

## Cardiovascular Immunotherapy and the Role of Imaging

Eva Zupančič, Zahi A. Fayad, Willem J.M. Mulder

Despite significant advances in prevention and treatment, cardiovascular diseases (CVDs) remain the most common cause of death in the United States.<sup>1-3</sup> In part, and compared with treatment developments in oncology, this is because of the lack of current generation CVD precision therapies.<sup>4-6</sup> Most CVDs are caused by atherosclerosis, a disease process that causes thickening of the arterial wall because of inflammation and lipid accumulation, resulting in plaque formation.<sup>7,8</sup> Macrophage dynamics plays a central role in atherosclerosis progression, vessel wall destabilization, and—as has been recently discovered—plaque aggravation because of myocardial infarction.<sup>9-11</sup> Therefore, and despite current standardized treatments, ≈25% of patients who had myocardial infarction or stroke will experience secondary major adverse cardiovascular events, often more harmful than the primary event.<sup>12-14</sup> To break this vicious cycle, innovative and tailored therapeutic strategies are needed.

Various targeted immunotherapies that showed anticancer potential in *in vitro* screens also displayed *in vivo* potential in mouse models. Such proof-of-concept studies in validated syngeneic mouse tumor models not only provide information on *in vivo* efficacy but also disclose the targeted immunotherapies' underlying mechanism of action and safety profile. However, immunotherapies' high costs and the identification of amendable patients compromise clinical translation.<sup>15,16</sup> Continued and focused efforts are needed to understand patient responsiveness and reduce high dropout rates in trials. A parallel effort focused on the development of novel imaging tools that can guide patient selection may offer a potential solution.

Positron emission tomography (PET) can be used to trace immunotherapies. So-called immunoPET uses antibodies, developed for cancer treatment, that selectively recognize a specific epitope on target cells. The antibodies are labeled with radioactive isotopes, such as zirconium-89 or iodine-124, with relatively long decay half-lives of 3.27 and 4.18 days, respectively, to visualize their accumulation in human tissues and study their interaction within the tumor microenvironment. ImmunoPET can be complemented with computed tomography and MRI to determine pathological and morphological changes.<sup>17-19</sup>

In a preclinical setting, immunoPET has been applied to study a novel combination therapy, involving a STEAP1 antibody conjugate with a chemotherapeutic agent, for the treatment of prostate cancer.<sup>18</sup> In an early stage clinical trial, implementation of immunoPET enabled visualizing bone metastases for the first time. ImmunoPET in a small cohort of 56 patients with advanced breast cancer, who received radio-labeled therapeutic trastuzumab, revealed that in 29% of the patients, antibody internalization was not ample robust.<sup>18,20</sup> Novel combination approaches of immunotherapy and noninvasive imaging enable clinicians and researchers to optimize dosage protocols and obtain deeper insights into tissue distribution and the immune response. In spite of these impressive recent advances in oncology, the principle of empowering a patient's own immune system to fight disease has yet to be applied for reversing atherosclerosis and its clinical manifestations.

Despite the slower development rate of atherosclerosis and CVD treatments, novel immunotherapies are on the rise. The first targeted CVD therapy that has been developed is a PCSK9 (proprotein convertase subtilisin-kexin type 9) antibody for the treatment of hypercholesterolemia.<sup>5</sup> Its approval paved the way for the clinical evaluation of an actual anti-inflammatory CVD immunotherapy directed against interleukin-1 $\beta$  (IL-1 $\beta$ ) in the CANTOS trial (Canakinumab Anti-Inflammatory Thrombosis Outcomes Study). The disclosure of the primary end point led to a lot of excitement about the trial's results, suggesting an extensive reduction of secondary cardiovascular events.<sup>21</sup>

In this review, we discuss integration of targeted therapy, immunologic approaches, and noninvasive imaging for the management of CVD. We highlight advancements in contemporary techniques that might improve our understanding of targeted therapies' behavior and their effect on plaque morphology, inflammation, and macrophage dynamics in CVD. Finally, we discuss merging possibilities of various techniques and applications, their limitations, and challenges.

### Targeted Therapies

Atherosclerosis is initiated by the arterial wall accumulation of atherogenic lipoproteins, namely low-density lipoproteins (LDL).<sup>22,23</sup> This is accompanied by monocyte recruitment and macrophage accumulation in the vessel wall.<sup>24-26</sup> Macrophages' uncontrolled phagocytosis of LDL leads to the generation of foam cells,<sup>27</sup> inducing early stage plaque deposition in childhood.<sup>13,28</sup> In advanced disease, the inflamed atherosclerotic plaque can rupture and induce an acute coronary syndrome or myocardial infarction.<sup>7,29-31</sup>

Current therapies are focused on lipid lowering and ineffectively reverse atherosclerosis. Targeting the immune system's capacity to resolve inflammation represents a compelling alternative strategy for treating atherosclerosis. At present, 3 main atherosclerosis immunotherapy approaches are under investigation, focusing on lipoprotein metabolism,

From the Translational and Molecular Imaging Institute, Icahn School of Medicine at Mount Sinai, New York, NY (E.Z., Z.A.F., W.J.M.M.); and Department of Medical Biochemistry, Academic Medical Center, Amsterdam, The Netherlands (W.J.M.M.).

Correspondence to Willem J.M. Mulder, PhD, Translational and Molecular Imaging Institute, Icahn School of Medicine at Mount Sinai, 1470 Madison Ave, New York, NY 10029. E-mail Willem.mulder@mssm.edu

(*Arterioscler Thromb Vasc Biol.* 2017;37:e167-e171. DOI: 10.1161/ATVBAHA.117.309227.)

