritability of apo A-I (h² = 0.66), mainly varying in HDL₂, and the lack of it in apo A-II, found mainly in HDL₃. These conclusions were strengthened by standardizing the data with relative ponderosity. Postheparin plasma HL activity had a high pairwise correlation coefficient in the MZ twins (r = 0.80, p < 0.001), whereas LPL displayed no within-pair correlation. Neither of the lipolytic enzymes, LPL or HL, showed any correlation in the DZ twins. Therefore, it is suggested that part of the genetic regulation of the HDL and its subfraction distribution might be mediated through the activity of HL.

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A familial clustering of coronary heart disease (CHD) is evident from several studies. However, it is not known whether genetic or shared environmental factors play a dominating role in the aggregation of the disease in some families. Recently, it has been shown in a large study on siblings that the clustering of CHD might not be due largely to the major classical risk factors of CHD such as serum cholesterol or blood pressure. This suggests that other lipoprotein abnormalities (as well as nonlipoprotein factors) warrant exploration as possible factors that might be involved in the familial aggregation of CHD.

A low level of serum high density lipoprotein (HDL) is a well-established risk factor of CHD. Accordingly, much interest has recently been focused on environmental and genetic factors regulating its serum level. In fact, HDL cholesterol shows a correlation between siblings ranging from 0.16 to 0.46. In twin studies, some genetic influence has been demonstrated in the variance of both major HDL apoproteins, apo A-I and apo A-II. However, studies on HDL in a large pedigree have supported a polygenic mode of inheritance. Studies on hypoalphalipoproteinemia have suggested a major gene effect in the familial aggregation of low HDL cholesterol in addition to the multifactorial background of the disorder. Thus, the regulation of plasma HDL by genetic factors appears to be quite complex in nature.

Plasma HDL is actually a mixture of different lipoproteins. At least two major subfractions, HDL₂ and HDL₃, can be identified in the HDL density range. The metabolism of the subfractions is interrelated, and it appears to be closely related to the function of two endothelial lipolytic enzymes, lipoprotein lipase (LPL) and hepatic endothelial lipase (HL). Thus, HDL₂ is formed during catabolism of plasma triglyceride-rich lipoproteins by LPL, whereas its degradation is associated with the function of HL. Information about the genetic aspects of interrelations between HDL and the two lipolytic enzymes are lacking. Therefore, we have calculated heritability estimates for plasma HDL, apo A-I and A-II, and the subfractions, HDL₂ and HDL₃, in twins. In addition, the postheparin plasma activities of LPL and HL were measured to get information on the possible role of the lipolytic enzymes in the genetic regulation of plasma HDL concentration.

Methods

Subjects

A group of 17 monozygotic (MZ) and 18 dizygotic (DZ) male twins living in the Helsinki metropolitan area were
randomly selected from the Finnish Twin Cohort Study that included about 15,000 twin pairs. The zygosity diagnosis in the registry is based on a mailed questionnaire which is validated by using blood markers as described in detail earlier. The twins in the present study were aged between 48 and 63 years and their weight ranged from 56 to 90 kg with a mean of 78 kg ± 9.5 kg (SD). The subjects were studied as outpatients at the Central University Hospital, and the protocol of the study was approved by the Ethical Committee. The subjects gave their written informed consent.

Lipids and Lipoproteins

Blood was obtained after an overnight fast at 8:00 A.M. Serum was separated after clotting at 4°C, and the lipoprotein analysis was started the same day. Very low density lipoprotein (VLDL) was separated by ultracentrifugation in a Beckman L-70 ultracentrifuge (Beckman Instruments Incorporated, Palo Alto, California) in a Kontron T75 45.6 rotor (Kontron AG, Basel, Switzerland) operated at 4°C for 18 hours at a density of 1.006 g/ml and at 38,000 rpm. LDL and HDL were then precipitated sequentially from the bottom fraction using heparin-MnCl₂ and dextran sulphate by the method of Gidez et al. Cholesterol concentration was determined using an enzymatic kit (No 187313, Boehringer GmbH, Mannheim, FRG) in a Kone Olli-D fully automated analyzer (Kone Limited, Espoo, Finland).

Apolipoprotein A-I and A-II

These concentrations were determined by immunoturbidimetry using monospecific antisera against apo A-I and A-II (Catalogue Nos. 726 428 and 726 486, respectively, Boehringer Mannheim, FRG).

