

## A Role for Partial Endothelial–Mesenchymal Transitions in Angiogenesis?

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**Abstract**—The contribution of epithelial-to-mesenchymal transitions (EMT) in both developmental and pathological conditions has been widely recognized and studied. In a parallel process, governed by a similar set of signaling and transcription factors, endothelial-to-mesenchymal transitions (EndoMT) contribute to heart valve formation and the generation of cancer-associated fibroblasts. During angiogenic sprouting, endothelial cells express many of the same genes and break down basement membrane; however, they retain intercellular junctions and migrate as a connected train of cells rather than as individual cells. This has been termed a partial endothelial-to-mesenchymal transition. A key regulatory check-point determines whether cells undergo a full or a partial epithelial-to-mesenchymal transition/endothelial-to-mesenchymal transition; however, very little is known about how this switch is controlled. Here we discuss these developmental/pathological pathways, with a particular focus on their role in vascular biology. (*Arterioscler Thromb Vasc Biol.* 2015;35:303-308. DOI: 10.1161/ATVBAHA.114.303220.)

**Key Words:** angiogenesis ■ EMT ■ endothelial ■ transcription factor

Morphological changes in tissues are invariably associated with phenotypic changes in the cells that comprise them. Often these are limited to temporary changes in protein expression patterns, but more dramatic changes can also occur, during which cells undergo changes in transcriptional programs that lead to significant changes in morphology and function. One class of such changes is called the epithelial-to-mesenchymal transition (EMT), and variants of traditional EMT include endothelial-to-mesenchymal transition (EndoMT) and partial EMT/EndoMT. Our focus will be to highlight the distinctions among the subsets, with an emphasis on angiogenesis as a unique example of a partial EndoMT.

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### Endothelial-to-Mesenchymal Transitions

Endothelial cells (EC) have many epithelial characteristics, including strong apical–basal polarity, the ability to form tubes, and the potential to undergo a transition to a mesenchymal-like cell (EndoMT). There are many reasons, therefore, to think that this process is related to EMT and may thus share some of the same pathways and effectors, including the key transcription factors Snail, Slug, Twist, Zeb1, and Zeb2, which we describe in detail further. There have been several excellent

reviews published on EMT,<sup>1–4</sup> and so we focus on EndoMT, with reference to EMT where clear overlaps exist. During embryogenesis, subsets of EC in the developing heart undergo EndoMT, acquire mesenchymal markers, invade the surrounding tissue, and form the valves and septa of the adult heart,<sup>5</sup> a process that involves transforming growth factor- $\beta$  (TGF $\beta$ ), bone morphogenetic protein, and Notch signaling pathways.<sup>6,7</sup> These pathways converge on a complex network of transcription factors that includes HES, HEY1/2, Twist, and SOX9.<sup>8,9</sup> Pathologically, EndoMT can be reactivated in the adult heart and has been shown to contribute to cardiac fibrosis, a characteristic common to most forms of heart failure. Using lineage-tracing techniques, Kalluri's group demonstrated that 27% to 35% of fibroblasts present in fibrotic heart tissue were of EC origin, strongly suggesting a role for EndoMT in this process. Importantly, EndoMT was TGF $\beta$ 1-dependent, whereas bone morphogenetic protein-7 preserved the EC phenotype and consequently reduced fibrosis.<sup>10</sup> Interestingly, however, a more recent study suggests that the accumulation of cardiac fibroblasts is not because of an EndoMT, but rather, the cells derive from a previously unrecognized fibroblast population, itself derived from ECs during development.<sup>11</sup> EndoMT has also been implicated as a source of fibroblasts in hypertrophic cardiomyopathy,<sup>12</sup> diabetes mellitus–induced cardiac fibrosis,<sup>13</sup> and chronic pulmonary hypertension,<sup>14,15</sup> although these studies lacked definitive lineage-tracing analyses.

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## Nonstandard Abbreviations and Acronyms

<b>E-cadherin</b>	epithelial cadherin
<b>EC</b>	endothelial cells
<b>EMT</b>	epithelial-to-mesenchymal transition
<b>EndoMT</b>	endothelial-to-mesenchymal transition
<b>FGF</b>	fibroblast growth factor
<b>HGF</b>	hepatocyte growth factor
<b>MMP</b>	matrix metalloproteinase
<b>N-cadherin</b>	neural cadherin
<b>NICD</b>	notch intracellular domain
<b>TGF<math>\beta</math></b>	transforming growth factor $\beta$
<b>VEGF</b>	vascular endothelial growth factor

There is also evidence supporting a role for EndoMT during both acute and chronic kidney injury.<sup>16</sup> In 3 distinct mouse models of chronic kidney disease,  $\approx 30\%$  to  $50\%$  of fibroblasts coexpressed the EC marker CD31 along with markers of myofibroblasts and fibroblasts, including fibroblast-specific protein-1 and  $\alpha$ -smooth muscle actin. Lineage tracing experiments confirmed the EC origin of these cells.<sup>16</sup> More recent work has suggested that only  $\approx 10\%$  of the myofibroblasts present in kidney fibrosis derive from an EndoMT, whereas the remainder come from proliferation of local fibroblasts and differentiation from bone marrow cells.<sup>17</sup> Other studies, however, have suggested that these fibroblasts may be derived from pericytes.<sup>18</sup> It should be borne in mind, however, that these are all mouse studies and strain-specific differences are always a possibility. Other fibrotic diseases where EndoMT has been implicated as a source of fibroblasts/stromal cells include intestinal fibrosis<sup>19</sup> and scleroderma.<sup>20,21</sup>

In aggregate, these studies provide evidence that EndoMT likely provides a source of fibroblasts in both damaged heart and kidney (although the extent is unclear) and may function to facilitate tissue remodeling and fibrosis.