© 2017 American Heart Association, Inc.

*Arterioscler Thromb Vasc Biol* is available at <http://atvb.ahajournals.org>  
DOI: 10.1161/ATVBAHA.117.309227

inflammation, and tissue repair. We will discuss all these approaches in the content of preclinical and clinical studies.

### Lipid Lowering

To this day, the best clinically established approach to treat atherosclerosis has been lowering LDL cholesterol<sup>32</sup> by gold standard statin therapy.<sup>33</sup> However, 2 recent statin trials, JUPITER (Justification for the Use of Statins in Primary Prevention: An Intervention Trial Evaluating Rosuvastatin trial) and ASTEROID (A Study To Evaluate the Effect of Rosuvastatin on Intravascular Ultrasound-Derived Coronary Atheroma Burden), have clearly shown that protection from CVD is limited. In the JUPITER trial, it was shown that despite standard treatments, 25% to 40% of patients who had a prior heart attack remain at increased risk of secondary major adverse cardiovascular events, that is, myocardial infarctions or stroke, within the first 5 years.<sup>30,34</sup> Consistent with this finding, the ASTEROID trial showed that after 2 years of treatment with rosuvastatin, a mere 0.6% reduction in plaque volume was achieved.<sup>35</sup> It is clear that atherosclerosis pathophysiology extends beyond lipids because 2 of 3 of patients experiencing a heart attack have normal LDL cholesterol levels. Hence, more effective approaches are being developed.

The first targeted therapy for the treatment of hypercholesterolaemia is an antibody against PCSK9.<sup>5</sup> PCSK9 is a circulating protein that regulates hepatic and serum LDL cholesterol levels.<sup>36,37</sup> Evolocumab, a human monoclonal antibody that blocks the interaction between PCSK9 and the LDL receptor, was evaluated in the FOURIER (Further Cardiovascular Outcomes Research With PCSK9 Inhibition in Subjects With Elevated Risk) randomized clinical trial. In combination with statin therapy, up to 59% of patients show LDL reductions from a median of 92 to 30 mg/dL. Moreover, during the 26-month study, an associated 15% decrease in cardiovascular risk was observed. ORION-1 (Trial to Evaluate the Effect of ALN-PCSSC Treatment on Low-Density Lipoprotein Cholesterol), a phase II trial designed to assess a small interfering RNA therapy directed against PCSK9, found reductions in LDL levels, ranging from 27.9% to 52.6%, depending on the regimen. Both studies suggest that targeting PCSK9 may help maintain consistent and effective reductions in LDL cholesterol levels.<sup>38–40</sup>

### Resolving Inflammation

It has been shown that in  $\approx 4$  of 10 patients that survived heart attack, the risk of secondary events is directly related to increased inflammation.<sup>34</sup> Chronic, nonresolving inflammatory atherosclerosis is a critical factor and a main culprit for atherosclerotic plaques progression, rupture, and thrombosis.<sup>40</sup>

IL-1 $\beta$  is a proinflammatory cytokine which is released by macrophages and plays a major role in vascular inflammation and modulation of atherosclerotic calcification.<sup>41,42</sup> In June 2017, it was announced that the primary end point of the CANTOS phase III study was met. Subcutaneous injections of canakinumab (ACZ885), a human monoclonal antibody that selectively neutralizes IL-1 $\beta$ , significantly lowered systemic inflammatory biomarker levels in postmyocardial infarction patients, reducing cardiovascular risk in patients with a prior heart attack and inflammatory atherosclerosis.<sup>21,43</sup> The detailed

trial's results' will be fully disclosed at an upcoming medical congress in the fall of 2017.

In recent years, various preclinical studies focused on the development of anti-inflammatory therapeutic strategies to reduce macrophage accumulation in plaques.<sup>44,45</sup> Cholesterol-lowering drugs called statins, also known as 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors, exhibit tangible anti-inflammatory actions at high doses.<sup>46</sup> However, oral statins are rapidly metabolized and secreted through the liver. Consequently, low amounts of drug are systemically available to exert anti-inflammatory effects at the vessel wall. Our group has developed a targeted approach to inhibit plaque macrophage proliferation in apolipoprotein E-deficient mice with advanced atherosclerosis.<sup>47,48</sup> To that aim, simvastatin was incorporated in high-density lipoprotein nanoparticles<sup>49</sup> to achieve targeted delivery to plaque macrophages.<sup>47,50</sup> A rapid reduction in macrophage burden was achieved as a result of a 1-week intravenous regimen. When supplemented with an ensuing 8-week oral statin treatment, the rapid inhibition of plaque inflammation could be maintained, resulting in favorable vessel wall remodeling.<sup>47,51</sup>

Although the plaque macrophage reduction in the above-mentioned study was achieved by targeting proliferation, blunting monocyte recruitment is an attractive alternative therapeutic strategy.<sup>52</sup> Dutta et al<sup>53</sup> have shown an effectiveness in impairing monocyte migration from the spleen to the vessel wall in atherosclerotic apolipoprotein E-deficient mice. The same group of principle investigators observed similar findings using polymeric (7C1) small interfering RNA-loaded nanoparticles with specific avidity toward endothelial cells. Concurrent delivery of 5 distinct small interfering RNAs inhibited adhesion molecule function and induced an  $\approx 40\%$  reduction in neutrophil, monocyte, and macrophage infiltration into atherosclerotic plaque lesions.<sup>54</sup>