Postheparin Plasma Lipase Activities

After taking samples for lipoprotein analysis, 100 IU/kg of heparin was injected intravenously. Blood was taken after 5 and 15 minutes into precooled heparinized tubes, and plasma was separated at 4°C and stored frozen at −20°C until assayed. Postheparin plasma LPL and HL activities were determined by a selective immunochromatographic method using gum arabic stabilized tris(1-14C-oleyl) glycerol as a substrate. The activity of LPL was determined after inactivation of HL by a specific antiserum (No 187313, Boehringer Mannheim, FRG) in a Kone Olli-D fully automated analyzer (Kone Limited, Espoo, Finland).

Statistical Analysis

Statistical treatment of the data was performed using a Honeywell DPS computer using BMDP statistical software (Biomedical Data Processing System, Department of Biostatistics, University of California, California). The MZ and DZ pairs were compared using the heritability estimate of Falconer calculated by the formula:

\[ h^2 = 2 \times (r_{MZ} - r_{DZ}) \]

where \( r_{MZ} \) and \( r_{DZ} \) are the pairwise intraclass correlations of the MZ and DZ twins calculated using double entry of the data. In addition, we calculated the intrapair variances by the formula:

\[ \Sigma(A - B)^2/2N \]

Values for cholesterol and apolipoproteins are given as mg/dl. Values for lipoprotein lipase and hepatic lipase are given as umol/hr/ml. All values are means ± SEM. MZ = monozygous; DZ = dizygous.

Results

The two groups of twins were relatively similar with respect to their plasma HDL cholesterol and apo A-I and A-II concentrations (Table 1). The cholesterol concentrations of the HDL subfractions, HDL₂ and HDL₃, and the postheparin plasma lipolytic enzymes, LPL and HL, were also similar in the MZ and DZ twins.

The plasma HDL cholesterol concentration was positively correlated with postheparin plasma LPL activity in all subjects (\( r = 0.33, p < 0.01 \)) and in the MZ group (Figure 1A). The respective correlation of plasma HDL cholesterol with postheparin plasma HL activity was negative in the MZ twins (Figure 1B), whereas no significant relationships existed between HDL cholesterol concentration and postheparin plasma lipase activities in DZ twins.

The pairwise correlation coefficients for plasma HDL cholesterol and postheparin plasma lipase activities are shown in Table 2. This correlation was significant for HDL cholesterol in both MZ and DZ twins. However, the correlations in the MZ pairs were higher in comparison with the DZ pairs. Thus, a heritability estimate of 0.34 could be obtained for HDL cholesterol. Calculation of the estimate for HDL subfractions revealed that this was mainly due to the

Table 1. Plasma Cholesterol and Apolipoprotein Concentrations and Postheparin Plasma Lipase Activities

<table>
<thead>
<tr>
<th>Parameters</th>
<th>MZ twins</th>
<th>DZ twins</th>
</tr>
</thead>
<tbody>
<tr>
<td>n = 34</td>
<td>n = 36</td>
<td></td>
</tr>
<tr>
<td>Cholesterol</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HDL</td>
<td>44.1 ± 9.3</td>
<td>50.7 ± 12.0</td>
</tr>
<tr>
<td>HDL₂*</td>
<td>13.5 ± 4.6</td>
<td>16.6 ± 9.3</td>
</tr>
<tr>
<td>HDL₃*</td>
<td>34.8 ± 7.0</td>
<td>37.2 ± 6.1</td>
</tr>
<tr>
<td>Apolipoprotein</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Apo A-I</td>
<td>132.5 ± 14.0</td>
<td>142.7 ± 21.0</td>
</tr>
<tr>
<td>Apo A-II</td>
<td>37.3 ± 4.2</td>
<td>36.0 ± 4.2</td>
</tr>
<tr>
<td>Lipoprotein lipase</td>
<td>31.1 ± 8.2</td>
<td>36.1 ± 15.8</td>
</tr>
<tr>
<td>Hepatic lipase</td>
<td>20.4 ± 12.7</td>
<td>26.8 ± 11.6</td>
</tr>
</tbody>
</table>

*HDL subfractionation was performed in 20 MZ and 24 DZ men.
high heritability for HDL2 cholesterol. In addition, genetic regulation could be observed for plasma apo A-I but not for apo A-II concentration (Table 2). Since there appeared to be considerable sharing of relative ponderosity between both sets of twins (r of body mass index (BMI) 0.58 and 0.40 for MZ and DZ twins, respectively) we also calculated the pairwise correlations after adjustment with BMI (shown in parenthesis in Table 2). Thus, it is evident that the heritability estimates for HDL and its subfractions were only slightly changed by the adjustment with BMI. However, the situation was opposite with apo A-I and A-II, as it appeared that the h² for both apoproteins were significantly improved by standardizing the data with BMI.