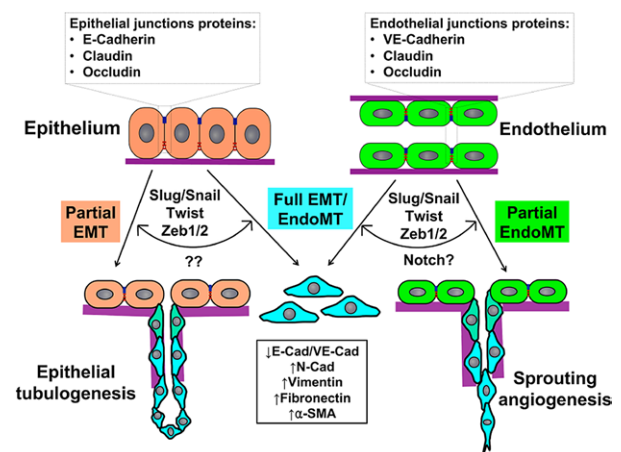
Finally, EndoMT also has a significant role to play in cancer. For example, Zeisberg et al,<sup>22</sup> using 2 different mouse models of cancer, demonstrated that EndoMT accounts for  $\leq 40\%$  of cancer-associated fibroblasts. A distinct population of fibroblasts coexpressed the EC marker CD31 along with either fibroblast-specific protein-1 or  $\alpha$ -smooth muscle actin. Use of transgenic mice with irreversibly tagged EC revealed strikingly similar results—unique populations of fibroblasts coexpressing endothelial and mesenchymal markers. These data suggest that EndoMT is a significant source of cancer-associated fibroblasts in tumors. Remarkably, it has also been demonstrated that Twist overexpression in head and neck cancer cells can drive them into an EC phenotype.<sup>23</sup>

### Angiogenesis: A Partial EndoMT

When epithelial cells commit to a mesenchymal phenotype, the event is designated as a complete EMT. Partial EMT is also possible, and this occurs when  $\geq 1$  of the key characteristics of complete EMT are not exhibited, such as loss of cell–cell contact. For example, during re-epithelialization of cutaneous wounds, keratinocytes undergo a series of changes reminiscent of EMT, including loss of polarity, rearrangement

of the actin cytoskeleton, alterations in cell–cell contacts, and breakdown of basement membrane; however, these cells retain some intercellular junctions and migrate as a cohesive cell sheet.<sup>24</sup> Similarly, during Madin–Darby canine kidney (MDCK) cell tubulogenesis, chains of epithelial cells migrate although again retaining intercellular junctions—a partial EMT driven by Slug activity.<sup>25</sup> Angiogenesis, the formation of new blood vessels from the preexisting vasculature, is essential during development and many normal physiological processes, but is also important in numerous pathological processes, including tumor growth. Interestingly, comparison of angiogenesis and EMT reveals several similarities. Among these, the tip cells that lead emerging sprouts lack apical–basal polarity, degrade both basement membrane and extracellular matrix and, by definition, are migratory. However, angiogenic EC do not usually separate from their neighbors, suggesting that angiogenesis may involve a partial EndoMT (Figure).<sup>26,27</sup>

Although much is known about the growth factors, receptors, and signaling pathways that govern angiogenesis, there is still much to learn about transcriptional changes that regulate each phase of angiogenesis, including sprouting. Our laboratory has recently published preliminary evidence demonstrating that the transcription factors Snail (*SNAIL* in human, *Snail* in mouse) and Slug (*SNAIL2/Snai2*) are indeed expressed and regulated by angiogenic EC during in vitro angiogenesis.<sup>26</sup> We demonstrated that inhibition of Snail or Slug expression results in a reduced ability of angiogenic EC to invade and migrate through multiple extracellular matrix environments. Importantly, lentiviral-mediated re-expression of membrane type-1 matrix metalloproteinase (MT1-MMP) rescued the inability of EC lacking Slug to



**Figure.** Complete vs partial epithelial-to-mesenchymal transition (EMT)/endothelial-to-mesenchymal transitions (EndoMT). Epithelial and endothelial cells comprise the quiescent epithelium and endothelium, respectively, and use junctional proteins to maintain connections. Once transcriptional reprogramming is initiated, an event led by the EMT/EndoMT transcription factors Slug, Snail, Twist, and Zeb1/2, the epithelial/endothelial cells lose apical–basal polarity, severe intercellular junctions, and become motile cells. However, the regulatory signal(s) that determine whether these cells undergo a complete EMT/EndoMT or partial EMT/EndoMT remains unclear. In the case of sprouting angiogenesis, the contact-dependent Notch signaling pathway may have a major role to play in this process. E-cadherin/E-cadherin indicates epithelial cadherin; N-Cad, neural cadherin;  $\alpha$ -SMA,  $\alpha$ -smooth muscle actin; and VE-cadherin/VE-Cad, vascular endothelial-cadherin.