### Tissue Repair and Remodeling

Maintaining tissue homeostasis and repairing injuries are fundamental survival mechanisms. In case of mechanical injury or infections, inflammation occurs in response to damage-associated and pathogen-associated molecular patterns. The activated immune system is primed to eliminate dead and dying cells from the site of injury.<sup>55–59</sup>

Key players in the regulation of the tissue repair and regeneration are macrophages,<sup>25,60,61</sup> which—on detection of anti-inflammatory cytokines that are released from the injured cells—shift into repair mode. IL-4 and IL-13 are the 2 main activators of host defense responses and prompt an anti-inflammatory and tissue repair macrophage phenotype.<sup>58,62,63</sup> These 2 cytokines are mainly secreted by T-helper 2 lymphocytes at the site of infection or injury.<sup>62,64</sup> On the other side, tissue repair macrophages are characterized by expressing high levels of IL-10, IL-1ra, and TGF- $\beta$  (transforming growth factor-beta). The latter drives tissue repair through targeting myofibroblast, by stimulating angiogenesis, enhancing production of extracellular matrix components, and thus recruitment of IL-13-producing leukocytes to the site of tissue injury.<sup>55</sup>

In a recently study by Bosurgi et al, it has been reported that IL-4 and IL-13 together with apoptotic cells are critical players in wound repair after helminth infections in the

lungs or in the gut.<sup>65–67</sup> RNA sequencing analyses identified increased expression of 61 wound healing genes involved in cell proliferation, chemotaxis, and cell adhesion.<sup>66</sup> Notably, epidemiological studies found that helminth infections reduce atherosclerosis risk,<sup>68,69</sup> suggesting the anti-inflammatory cytokines' unique role in inflammation resolution and tissue repair through, for example, the synthesis of collagen.<sup>70</sup>

### Integration of Imaging

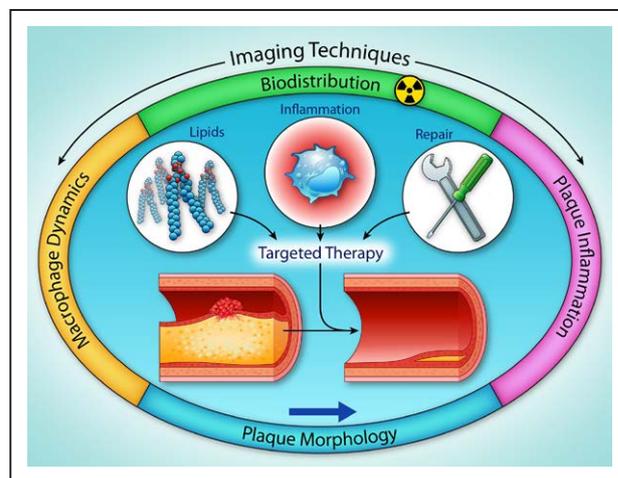
In the oncology field, noninvasive imaging is increasingly applied to study and assess immunotherapies, in the early phases of preclinical development, in clinical trials, as well as to select patients and predict therapeutic outcome. As described in the introduction, immunoPET with therapeutic antibodies is a promising tool for diagnostic and therapeutic treatments.

In the context of atherosclerosis, radiolabeling may help in understanding the targeted therapy's behavioral parameters, such as pharmacokinetics and plaque (macrophage) specificity.<sup>71</sup> At the same time, imaging can also be used to study immunotherapeutic agents' effects on vessel morphology and plaque stability through readouts reporting on vessel wall thickness and permeability, inflammation, and macrophage proliferation.<sup>72,73</sup>

Compared with other available anatomic imaging methods, such as ultrasound,<sup>74</sup> magnetic resonance, or computed tomography, PET imaging<sup>75–77</sup> has the advantageous ability to visualize minuscule concentrations of tracers that target specific molecular process. PET vascular inflammation imaging with fluorodeoxyglucose has become important in atherosclerosis research<sup>78</sup> because of fluorodeoxyglucose's accumulation in cells with glycolytic rate.<sup>79</sup> Several studies showed that fluorodeoxyglucose uptake, when measured in the arterial wall in vivo, reflects the level of macrophage accumulation within the atheroma.<sup>80</sup> Consequently, fluorodeoxyglucose-PET imaging is increasingly used to evaluate therapeutic approaches targeting atherosclerosis.<sup>81,82</sup>

Although fluorodeoxyglucose-PET imaging adequately addresses certain pathophysiological and treatment-related questions, the specificity for inflammation with this agent is not clearly defined because of the variable affinity for glucose of all cells in the body. In addition, high fluorodeoxyglucose activity in the blood pool and tissues near the vessel wall complicates quantification. Thus, there are opportunities for other imaging agents for atherosclerosis, such as vascular microcalcification imaged with <sup>18</sup>F-sodium fluoride<sup>83,84</sup> or somatostatin receptors imaged, present in high concentration on inflammatory leukocytes, imaged with <sup>68</sup>Ga-DOTATATE.<sup>76</sup> Further investigation of the performance characteristics of these new and other agents<sup>75</sup> are ongoing.

Combined and simultaneous PET/magnetic resonance is an exciting novel imaging modality that can assess disease activity alongside assessments of cardiac anatomy, function, and tissue composition during a single scan.<sup>77,85,86</sup> The lower associated radiation doses may be of particular importance for the clinical imaging of younger patients. In this research arena, beyond the ability to easily combine and coregister already established magnetic resonance and PET imaging techniques in a single scan, many researchers are seeking novel applications that may advance the state-of-the-art even further. Although technological and operational obstacles



**Figure.** Cardiovascular immunotherapy can be integrated with noninvasive imaging to probe biodistribution and targeting, as well as to monitor therapeutic efficacy, systemically and at the level of atherosclerotic lesions.

persist, these are rapidly being overcome, positioning PET/magnetic resonance as a useful new imaging modality for the investigation of CVD. Further clinical trials are now required to explore and disclose this technique's full potential.