Postheparin plasma HL activity displayed a highly significant pairwise correlation in MZ twins, whereas no significant correlation could be observed in their LPL activities. The postheparin plasma lipases, LPL and HL, showed low pairwise correlation in the DZ group (Table 2). Accordingly, a high heritability estimate (h² = 1.18) could be calculated for HL activity in our twin material, and this appeared to be rather independent of the effect of ponderosity as judged from the correlations obtained after adjusting with BMI.

The heritability data on HDL and the factors that influence its concentrations were also evaluated by calculating the intrapair variances. The F-values thus obtained were most significant for HDL2, HL, and apo A-I (Table 3). This suggests that HDL subfraction distribution and its regulators might have a marked role in the mediation of genetic influence on plasma HDL concentration.

Discussion

Several twin studies have been performed to evaluate the role of nature vs. nurture as a determinant of plasma HDL cholesterol. This research was largely stimulated by the finding of low HDL cholesterol concentrations in male relatives of patients with coronary heart disease. CHD is common in families with hypolalphalipoproteinemia and Tangier disease but is rare in pedigrees with hyperalphalipoproteinemia, a familial disorder with grossly elevated plasma HDL cholesterol. The common risk factors such as serum cholesterol and hypertension have failed to explain much of the genetic/familial transmission of CHD. Thus familial aggregation of low HDL cholesterol might have a role in the familial aggregation of atherosclerotic disease. Third et al. have, in fact, demonstrated that low HDL cholesterol as an isolated dyslipoproteinemia is a heritable disorder and is associated with accelerated premature coronary heart disease and stroke.

HDL cholesterol concentration is strongly regulated by many exogenous factors, such as diet, exercise, and alcohol intake. In the present study the genetic influence on HDL cholesterol concentration was evaluated by comparing the pairwise correlation of HDL in both MZ and DZ twins. The heritability estimate thus obtained (h² = 0.34) is very close to that calculated in a previous twin study (h² = 0.35) and in a large pedigree using path analysis (h² = 0.59). In the present study, we also performed HDL subfractionation, which was not reported in earlier twin studies, to evaluate the contribution of the HDL subfractions to the inheritance of HDL cholesterol level. It is well known that most variation in plasma HDL cholesterol is due to the HDL₂ subfraction. Therefore it was not surprising to find high h² for HDL₂ but not for HDL₃ cholesterol (Table 2). This was confirmed by calculating the heritability estimates by the intrapair variance method (Table 3). These results suggest that factors regulating the HDL subfraction distribution might also participate in a genetic regulation of plasma HDL.
Earlier studies have resulted in conflicting data on the heritability of the major HDL apoproteins, apo A-I and A-II. Thus, high heritability has been reported for apo A-I but not for apo A-II, a finding contrasted in other studies. In the present study we could demonstrate a genetic influence on serum apo A-I but not on apo A-II by both the Falconer and the Intrapair variance methods. The adjusted heritability estimate was obtained (Tables 2 and 3). This occurs at least partly during catabolism of triglyceride-rich lipoproteins by LPL. Degradation of HDL2 and its conversion back to HDL3 is initiated by another lipolytic enzyme, HL. Familially increased HDL cholesterol is associated with either increased LPL or decreased HL activities. In normal subjects, plasma HDL (HDL3) cholesterol concentration is positively correlated with LPL and negatively with HL activity. This was also the case in all the subjects and in the MZ twins of the present study (Figure 1).