migrate. This finding, therefore, suggests that membrane type-1-MMP is a critical downstream target of Slug during angiogenesis. Importantly, we and others have observed increased expression of Snail and Slug in the vasculature of colon, breast,<sup>28</sup> and ovarian carcinoma.<sup>29</sup> It is interesting to speculate that the same factors that drive epithelial cells toward a mesenchymal, prometastatic phenotype may also drive EC toward a proangiogenic phenotype, which is also associated with metastasis. Key factors here may include vascular endothelial growth factor (VEGF), TGF $\beta$ , bone morphogenetic proteins, hepatocyte growth factor, and Wnts. The permanently activated phenotype of tumor vasculature may well reflect the chronic activation of the EndoMT process, driven by persistently elevated VEGF and a hypoxic environment, leading to excessive sprouting and a failure to settle back into the mature, stable phenotype associated with nontumor EC. It is possible therefore that drugs that target EMT/EndoMT, potentially through these pathways, may be doubly effective in slowing metastatic spread of epithelial tumors. Finally, we have preliminary data suggesting that Slug deficiency in mice leads both to impaired developmental and to pathological angiogenesis (Welch-Reardon KM, Wu N, Hughes CCW, unpublished data, 2014). In aggregate, these data clearly point to a role for the Snail family of transcription factors during angiogenesis. Furthermore, the findings that cell–cell contact is retained and expression of vascular endothelial-cadherin is not reduced are reminiscent of the partial EMT seen during keratinocyte migration in wound closure and during mammary gland or kidney epithelial cell tubule formation.<sup>30</sup> We therefore think that angiogenic sprouting may represent a partial EndoMT.

### Signaling Pathways Governing EndoMT

EndoMT and EMT share many of the same regulators, with members of the TGF $\beta$  superfamily being arguably the most prominent players. TGF $\beta$  signaling through Smad-dependent and independent pathways leads to direct transcriptional regulation of multiple genes, including several EMT/EndoMT-inducing transcription factors.<sup>31</sup> Expression of these transcription factors subsequently drives loss of cell–cell adhesion by repression of epithelial/endothelial genes encoding junction proteins, regulation of cytoskeletal rearrangement, and increased expression and activity of both MT-MMPs and secreted MMPs.<sup>32</sup> Moreover, during EndoMT, upregulation of EC Slug by TGF $\beta$  and other growth factors results in increased migration and invasion into extracellular matrices of diverse composition, and this is due in part to the indirect activation of membrane type-1-MMP, MMP-2, and MMP-9.<sup>26</sup> Interestingly, nuclear Smads form multiprotein complexes with EMT/EndoMT-transcription factors, including Snail, Zeb1, and Zeb2, resulting in suppression or activation of promoters of epithelial (E-cadherin, Occludin, ZO-1) or mesenchymal (Vimentin, N-cadherin) genes, respectively.<sup>4</sup> TGF $\beta$  can also activate Smad-independent pathways, such as MAPK/ERK/JNK (mitogen-activated protein kinases/extracellular signal-regulated kinases/c-Jun N-terminal kinases), all of which are implicated in EndoMT.<sup>31,33,34</sup> Finally, a recent study has shown a requirement for PKC $\delta$  (protein kinase C) and c-Abl in mediating TGF $\beta$ -induced EndoMT in mouse pulmonary EC.<sup>35</sup>

Aside from TGF $\beta$ , several other signaling pathways associated with EMT have also been reported to regulate EndoMT. The relationship between canonical Wnt signaling and the onset of EMT and metastasis is well established in many cancer models. In human prostate cancer, the expression and nuclear activity of  $\beta$ -catenin correlates with the level of hypoxia-induced factor 1 alpha and hypoxia-induced factor 1 alpha–induced EMT.<sup>36</sup> The degree of hypoxia-induced EMT can also be enhanced by Wnt3a-induced activation of  $\beta$ -catenin in hepatic carcinoma.<sup>37</sup> Furthermore, it has been demonstrated that canonical Wnt signaling stabilizes Slug expression through regulating glycogen synthase kinase 3- $\beta$  phosphorylation and  $\beta$ Trcp-1-mediated ubiquitination, thereby inducing EMT in triple-negative breast cancer.<sup>38</sup> In contrast, the understanding of canonical Wnt signaling in EndoMT was mostly limited to developmental processes until recently. In an experimentally induced myocardial infarction model, Aisagbonhi et al, using lineage-tracing experiments, demonstrated that the canonical Wnt pathway is transiently activated in ECs, and this in turn leads to EndoMT.<sup>39</sup> More recently, the effect of Wnt7a and its antagonist Dkk-1 on EndoMT during arteriosclerosis was explored. In contrast to its effect on myofibroblasts, the activation of Wnt signaling through Wnt7b expression preserves the phenotype of ECs, whereas the expression of Dkk-1 promotes EndoMT.<sup>40</sup>

Fibroblast growth factor (FGF) has been proposed as a gatekeeper of partial EndoMTs through its regulation of the *let-7* miRNA, which normally acts to suppress TGF $\beta$ -induced EndoMT.<sup>41</sup> When FGF signaling is reduced in a murine model of transplant arteriopathy, in this case by inflammatory signals that downregulate the FGF receptor, *let-7*-mediated suppression of TGF $\beta$  signaling is relieved and EC undergo an EndoMT, leading to intimal fibrosis. Given that the sprouty genes, which regulate FGF receptor signaling, have previously been implicated in the regulation of EC sprouting,<sup>42</sup> it is tempting to speculate that they may be acting as a rheostat to fine-tune FGF signaling<sup>43</sup> and thereby control whether EC undergo a partial or full EndoMT in response to TGF $\beta$  signaling.