### Final Remarks

In this review, we have highlighted the most recent advances in immunotherapy for the management of CVD. A large amount of basic and translational atherosclerosis research has yielded important new insights in the disease's molecular pathogenesis. Similar to oncological applications and with the first immunotherapy on the clinical horizon, atherosclerosis immunoimaging will likely also become a reality, as outline in the Figure. The high costs associated with this approach may make it a treatment modality that may become available to high-risk patients who do not benefit from the current standard of care.

### Sources of Funding

This work was supported by National Institute of Health Grants R01HL118440, R01HL125703, P01HL131478, a Netherlands Organisation for Scientific Research Vidi (all to W.J.M. Mulder), as well as R01 EB009638, R01HL128056, R01HL135878, and P01HL131478 (to Z.A. Fayad)

### Disclosures

None.

### References

1. Mozaffarian D, Benjamin E, Go A, et al. Heart Disease and Stroke Statistics—update: a report from the American Heart Association. *Circulation*. 2016;135:e1–e324.
2. Lu H, Daugherty A. Atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2015;35:485–491. doi: 10.1161/ATVBAHA.115.305380.
3. Hedrick CC. Lymphocytes in atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2015;35:253–257. doi: 10.1161/ATVBAHA.114.305144.
4. Gadde S, Rayner KJ. Nanomedicine meets microRNA: current advances in RNA-based nanotherapies for atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2016;36:e73–e79. doi: 10.1161/ATVBAHA.116.307481.
5. Zhang H, de Aguiar Vallim TQ, Martel C; Early Career Committee. Translational and therapeutic approaches to the understanding and treatment of dyslipidemia. *Arterioscler Thromb Vasc Biol*. 2016;36:e56–e61. doi: 10.1161/ATVBAHA.116.307808.