Table 2. Within-Pair Correlation Coefficients for Cholesterol Concentrations and Lipase Activities in Monozygotic and Dizygotic Twins

<table>
<thead>
<tr>
<th>Parameters</th>
<th>MZ twins</th>
<th>DZ twins</th>
<th>Heritability estimates*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cholesterol</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HDL</td>
<td>0.70</td>
<td>&lt;0.01</td>
<td>0.53</td>
</tr>
<tr>
<td></td>
<td>(0.73)</td>
<td></td>
<td>(0.56)</td>
</tr>
<tr>
<td>HDL2</td>
<td>0.74</td>
<td>&lt;0.05</td>
<td>0.46</td>
</tr>
<tr>
<td></td>
<td>(0.79)</td>
<td></td>
<td>(0.58)</td>
</tr>
<tr>
<td>HDL3</td>
<td>0.14</td>
<td>NS</td>
<td>0.39</td>
</tr>
<tr>
<td></td>
<td>(0.37)</td>
<td></td>
<td>(0.42)</td>
</tr>
<tr>
<td>Apolipoprotein</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Apo A-I</td>
<td>0.36</td>
<td>NS</td>
<td>0.03</td>
</tr>
<tr>
<td></td>
<td>(0.63)</td>
<td></td>
<td>(0.06)</td>
</tr>
<tr>
<td>Apo A-II</td>
<td>0.26</td>
<td>NS</td>
<td>0.32</td>
</tr>
<tr>
<td></td>
<td>(0.49)</td>
<td></td>
<td>(0.24)</td>
</tr>
<tr>
<td>Lipoprotein lipase</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>0.22</td>
<td>NS</td>
<td>0.37</td>
</tr>
<tr>
<td></td>
<td>(0.50)</td>
<td></td>
<td>(0.35)</td>
</tr>
<tr>
<td>Hepatic lipase</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>0.80</td>
<td>&lt;0.001</td>
<td>0.21</td>
</tr>
<tr>
<td></td>
<td>(0.77)</td>
<td></td>
<td>(0.23)</td>
</tr>
</tbody>
</table>

The correlation coefficients calculated after standardization of the data with body mass index are given in parentheses. MZ, monozygous; DZ, dizygous.

*Heritability estimates were calculated according to the formula of Falconer: h^2 = 2 x (r^2 - r^2_DZ). Determined in 10 MZ and 12 DZ twins.

Table 3. Intrapair Variances for High Density Lipoproteins, Apolipoproteins A, and Lipase Activities in Monozygotic and Dizygotic Twins

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Intrapair variance</th>
<th>Heritability index</th>
<th>F ratio†</th>
</tr>
</thead>
<tbody>
<tr>
<td>HDL</td>
<td>24.4</td>
<td>0.62</td>
<td>2.64‡</td>
</tr>
<tr>
<td>HDL2</td>
<td>5.9</td>
<td>0.87</td>
<td>7.60§</td>
</tr>
<tr>
<td>HDL3</td>
<td>39.4</td>
<td>0.64</td>
<td>0.81 NS</td>
</tr>
<tr>
<td>Apo A-I</td>
<td>145.4</td>
<td>0.64</td>
<td>2.77‡</td>
</tr>
<tr>
<td>Apo A-II</td>
<td>20.7</td>
<td>0.60</td>
<td>0.63 NS</td>
</tr>
<tr>
<td>LPL</td>
<td>67.8</td>
<td>0.55</td>
<td>2.23 NS</td>
</tr>
<tr>
<td>HL</td>
<td>31.7</td>
<td>0.68</td>
<td>3.17‡</td>
</tr>
</tbody>
</table>

LPL = lipoprotein lipase, HL = hepatic lipase, NS = not significant. MZ = monozygous; DZ = dizygous.

†The heritability index was calculated using the formula h^2 = (intrapair variance_DZ - intrapair variance_MZ)/intrapair variance_MZ. The F ratios were calculated by dividing the intrapair variance for DZ with that for MZ.

‡p<0.05; §p<0.01.
data support these previous findings by demonstrating that the genetic regulation is stronger for HL than LPL.

The MZ twins in the present study, even though living apart, may have shared similar dietary habits, physical activities, and other environmental factors. Thus, the heritability estimates for HDL and HL activity are likely to be maximal estimations for the genetic regulation rather than strict absolute values. At any rate, it seems quite obvious that HL, and probably through its activity, HDL₂ are more powerfully genetically regulated than LPL.

References


Index Terms: HDL • apolipoprotein A-I • apolipoprotein A-II • hepatic lipase • lipoprotein lipase • twin studies • heritability
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