Notch activation is a well-known regulator of angiogenesis<sup>44–47</sup> and is linked to both EMT and EndoMT events. The cleavage and nuclear translocation of the Notch intracellular domain can induce transcriptional alterations and hence a series of morphological and functional changes related to a mesenchymal transition.<sup>48</sup> Notch can suppress (or activate) gene expression directly or through upregulation of Snail and Slug in both epithelial cells and EC and, thus, initiate EMT and EndoMT in both developmental and pathological conditions.<sup>49–51</sup> Notch ligands can also be induced by TGF $\beta$  signaling to activate Notch receptors<sup>52</sup> and enhance EMT synergistically.<sup>53</sup> Blockage of either Jagged-1, or its downstream signaling target Hey-1, can attenuate TGF $\beta$ -induced EMT in mammary gland, kidney tubule, and epidermal epithelial cells.<sup>50,54</sup> Notch and VEGF are both induced in the hypoxic tumor environment, and they work together to drive metastasis. On the one hand, interaction of Notch and hypoxia-induced factor pathways leads to increased stemness of cancer cells, self-renewal ability, and a complete EMT.<sup>50,55</sup> On the other hand, hypoxia-dependent induction of VEGF expression



augments tumor angiogenesis, which provides increased opportunities for tumor cell intravasation. Finally, the cross-talk between Notch and VEGF pathways in the context of hypoxic tumors also promotes partial EndoMT in angiogenic tumor EC, leading to the formation of unstable, leaky vessels.<sup>52</sup> Altered vessel integrity and permeability correlates with enhanced tumor cell dissemination to distant sites.<sup>56</sup>

Notch-mediated EMT/EndoMT is unusual, and somewhat paradoxical, as it is contact-dependent. Importantly, the ability of cells to retain intercellular adhesion complexes although migrating as a group is crucial to tubulogenesis. As described earlier, processes involving tubulogenesis, such as angiogenesis and kidney tubule formation, both require a partial EMT/EndoMT, during which the participating cells temporarily lose apical-basal polarity and gain migratory capacity, but never fully acquire all mesenchymal phenotypes, nor completely lose cellular adhesion. Although other signaling pathways, such as TGF $\beta$ , hepatocyte growth factor, and FGF, are capable of promoting this process, it is intriguing to speculate that Notch activation, perhaps in conjunction with sprouty, is a crucial determinant of a partial versus full EMT/EndoMT.

Aside from the major signaling pathways discussed earlier, miRNA, epigenetic regulation, and histone modification have also recently emerged as regulators of EMT<sup>30,32</sup> and may also have a role in EndoMT.<sup>41</sup> These alterations control the expression level of the Snail/Slug, ZEB, and Twist families of transcription factors, and these in turn feed back to affect either the expression of the miRNA or histone-modifying enzymes, or their activity.<sup>30,32</sup> Clearly, the relationship(s) between the master regulators governing EMT/EndoMT are extremely complex.<sup>4,31</sup>

### Transcription Factor Interactions Governing EndoMT

Snail (*SNAI1* in human, *Snai1* in mouse), Slug (*SNAI2/Snai2*), Zeb1 (*ZEB1/Zeb1*), Zeb2 (*ZEB2/Zeb2*), and Twist (*TWIST1/Twist1*) have been identified as the key transcriptional regulators of EMTs and EndoMTs. A shared function of these proteins is their ability to repress the transcription of epithelial-cadherin (E-cadherin); however, numerous studies have demonstrated that they have overlapping but nonredundant roles in EMT and tumor progression. In human carcinomas, it is generally accepted that Snail plays a major role in inducing EMT, whereas Zeb1/2 and Twist are mainly involved in maintaining the invasive mesenchymal phenotype.<sup>32</sup> However, our recent study on EndoMT suggests that at least in the case of sprouting angiogenesis, Slug is the primary initiator of this process, whereas the induction of Snail occurs at a much later time.<sup>26</sup> It is therefore unclear if each of these transcription factors has a distinct and specific role during EMT/EndoMT or if they rather act in symphony to promote a mesenchymal phenotype. Accumulating evidence from studies observing their expression patterns and their ability to regulate each other has begun to reveal a nonlinear map that suggests that these transcription factors mostly act in concert. For example, Snail can upregulate Zeb1 and Zeb2 in oral squamous carcinoma and, at the same time, negatively regulate its own expression through direct promoter binding.<sup>57,58</sup> Moreover, Slug indirectly upregulates Snail through epithelial growth factor or hepatocyte growth factor signaling, thereby promoting mammary gland

branching morphogenesis.<sup>59</sup> Slug can also activate Zeb1 and its own expression through direct transcriptional regulation.<sup>60,61</sup> In addition, many have shown that Twist1 can regulate the expression level of Snail and Slug by either directly influencing transcription<sup>62,63</sup> or through post-translational regulation via the NF- $\kappa$ B (nuclear factor kappa-light-chain-enhancer of activated B cells)/glycogen synthase kinase 3- $\beta$  axis.<sup>64</sup>