6. Marrache S, Dhar S. Biodegradable synthetic high-density lipoprotein nanoparticles for atherosclerosis. *Proc Natl Acad Sci U S A*. 2013;110:9445–9450. doi: 10.1073/pnas.1301929110.
7. Rader DJ, Daugherty A. Translating molecular discoveries into new therapies for atherosclerosis. *Nature*. 2008;451:904–913. doi: 10.1038/nature06796.
8. Abe J, Berk BC. Novel mechanisms of endothelial mechanotransduction. *Arterioscler Thromb Vasc Biol*. 2014;34:2378–2386. doi: 10.1161/ATVBAHA.114.303428.
9. Dutta P, Courties G, Wei Y, et al. Myocardial infarction accelerates atherosclerosis. *Nature*. 2012;487:325–329. doi: 10.1038/nature11260.
10. Schrijvers DM, De Meyer GR, Kockx MM, Herman AG, Martinet W. Phagocytosis of apoptotic cells by macrophages is impaired in atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2005;25:1256–1261. doi: 10.1161/01.ATV.0000166517.18801.a7.
11. Vergallo R, Uemura S, Soeda T, Minami Y, Cho JM, Ong DS, Aguirre AD, Gao L, Biasucci LM, Crea F, Yu B, Lee H, Kim CJ, Jang IK. Prevalence and predictors of multiple coronary plaque ruptures: *in vivo* 3-vessel optical coherence tomography imaging study. *Arterioscler Thromb Vasc Biol*. 2016;36:2229–2238. doi: 10.1161/ATVBAHA.116.307891.
12. Randolph GJ. Mechanisms that regulate macrophage burden in atherosclerosis. *Circ Res*. 2014;114:1757–1771. doi: 10.1161/CIRCRESAHA.114.301174.
13. Moore KJ, Sheedy FJ, Fisher EA. Macrophages in atherosclerosis: a dynamic balance. *Nat Rev Immunol*. 2013;13:709–721. doi: 10.1038/nri3520.
14. Feig JE, Hewing B, Smith JD, Hazen SL, Fisher EA. High-density lipoprotein and atherosclerosis regression: evidence from preclinical and clinical studies. *Circ Res*. 2014;114:205–213. doi: 10.1161/CIRCRESAHA.114.300760.
15. Zou W, Wolchok JD, Chen L. PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: mechanisms, response biomarkers, and combinations. *Sci Transl Med*. 2016;8:328rv4. doi: 10.1126/scitranslmed.aad7118.
16. Yuan J, Hegde PS, Clynes R, et al. Novel technologies and emerging biomarkers for personalized cancer immunotherapy. *J Immunother Cancer*. 2016;4:3. doi: 10.1186/s40425-016-0107-3.
17. Belov V, Levine D, Belova E, et al. Pharmacological PET imaging performance with I-124 and Zr-89. *J Nucl Med*. 2014;55(suppl 1):590–590.
18. Mestel R. Cancer: imaging with antibodies. *Nature*. 2017;543:743–746. doi: 10.1038/543743a.
19. Tavaré R, Escuin-Ordinas H, Mok S, McCracken MN, Zettlitz KA, Salazar FB, Witte ON, Ribas A, Wu AM. An effective immuno-PET imaging method to monitor CD8-dependent responses to immunotherapy. *Cancer Res*. 2016;76:73–82. doi: 10.1158/0008-5472.CAN-15-1707.
20. Gebhart G, Lamberts LE, Wimana Z, et al. Molecular imaging as a tool to investigate heterogeneity of advanced HER2-positive breast cancer and to predict patient outcome under trastuzumab emtansine (T-DM1): the ZEPHIR trial. *Ann Oncol*. 2016;27:619–624. doi: 10.1093/annonc/mdv577.
21. Novartis official web site. <https://www.novartis.com/news/media-releases/novartis-phase-iii-study-shows-acz885-canakinumab-reduces-cardiovascular-risk>. Accessed July 7, 2017.
22. Varvel S, McConnell JP, Tsimikas S. Prevalence of elevated Lp(a) mass levels and patient thresholds in 532 359 patients in the United States. *Arterioscler Thromb Vasc Biol*. 2016;36:2239–2245. doi: 10.1161/ATVBAHA.116.308011.
23. Li J, Ley K. Lymphocyte migration into atherosclerotic plaque. *Arterioscler Thromb Vasc Biol*. 2015;35:40–49. doi: 10.1161/ATVBAHA.114.303227.
24. Xu L, Dai Perrard X, Perrard JL, Yang D, Xiao X, Teng BB, Simon SI, Ballantyne CM, Wu H. Foamy monocytes form early and contribute to nascent atherosclerosis in mice with hypercholesterolemia. *Arterioscler Thromb Vasc Biol*. 2015;35:1787–1797. doi: 10.1161/ATVBAHA.115.305609.
25. Dutta P, Nahrendorf M. Monocytes in myocardial infarction. *Arterioscler Thromb Vasc Biol*. 2015;35:1066–1070. doi: 10.1161/ATVBAHA.114.304652.
26. Hilgendorf I, Swirski FK, Robbins CS. Monocyte fate in atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2015;35:272–279. doi: 10.1161/ATVBAHA.114.303565.
27. Tsiantoulas D, Sage AP, Mallat Z, Binder CJ. Targeting B cells in atherosclerosis: closing the gap from bench to bedside. *Arterioscler Thromb Vasc Biol*. 2015;35:296–302. doi: 10.1161/ATVBAHA.114.303569.
28. Odrí Komazec I, Posod A, Schwienbacher M, Resch M, Pupp Peglow U, Kiechl S, Baumgartner D, Kiechl-Kohlendorfer U. Aortic elastic properties in preschool children born preterm. *Arterioscler Thromb Vasc Biol*. 2016;36:2268–2274. doi: 10.1161/ATVBAHA.116.308144.
29. Stary HC. Natural history and histological classification of atherosclerotic lesions: an update. *Arterioscler Thromb Vasc Biol*. 2000;20:1177–1178.
30. Puri R, Nissen SE, Shao M, Elshazly MB, Kataoka Y, Kapadia SR, Tuzcu EM, Nicholls SJ. Non-HDL cholesterol and triglycerides: implications for coronary atheroma progression and clinical events. *Arterioscler Thromb Vasc Biol*. 2016;36:2220–2228. doi: 10.1161/ATVBAHA.116.307601.
31. Varbo A, Nordestgaard BG. Remnant cholesterol and triglyceride-rich lipoproteins in atherosclerosis progression and cardiovascular disease. *Arterioscler Thromb Vasc Biol*. 2016;36:2133–2135. doi: 10.1161/ATVBAHA.116.308305.
32. Robinson JG, Ray K. Counterpoint: low-density lipoprotein cholesterol targets are not needed in lipid treatment guidelines. *Arterioscler Thromb Vasc Biol*. 2016;36:586–590. doi: 10.1161/ATVBAHA.116.306887.
33. Herder M, Arntzen KA, Johnsen SH, Eggen AE, Mathiesen EB. Long-term use of lipid-lowering drugs slows progression of carotid atherosclerosis: the Tromsø study 1994 to 2008. *Arterioscler Thromb Vasc Biol*. 2013;33:858–862. doi: 10.1161/ATVBAHA.112.300767.
34. Ridker PM. How common is residual inflammatory risk? *Circ Res*. 2017;120:617–619. doi: 10.1161/CIRCRESAHA.116.310527.
35. Ballantyne CM, Raichlen JS, Nicholls SJ, Erbel R, Tardif JC, Brener SJ, Cain VA, Nissen SE; ASTEROID Investigators. Effect of rosuvastatin therapy on coronary artery stenoses assessed by quantitative coronary angiography: a study to evaluate the effect of rosuvastatin on intravascular ultrasound-derived coronary atheroma burden. *Circulation*. 2008;117:2458–2466. doi: 10.1161/CIRCULATIONAHA.108.773747.
36. Lu H, Howatt D, Balakrishnan A, et al. Hypercholesterolemia induced by a PCSK9 gain-of-function mutation augments angiotensin II-induced abdominal aortic aneurysms in C57BL/6 mice. *Arterioscler Thromb Vasc Biol*. 2016;36:1753–1757.
37. Demers A, Samami S, Lauzier B, Des Rosiers C, Ngo Sock ET, Ong H, Mayer G. PCSK9 induces CD36 degradation and affects long-chain fatty acid uptake and triglyceride metabolism in adipocytes and in mouse liver. *Arterioscler Thromb Vasc Biol*. 2015;35:2517–2525. doi: 10.1161/ATVBAHA.115.306032.
38. Tavori H, Giunzioni I, Fazio S. PCSK9 inhibition to reduce cardiovascular disease risk: recent findings from the biology of PCSK9. *Curr Opin Endocrinol Diabetes Obes*. 2015;22:126–132. doi: 10.1097/MED.000000000000137.
39. Slomski A. Therapies that target PCSK9 effective at reducing LDL cholesterol. *JAMA*. 2017;317:2054–2054.
40. Ray KK, Landmesser U, Leiter LA, Kallend D, Dufour R, Karakas M, Hall T, Troquay RP, Turner T, Visseren FL, Wijngaard P, Wright RS, Kastelein JJ. Inclisiran in patients at high cardiovascular risk with elevated LDL cholesterol. *N Engl J Med*. 2017;376:1430–1440. doi: 10.1056/NEJMoa1615758.
41. Herder C, de Las Heras Gala T, Carstensen-Kirberg M, et al. Circulating levels of interleukin 1-receptor antagonist and risk of cardiovascular disease: meta-analysis of six population-based cohorts. *Arterioscler Thromb Vasc Biol*. 2017;37:1222–1227. doi: 10.1161/ATVBAHA.117.309307.
42. Ceneri N, Zhao L, Young BD, et al. Rac2 modulates atherosclerotic calcification by regulating macrophage interleukin-1 $\beta$  production. *Arterioscler Thromb Vasc Biol*. 2017;37:328–340. doi: 10.1161/ATVBAHA.116.308507.
43. Ridker PM. From C-reactive protein to interleukin-6 to interleukin-1: moving upstream to identify novel targets for atheroprotection. *Circ Res*. 2016;118:145–156. doi: 10.1161/CIRCRESAHA.115.306656.
44. Potteaux S, Gautier EL, Hutchison SB, van Rooijen N, Rader DJ, Thomas MJ, Sorci-Thomas MG, Randolph GJ. Suppressed monocyte recruitment drives macrophage removal from atherosclerotic plaques of ApoE $^{-/-}$  mice during disease regression. *J Clin Invest*. 2011;121:2025–2036. doi: 10.1172/JCI43802.
45. Feig JE, Shang Y, Rotllan N, Vengrenyuk Y, Wu C, Shamir R, Torra IP, Fernandez-Hernando C, Fisher EA, Garabedian MJ. Statins promote the regression of atherosclerosis via activation of the CCR7-dependent emigration pathway in macrophages. *PLoS One*. 2011;6:e28534. doi: 10.1371/journal.pone.0028534.
46. Banach M, Serban C, Sahebkar A, Mikhailidis DP, Ursoniu S, Ray KK, Rysz J, Toth PP, Muntner P, Mosteoru S, García-García HM, Hovingh GK, Kastelein JJ, Serruys PW; Lipid and Blood Pressure Meta-analysis Collaboration (LBPMC) Group. Impact of statin therapy on coronary plaque composition: a systematic review and meta-analysis of virtual histology intravascular ultrasound studies. *BMC Med*. 2015;13:229. doi: 10.1186/s12916-015-0459-4.
47. Duivenvoorden R, Tang J, Cormode DP, et al. A statin-loaded reconstituted high-density lipoprotein nanoparticle inhibits atherosclerotic plaque inflammation. *Nat Commun*. 2014;5:3065. doi: 10.1038/ncomms4065.
48. Getz GS, Reardon CA. Do the ApoE $^{-/-}$  and Ldlr $^{-/-}$  mice yield the same insight on atherogenesis? *Arterioscler Thromb Vasc Biol*. 2016;36:1734–1741. doi: 10.1161/ATVBAHA.116.306874.