### Dynamic Functions of EndoMT Transcriptional Regulators

The master regulators of EMT mediate repression of E-cadherin expression, and this is often described as the hallmark of EMT. However, several recent studies show that in both in vitro and in vivo models, EMT master regulators can induce EMT/EndoMT-like phenotypes in cells without complete loss of membrane E-cadherin—a partial EMT. Similarly, we observed that overexpression of Slug in EC promotes EC sprouting, a process suggestive of a partial EndoMT, without altering the mRNA levels or surface expression of vascular endothelial-cadherin, the EC equivalent of E-cadherin.<sup>26</sup> Interestingly, Leroy et al others have shown that Slug upregulation prevents apoptosis and promotes cell proliferation through p53,<sup>25</sup> 2 processes associated with angiogenic sprouting. The induction of Twist alone, perhaps surprisingly, is sufficient to induce single cell dissemination/local invasion without the loss of epithelial identity, and moreover, E-cadherin expression is required for this process.<sup>65</sup> Conversely, the deletion of E-cadherin alone is not sufficient to induce EMT. Indeed, in the absence of E-cadherin, and despite a reduction in multiple classes of junction proteins, epithelial cells are still able to invade an extracellular matrix as a chain rather than single cells.<sup>65</sup> Collectively, these data suggest that master regulators of EMT and EndoMT serve more functions than simply acting as repressors of the epithelial/endothelial phenotype.

### Remaining Questions and Perspectives

Our knowledge of the mechanisms underlying EMT and EndoMT is rapidly advancing; however, there are still several critical questions that have to be answered, including the following:

- Are Snail, Slug, Twist, Zeb1, and Zeb2 all required for an EndoMT?
- Do these genes work sequentially, in parallel or in feedback loops?
- What regulates expression of EndoMT-promoting transcription factors?
- What are the target genes for EndoMT transcription factors?
- What controls whether cells undergo a full or partial EndoMT?
- How is vascular endothelial-cadherin protected from downregulation in partial EndoMT?
- Are there fundamental differences between EMT and EndoMT or are the basic mechanisms identical?
- Do the same factors that promote EMT in cancer promote EndoMT in angiogenic EC?
- How does EndoMT contribute to progression of diseases, such as cancer, arteriosclerosis, and fibrosis?
- Are there fundamental differences between pathological EndoMT and developmental EndoMT?

- Can EndoMT be targeted therapeutically in cancer and other diseases involving pathological angiogenesis?

Answers to these questions have the potential to fundamentally affect how we target pathological angiogenesis.

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### Disclosures

None.