49. Hoekstra M, Van Berkel TJ. Functionality of high-density lipoprotein as antiatherosclerotic therapeutic target. *Arterioscler Thromb Vasc Biol.* 2016;36:e87–e94. doi: 10.1161/ATVBAHA.116.308262.
50. Galvani S, Hla T. Quality versus quantity: making HDL great again. *Arterioscler Thromb Vasc Biol.* 2017;37:1018–1019. doi: 10.1161/ATVBAHA.117.309441.
51. Tang J, Lobatto M, Hassing L, et al. Inhibiting macrophage proliferation suppresses atherosclerotic plaque inflammation. *Sci Adv.* 2015;1:e1400223.
52. Kratochvil RM, Kubes P, Deniset JF. Monocyte conversion during inflammation and injury. *Arterioscler Thromb Vasc Biol.* 2017;37:35–42. doi: 10.1161/ATVBAHA.116.308198.
53. Dutta P, Hoyer FF, Sun Y, Iwamoto Y, Tricot B, Weissleder R, Magnani JL, Swirski FK, Nahrendorf M. E-selectin inhibition mitigates splenic HSC activation and myelopoiesis in hypercholesterolemic mice with myocardial infarction. *Arterioscler Thromb Vasc Biol.* 2016;36:1802–1808. doi: 10.1161/ATVBAHA.116.307519.
54. Sager HB, Dutta P, Dahlman JE, et al. RNAi targeting multiple cell adhesion molecules reduces immune cell recruitment and vascular inflammation after myocardial infarction. *Sci Transl Med.* 2016;8:342ra80. doi: 10.1126/scitranslmed.aaf1435.
55. Wynn TA, Vannella KM. Macrophages in tissue repair, regeneration, and fibrosis. *Immunity.* 2016;44:450–462. doi: 10.1016/j.immuni.2016.02.015.
56. Das A, Sinha M, Datta S, Abas M, Chaffee S, Sen CK, Roy S. Monocyte and macrophage plasticity in tissue repair and regeneration. *Am J Pathol.* 2015;185:2596–2606. doi: 10.1016/j.ajpath.2015.06.001.
57. Zheng D, Wang Y, Cao Q, Lee VW, Zheng G, Sun Y, Tan TK, Wang Y, Alexander SI, Harris DC. Transfused macrophages ameliorate pancreatic and renal injury in murine diabetes mellitus. *Nephron Exp Nephrol.* 2011;118:e87–e99. doi: 10.1159/000321034.
58. Colin S, Chinetti-Gbaguidi G, Staels B. Macrophage phenotypes in atherosclerosis. *Immunol Rev.* 2014;262:153–166. doi: 10.1111/imr.12218.
59. He H, Mack J, Güç E, et al. Perivascular macrophages limit permeability. *Arterioscler Thromb Vasc Biol.* 2016;36:2203–2212.
60. Mallat Z. Macrophages. *Arterioscler Thromb Vasc Biol.* 2014;34:2509–2519. doi: 10.1161/ATVBAHA.114.304794.
61. Fisher EA. Regression of atherosclerosis: the journey from the liver to the plaque and back. *Arterioscler Thromb Vasc Biol.* 2016;36:226–235. doi: 10.1161/ATVBAHA.115.301926.
62. Gordon S, Martinez FO. Alternative activation of macrophages: mechanism and functions. *Immunity.* 2010;32:593–604. doi: 10.1016/j.immuni.2010.05.007.
63. Van Dyken SJ, Locksley RM. Interleukin-4- and interleukin-13-mediated alternatively activated macrophages: roles in homeostasis and disease. *Annu Rev Immunol.* 2013;31:317–343. doi: 10.1146/annurev-immunol-032712-095906.
64. Gombozhapova A, Rogovskaya Y, Shurupov V, Rebenkova M, Kzhyshkowska J, Popov SV, Karpov RS, Ryabov V. Macrophage activation and polarization in post-infarction cardiac remodeling. *J Biomed Sci.* 2017;24:13. doi: 10.1186/s12929-017-0322-3.
65. Wynn T, Chawla A, Pollard J. Origins and hallmarks of macrophages: development, homeostasis, and disease. *Nature.* 2013;496:445.
66. Bosurgi L, Cao YG, Cabeza-Cabrero M, Tucci A, Hughes LD, Kong Y, Weinstein JS, Licon-Limon P, Schmid ET, Pelorosso F, Gagliani N, Craft JE, Flavell RA, Ghosh S, Rothlin CV. Macrophage function in tissue repair and remodeling requires IL-4 or IL-13 with apoptotic cells. *Science.* 2017;356:1072–1076. doi: 10.1126/science.aai8132.
67. Allen JE, Maizels RM. Diversity and dialogue in immunity to helminths. *Nat Rev Immunol.* 2011;11:375–388. doi: 10.1038/nri2992.
68. Shen SW, Lu Y, Li F, Shen ZH, Xu M, Yao WF, Feng YB, Yun JT, Wang YP, Ling W, Qi HJ, Tong DX. Potential long-term effects of previous schistosome infection may reduce the atherogenic index of plasma in Chinese men. *Int J Parasitol.* 2015;45:289–294. doi: 10.1016/j.ijpara.2015.01.001.
69. Magen E, Bychkov V, Ginovker A, Kashuba E. Chronic *Opisthorchis felineus* infection attenuates atherosclerosis—an autopsy study. *Int J Parasitol.* 2013;43:819–824. doi: 10.1016/j.ijpara.2013.04.008.
70. Knipper JA, Willenborg S, Brinckmann J, Bloch W, Maaß T, Wagener R, Krieg T, Sutherland T, Munitz A, Rothenberg ME, Niehoff A, Richardson R, Hammerschmidt M, Allen JE, Eming SA. Interleukin-4 receptor  $\alpha$  signaling in myeloid cells controls collagen fibril assembly in skin repair. *Immunity.* 2015;43:803–816. doi: 10.1016/j.immuni.2015.09.005.
71. Chen YC, Huang AL, Kyaw TS, Bobik A, Peter K. Atherosclerotic plaque rupture: identifying the straw that breaks the camel's back. *Arterioscler Thromb Vasc Biol.* 2016;36:e63–e72. doi: 10.1161/ATVBAHA.116.307993.
72. Calcagno C, Mulder WJ, Nahrendorf M, Fayad ZA. Systems biology and noninvasive imaging of atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2016;36:e1–e8. doi: 10.1161/ATVBAHA.115.306350.
73. Herbin O, Regelmann AG, Ramkhalawon B, Weinstein EG, Moore KJ, Alexandropoulos K. Monocyte adhesion and plaque recruitment during atherosclerosis development is regulated by the adapter protein Chat-H/SHEP1. *Arterioscler Thromb Vasc Biol.* 2016;36:1791–1801. doi: 10.1161/ATVBAHA.116.308014.
74. Curaj A, Wu Z, Fokong S, Liehn EA, Weber C, Burlacu A, Lammers T, van Zandvoort M, Kiessling F. Noninvasive molecular ultrasound monitoring of vessel healing after intravascular surgical procedures in a pre-clinical setup. *Arterioscler Thromb Vasc Biol.* 2015;35:1366–1373. doi: 10.1161/ATVBAHA.114.304857.
75. Derlin T, Thiele J, Weiber D, Thackeray JT, Püschel K, Wester HJ, Aguirre Dávila L, Larena-Avellaneda A, Daum G, Bengel FM, Schumacher U. Evaluation of  $^{68}\text{Ga}$ -glutamate carboxypeptidase II ligand positron emission tomography for clinical molecular imaging of atherosclerotic plaque neovascularization. *Arterioscler Thromb Vasc Biol.* 2016;36:2213–2219. doi: 10.1161/ATVBAHA.116.307701.
76. Tarkin JM, Joshi FR, Evans NR, et al. Detection of atherosclerotic inflammation by  $(^{68}\text{Ga})\text{-DOTATATE}$  PET compared to  $[(^{18}\text{F})\text{FDG}]$  PET imaging. *J Am Coll Cardiol.* 2017;69:1774–1791. doi: 10.1016/j.jacc.2017.01.060.
77. Blasi F, Oliveira BL, Rietz TA, Rotile NJ, Naha PC, Cormode DP, Izquierdo-Garcia D, Catana C, Caravan P. Multisite thrombus imaging and fibrin content estimation with a single whole-body PET scan in rats. *Arterioscler Thromb Vasc Biol.* 2015;35:2114–2121. doi: 10.1161/ATVBAHA.115.306055.
78. Fayad ZA, Mani V, Woodward M, Kallend D, Abt M, Burgess T, Fuster V, Ballantyne CM, Stein EA, Tardif JC, Rudd JH, Farkouh ME, Tawakol A; dal-PLAQUE Investigators. Safety and efficacy of dalcetrapib on atherosclerotic disease using novel non-invasive multimodality imaging (dal-PLAQUE): a randomised clinical trial. *Lancet.* 2011;378:1547–1559. doi: 10.1016/S0140-6736(11)61383-4.
79. Tarkin JM, Dweck MR, Evans NR, Takx RA, Brown AJ, Tawakol A, Fayad ZA, Rudd JH. Imaging atherosclerosis. *Circ Res.* 2016;118:750–769. doi: 10.1161/CIRCRESAHA.115.306247.
80. Tawakol A, Singh P, Mojena M, et al. HIF-1 $\alpha$  and PFKFB3 mediate a tight relationship between proinflammatory activation and anaerobic metabolism in atherosclerotic macrophages. *Arterioscler Thromb Vasc Biol.* 2015;35:1463–1471. doi: 10.1161/ATVBAHA.115.305551.
81. Bucerius J, Hyafil F, Verberne HJ, Slart RH, Lindner O, Sciacra R, Agostini D, Übleis C, Gimelli A, Hacker M; Cardiovascular Committee of the European Association of Nuclear Medicine (EANM). Position paper of the Cardiovascular Committee of the European Association of Nuclear Medicine (EANM) on PET imaging of atherosclerosis. *Eur J Nucl Med Mol Imaging.* 2016;43:780–792. doi: 10.1007/s00259-015-3259-3.
82. Tawakol A, Fayad ZA, Mogg R, Alon A, Klimas MT, Dansky H, Subramanian SS, Abdelbaky A, Rudd JH, Farkouh ME, Nunes IO, Beals CR, Shankar SS. Intensification of statin therapy results in a rapid reduction in atherosclerotic inflammation: results of a multicenter fluorodeoxyglucose-positron emission tomography/computed tomography feasibility study. *J Am Coll Cardiol.* 2013;62:909–917. doi: 10.1016/j.jacc.2013.04.066.
83. Joshi NV, Vesey AT, Williams MC, et al.  $^{18}\text{F}$ -fluoride positron emission tomography for identification of ruptured and high-risk coronary atherosclerotic plaques: a prospective clinical trial. *Lancet.* 2014;383:705–713. doi: 10.1016/S0140-6736(13)61754-7.
84. Honda A, Tahara N, Nitta Y, et al. Vascular inflammation evaluated by  $^{18}\text{F}$ -fluorodeoxyglucose-positron emission tomography/computed tomography is associated with endothelial dysfunction. *Arterioscler Thromb Vasc Biol.* 2016;36:1980–1988. doi: 10.1161/ATVBAHA.116.307293.
85. Robson P, Dweck M, Trivieri M, et al. Coronary artery PET/MR imaging. *JACC Cardiovasc Imaging.* 2017;2154:30976–30977.
86. Pedersen SF, Sandholt BV, Keller SH, Hansen AE, Clemmensen AE, Sillesen H, Højgaard L, Ripa RS, Kjær A.  $^{64}\text{Cu}$ -DOTATATE PET/MRI for detection of activated macrophages in carotid atherosclerotic plaques: studies in patients undergoing endarterectomy. *Arterioscler Thromb Vasc Biol.* 2015;35:1696–1703. doi: 10.1161/ATVBAHA.114.305067.