### References

- Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *J Clin Invest*. 2009;119:1420–1428. doi: 10.1172/JCI39104.
- Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. *Cell*. 2009;139:871–890. doi: 10.1016/j.cell.2009.11.007.
- Tsai JH, Yang J. Epithelial-mesenchymal plasticity in carcinoma metastasis. *Genes Dev*. 2013;27:2192–2206. doi: 10.1101/gad.225334.113.
- Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol*. 2014;15:178–196. doi: 10.1038/nrm3758.
- Armstrong EJ, Bischoff J. Heart valve development: endothelial cell signaling and differentiation. *Circ Res*. 2004;95:459–470. doi: 10.1161/01.RES.0000141146.95728.da.
- Nakajima Y, Yamagishi T, Hokari S, Nakamura H. Mechanisms involved in valvuloseptal endocardial cushion formation in early cardiogenesis: roles of transforming growth factor (TGF)-beta and bone morphogenetic protein (BMP). *Anat Rec*. 2000;258:119–127.
- Timmerman LA, Grego-Bessa J, Raya A, Bertran E, Perez-Pomares JM, Diez J, Aranda S, Palomo S, McCormick F, Izpisua-Belmonte JC, de la Pompa JL. Notch promotes epithelial-mesenchymal transition during cardiac development and oncogenic transformation. *Genes Dev*. 2004;18:99–115. doi: 10.1101/gad.276304.
- Fischer A, Gessler M. Delta-Notch--and then? Protein interactions and proposed modes of repression by Hes and Hey bHLH factors. *Nucleic Acids Res*. 2007;35:4583–4596. doi: 10.1093/nar/gkm477.
- Wrigg EE, Yutzy KE. Conserved transcriptional regulatory mechanisms in aortic valve development and disease. *Arterioscler Thromb Vasc Biol*. 2014;34:737–741. doi: 10.1161/ATVBAHA.113.302071.
- Zeisberg EM, Tarnavski O, Zeisberg M, Dorfman AL, McMullen JR, Gustafsson E, Chandraker A, Yuan X, Pu WT, Roberts AB, Neilson EG, Sayegh MH, Izumo S, Kalluri R. Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat Med*. 2007;13:952–961. doi: 10.1038/nm1613.
- Moore-Morris T, Guimarães-Cambo A, Banerjee I, et al. Resident fibroblast lineages mediate pressure overload-induced cardiac fibrosis. *J Clin Invest*. 2014;124:2921–2934. doi: 10.1172/JCI74783.
- Teekakirikul P, Eminaga S, Toka O, et al. Cardiac fibrosis in mice with hypertrophic cardiomyopathy is mediated by non-myocyte proliferation and requires Tgf-β. *J Clin Invest*. 2010;120:3520–3529. doi: 10.1172/JCI42028.
- Widiantoro B, Emoto N, Nakayama K, Anggrahini DW, Adiarto S, Iwasa N, Yagi K, Miyagawa K, Rikitake Y, Suzuki T, Kisanuki YY, Yanagisawa M, Hirata K. Endothelial cell-derived endothelin-1 promotes cardiac fibrosis in diabetic hearts through stimulation of endothelial-to-mesenchymal transition. *Circulation*. 2010;121:2407–2418. doi: 10.1161/CIRCULATIONAHA.110.938217.
- Arciniegas E, Frid MG, Douglas IS, Stenmark KR. Perspectives on endothelial-to-mesenchymal transition: potential contribution to vascular remodeling in chronic pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol*. 2007;293:L1–L8. doi: 10.1152/ajplung.00378.2006.
- Zhu P, Huang L, Ge X, Yan F, Wu R, Ao Q. Transdifferentiation of pulmonary arteriolar endothelial cells into smooth muscle-like cells regulated by myocardium involved in hypoxia-induced pulmonary vascular remodeling. *Int J Exp Pathol*. 2006;87:463–474. doi: 10.1111/j.1365-2613.2006.00503.x.
- Zeisberg EM, Potenta SE, Sugimoto H, Zeisberg M, Kalluri R. Fibroblasts in kidney fibrosis emerge via endothelial-to-mesenchymal transition. *J Am Soc Nephrol*. 2008;19:2282–2287. doi: 10.1681/ASN.2008050513.
- LeBleu VS, Taduri G, O'Connell J, Teng Y, Cooke VG, Woda C, Sugimoto H, Kalluri R. Origin and function of myofibroblasts in kidney fibrosis. *Nat Med*. 2013;19:1047–1053. doi: 10.1038/nm.3218.
- Schrumpf C, Xin C, Campanholle G, Gill SE, Stallcup W, Lin SL, Davis GE, Gharib SA, Humphreys BD, Duffield JS. Pericyte TIMP3 and ADAMTS1 modulate vascular stability after kidney injury. *J Am Soc Nephrol*. 2012;23:868–883. doi: 10.1681/ASN.2011080851.
- Rieder F, Kessler SP, West GA, Bhilocha S, de la Motte C, Sadler TM, Gopalan B, Stylianou E, Fiocchi C. Inflammation-induced endothelial-to-mesenchymal transition: a novel mechanism of intestinal fibrosis. *Am J Pathol*. 2011;179:2660–2673. doi: 10.1016/j.ajpath.2011.07.042.
- Jimenez SA. Role of endothelial to mesenchymal transition in the pathogenesis of the vascular alterations in systemic sclerosis. *ISRN Rheumatol*. 2013;2013:835948. doi: 10.1155/2013/835948.
- Leach HG, Chrobak I, Han R, Trojanowska M. Endothelial cells recruit macrophages and contribute to a fibrotic milieu in bleomycin lung injury. *Am J Respir Cell Mol Biol*. 2013;49:1093–1101. doi: 10.1165/rmb.2013-0152OC.
- Zeisberg EM, Potenta S, Xie L, Zeisberg M, Kalluri R. Discovery of endothelial to mesenchymal transition as a source for carcinoma-associated fibroblasts. *Cancer Res*. 2007;67:10123–10128. doi: 10.1158/0008-5472.CAN-07-3127.
- Chen HF, Huang CH, Liu CJ, Hung JJ, Hsu CC, Teng SC, Wu KJ. Twist1 induces endothelial differentiation of tumour cells through the Jagged1-KLF4 axis. *Nat Commun*. 2014;5:4697. doi: 10.1038/ncomms5697.
- Savagner P, Kusewitt DF, Carver EA, Magnino F, Choi C, Gridley T, Hudson LG. Developmental transcription factor slug is required for effective re-epithelialization by adult keratinocytes. *J Cell Physiol*. 2005;202:858–866. doi: 10.1002/jcp.20188.
- Leroy P, Mostov KE. Slug is required for cell survival during partial epithelial-mesenchymal transition of HGF-induced tubulogenesis. *Mol Biol Cell*. 2007;18:1943–1952. doi: 10.1091/mbc.E06-09-0823.
- Welch-Reardon KM, Ehsan SM, Wang K, Wu N, Newman AC, Romero-Lopez M, Fong AH, George SC, Edwards RA, Hughes CC. Angiogenic sprouting is regulated by endothelial cell expression of Slug. *J Cell Sci*. 2014;127(Pt 9):2017–2028. doi: 10.1242/jcs.143420.
- Potenta S, Zeisberg E, Kalluri R. The role of endothelial-to-mesenchymal transition in cancer progression. *Br J Cancer*. 2008;99:1375–1379. doi: 10.1038/sj.bjc.6604662.
- Parker BS, Argani P, Cook BP, Liangfeng H, Chartrand SD, Zhang M, Saha S, Bardelli A, Jiang Y, St Martin TB, Nacht M, Teicher BA, Klinger KW, Sukumar S, Madden SL. Alterations in vascular gene expression in invasive breast carcinoma. *Cancer Res*. 2004;64:7857–7866. doi: 10.1158/0008-5472.CAN-04-1976.
- Lu C, Bonome T, Li Y, Kamat AA, Han LY, Schmandt R, Coleman RL, Gershenson DM, Jaffe RB, Birrer MJ, Sood AK. Gene alterations identified by expression profiling in tumor-associated endothelial cells from invasive ovarian carcinoma. *Cancer Res*. 2007;67:1757–1768. doi: 10.1158/0008-5472.CAN-06-3700.
- Yang J, Weinberg RA. Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. *Dev Cell*. 2008;14:818–829. doi: 10.1016/j.devcel.2008.05.009.
- Heldin CH, Vanlandewijck M, Moustakas A. Regulation of EMT by TGFβ in cancer. *FEBS Lett*. 2012;586:1959–1970. doi: 10.1016/j.febslet.2012.02.037.
- Peinado H, Olmeda D, Cano A. Snail, Zeb and bHLH factors in tumour progression: an alliance against the epithelial phenotype? *Nat Rev Cancer*. 2007;7:415–428. doi: 10.1038/nrc2131.
- Medici D, Potenta S, Kalluri R. Transforming growth factor-β2 promotes Snail-mediated endothelial-mesenchymal transition through convergence of Smad-dependent and Smad-independent signalling. *Biochem J*. 2011;437:515–520. doi: 10.1042/BJ20101500.
- van Meeteren LA, ten Dijke P. Regulation of endothelial cell plasticity by TGF-β. *Cell Tissue Res*. 2012;347:177–186. doi: 10.1007/s00441-011-1222-6.

35. Li Z, Jimenez SA. Protein kinase C $\delta$  and c-Abl kinase are required for transforming growth factor  $\beta$  induction of endothelial-mesenchymal transition *in vitro*. *Arthritis Rheum*. 2011;63:2473–2483. doi: 10.1002/art.30317.
36. Jiang YG, Luo Y, He DL, Li X, Zhang LL, Peng T, Li MC, Lin YH. Role of Wnt/ $\beta$ -catenin signaling pathway in epithelial-mesenchymal transition of human prostate cancer induced by hypoxia-inducible factor-1 $\alpha$ . *Int J Urol*. 2007;14:1034–1039. doi: 10.1111/j.1442-2042.2007.01866.x.
37. Zhang Q, Bai X, Chen W, Ma T, Hu Q, Liang C, Xie S, Chen C, Hu L, Xu S, Liang T. Wnt/ $\beta$ -catenin signaling enhances hypoxia-induced epithelial-mesenchymal transition in hepatocellular carcinoma via crosstalk with hif-1 $\alpha$  signaling. *Carcinogenesis*. 2013;34:962–973. doi: 10.1093/carcin/bgt027.
38. Wu ZQ, Li XY, Hu CY, Ford M, Kleer CG, Weiss SJ. Canonical Wnt signaling regulates Slug activity and links epithelial-mesenchymal transition with epigenetic Breast Cancer 1, Early Onset (BRCA1) repression. *Proc Natl Acad Sci U S A*. 2012;109:16654–16659. doi: 10.1073/pnas.1205822109.
39. Aisagbonhi O, Rai M, Ryzhov S, Atria N, Feoktistov I, Hatzopoulos AK. Experimental myocardial infarction triggers canonical Wnt signaling and endothelial-to-mesenchymal transition. *Dis Model Mech*. 2011;4:469–483. doi: 10.1242/dmm.006510.
40. Cheng SL, Shao JS, Behrmann A, Krchma K, Towler DA. Dkk1 and MSX2-Wnt7b signaling reciprocally regulate the endothelial-mesenchymal transition in aortic endothelial cells. *Arterioscler Thromb Vasc Biol*. 2013;33:1679–1689. doi: 10.1161/ATVBAHA.113.300647.
41. Chen PY, Qin L, Barnes C, Charisse K, Yi T, Zhang X, Ali R, Medina PP, Yu J, Slack FJ, Anderson DG, Kotelianski V, Wang F, Tellides G, Simons M. FGF regulates TGF- $\beta$  signaling and endothelial-to-mesenchymal transition via control of let-7 miRNA expression. *Cell Rep*. 2012;2:1684–1696. doi: 10.1016/j.celrep.2012.10.021.
42. Lee SH, Schloss DJ, Jarvis L, Krasnow MA, Swain JL. Inhibition of angiogenesis by a mouse sprout protein. *J Biol Chem*. 2001;276:4128–4133. doi: 10.1074/jbc.M006922200.
43. Impagnatiello MA, Weitzer S, Gannon G, Compagni A, Cotten M, Christofori G. Mammalian sprouty-1 and -2 are membrane-anchored phosphoprotein inhibitors of growth factor signaling in endothelial cells. *J Cell Biol*. 2001;152:1087–1098.
44. Taylor KL, Henderson AM, Hughes CC. Notch activation during endothelial cell network formation *in vitro* targets the basic HLH transcription factor HESR-1 and downregulates VEGFR-2/KDR expression. *Microvasc Res*. 2002;64:372–383.
45. Sainson RC, Aoto J, Nakatsu MN, Holderfield M, Conn E, Koller E, Hughes CC. Cell-autonomous notch signaling regulates endothelial cell branching and proliferation during vascular tubulogenesis. *FASEB J*. 2005;19:1027–1029. doi: 10.1096/fj.04-3172fje.
46. Siekmann AF, Lawson ND. Notch signalling limits angiogenic cell behaviour in developing zebrafish arteries. *Nature*. 2007;445:781–784. doi: 10.1038/nature05577.
47. Hellström M, Phng LK, Hofmann JJ, et al. Dll4 signalling through Notch1 regulates formation of tip cells during angiogenesis. *Nature*. 2007;445:776–780. doi: 10.1038/nature05571.
48. Espinoza I, Miele L. Deadly crosstalk: Notch signaling at the intersection of EMT and cancer stem cells. *Cancer Lett*. 2013;341:41–45. doi: 10.1016/j.canlet.2013.08.027.
49. Becker KF, Rosivatz E, Blechschmidt K, Kremmer E, Sarbia M, Höfler H. Analysis of the E-cadherin repressor Snail in primary human cancers. *Cells Tissues Organs*. 2007;185:204–212. doi: 10.1159/000101321.
50. Sahlgren C, Gustafsson MV, Jin S, Poellinger L, Lendahl U. Notch signaling mediates hypoxia-induced tumor cell migration and invasion. *Proc Natl Acad Sci U S A*. 2008;105:6392–6397. doi: 10.1073/pnas.0802047105.
51. Niessen K, Fu Y, Chang L, Hoodless PA, McFadden D, Karsan A. Slug is a direct Notch target required for initiation of cardiac cushion cellularization. *J Cell Biol*. 2008;182:315–325. doi: 10.1083/jcb.200710067.
52. Holderfield MT, Hughes CC. Crosstalk between vascular endothelial growth factor, notch, and transforming growth factor-beta in vascular morphogenesis. *Circ Res*. 2008;102:637–652. doi: 10.1161/CIRCRESAHA.107.167171.