KEY WORDS: atherosclerosis ■ cardiovascular disease ■ imaging ■ immunotherapy ■ inflammation ■ lipids

# Arteriosclerosis, Thrombosis, and Vascular Biology



JOURNAL OF THE AMERICAN HEART ASSOCIATION

## Cardiovascular Immunotherapy and the Role of Imaging Eva Zupancic, Zahi A. Fayad and Willem J.M. Mulder

*Arterioscler Thromb Vasc Biol.* 2017;37:e167-e171

doi: 10.1161/ATVBAHA.117.309227

*Arteriosclerosis, Thrombosis, and Vascular Biology* is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231

Copyright © 2017 American Heart Association, Inc. All rights reserved.

Print ISSN: 1079-5642. Online ISSN: 1524-4636

The online version of this article, along with updated information and services, is located on the World Wide Web at:

<http://atvb.ahajournals.org/content/37/11/e167>

**Permissions:** Requests for permissions to reproduce figures, tables, or portions of articles originally published in *Arteriosclerosis, Thrombosis, and Vascular Biology* can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the [Permissions and Rights Question and Answer](#) document.

**Reprints:** Information about reprints can be found online at:  
<http://www.lww.com/reprints>

**Subscriptions:** Information about subscribing to *Arteriosclerosis, Thrombosis, and Vascular Biology* is online at:  
<http://atvb.ahajournals.org/subscriptions/>