53. Niimi H, Pardali K, Vanlandewijck M, Heldin CH, Moustakas A. Notch signaling is necessary for epithelial growth arrest by TGF-beta. *J Cell Biol*. 2007;176:695–707. doi: 10.1083/jcb.200612129.
54. Zavadil J, Cermak L, Soto-Nieves N, Böttinger EP. Integration of TGF-beta/Smad and Jagged1/Notch signalling in epithelial-to-mesenchymal transition. *EMBO J*. 2004;23:1155–1165. doi: 10.1038/sj.emboj.7600069.
55. Gustafsson MV, Zheng X, Pereira T, Gradin K, Jin S, Lundkvist J, Ruas JL, Poellinger L, Lendahl U, Bondesson M. Hypoxia requires notch signaling to maintain the undifferentiated cell state. *Dev Cell*. 2005;9:617–628. doi: 10.1016/j.devcel.2005.09.010.
56. Sullivan R, Graham CH. Hypoxia-driven selection of the metastatic phenotype. *Cancer Metastasis Rev*. 2007;26:319–331. doi: 10.1007/s10555-007-9062-2.
57. Takkunen M, Grenman R, Hukkanen M, Korhonen M, García de Herreros A, Virtanen I. Snail-dependent and -independent epithelial-mesenchymal transition in oral squamous carcinoma cells. *J Histochem Cytochem*. 2006;54:1263–1275. doi: 10.1369/jhc.6A6958.2006.
58. Peiró S, Escrivà M, Puig I, Barberà MJ, Dave N, Herranz N, Larriba MJ, Takkunen M, Francí C, Muñoz A, Virtanen I, Baulida J, García de Herreros A. Snail1 transcriptional repressor binds to its own promoter and controls its expression. *Nucleic Acids Res*. 2006;34:2077–2084. doi: 10.1093/nar/gkl141.
59. Lee K, Gjorevski N, Boghaert E, Radisky DC, Nelson CM, Snail1, Snail2, and E47 promote mammary epithelial branching morphogenesis. *EMBO J*. 2011;30:2662–2674. doi: 10.1038/emboj.2011.159.
60. Wels C, Joshi S, Koefinger P, Bergler H, Schaidler H. Transcriptional activation of ZEB1 by Slug leads to cooperative regulation of the epithelial-mesenchymal transition-like phenotype in melanoma. *J Invest Dermatol*. 2011;131:1877–1885. doi: 10.1038/jid.2011.142.
61. Sakai D, Suzuki T, Osumi N, Wakamatsu Y. Cooperative action of Sox9, Snail2 and PKA signaling in early neural crest development. *Development*. 2006;133:1323–1333. doi: 10.1242/dev.02297.
62. Yu W, Zhang Y, Ruest LB, Svoboda KK. Analysis of Snail1 function and regulation by Twist1 in palatal fusion. *Front Physiol*. 2013;4:12. doi: 10.3389/fphys.2013.00012.
63. Casas E, Kim J, Bendesky A, Ohno-Machado L, Wolfe CJ, Yang J. Snail2 is an essential mediator of Twist1-induced epithelial mesenchymal transition and metastasis. *Cancer Res*. 2011;71:245–254. doi: 10.1158/0008-5472.CAN-10-2330.
64. Lander N, Nasr T, Ochoa SD, Nordin K, Prasad MS, Labonne C. Interactions between Twist and other core epithelial-mesenchymal transition factors are controlled by GSK3-mediated phosphorylation. *Nat Commun*. 2013;4:1542. doi: 10.1038/ncomms2543.
65. Shamir ER, Pappalardo E, Jorgens DM, Coutinho K, Tsai WT, Aziz K, Auer M, Tran PT, Bader JS, Ewald AJ. Twist1-induced dissemination preserves epithelial identity and requires E-cadherin. *J Cell Biol*. 2014;204:839–856. doi: 10.1083/jcb.201306088.

## Significance

As a single cell multiplies and differentiates to generate a fully developed multicellular organism, daughter cells often undergo phenotypic changes that can be either permanent or temporary. One such change is termed an epithelial-to-mesenchymal transition, and this has been widely studied in both developmental and pathological conditions. It contributes to gastrulation and neural crest formation during development, and metastasis of epithelial tumors is also thought to involve an epithelial-to-mesenchymal transition. In a somewhat similar process, governed by an overlapping set of signaling and transcription factors, endothelial-to-mesenchymal transitions contribute to heart valve formation, the generation of cancer-associated fibroblasts, and the activated endothelial cells that drive angiogenic sprouting. A key regulatory check point determines whether cells undergo a full endothelial-to-mesenchymal transition (heart valve development) or a partial endothelial-to-mesenchymal transition (angiogenesis); however, little is known about how this switch is controlled. Here we discuss these developmental/pathological pathways, with a particular focus on their role in vascular biology.

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