Peroxisome Proliferator–Activated Receptor δ Agonist GW1516 Attenuates Diet-Induced Aortic Inflammation, Insulin Resistance, and Atherosclerosis in Low-Density Lipoprotein Receptor Knockout Mice

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Objective—The peroxisome proliferator–activated receptor (PPAR) δ regulates systemic lipid homeostasis and inflammation. However, the ability of PPARδ agonists to improve the pathology of pre-established lesions and whether PPARδ activation is atheroprotective in the setting of insulin resistance have not been reported. Here, we examine whether intervention with a selective PPARδ agonist corrects metabolic dysregulation and attenuates aortic inflammation and atherosclerosis.

Approach and Results—Low-density lipoprotein receptor knockout mice were fed a chow or a high-fat, high-cholesterol (HFHC) diet (42% fat, 0.2% cholesterol) for 4 weeks. For a further 8 weeks, the HFHC group was fed either HFHC or HFHC plus GW1516 (3 mg/kg per day). GW1516 significantly attenuated pre-established fasting hyperlipidemia, hyperglycemia, and hyperinsulinemia, as well as glucose and insulin intolerance. GW1516 intervention markedly reduced aortic sinus lesions and lesion macrophages, whereas smooth muscle α-actin was unchanged and collagen deposition enhanced. In aortae, GW1516 increased the expression of the PPARδ-specific gene Adip but not PPARα- or γ-specific genes. GW1516 intervention decreased the expression of aortic proinflammatory M1 cytokines, increased the expression of the anti-inflammatory M2 cytokine Arg1, and attenuated the iNos/Arg1 ratio. Enhanced mitogen-activated protein kinase signaling, known to induce inflammatory cytokine expression in vitro, was enhanced in aortae of HFHC-fed mice. Furthermore, the HFHC diet impaired aortic insulin signaling through Akt and forkhead box O1, which was associated with elevated endoplasmic reticulum stress markers CCAAT-enhancer-binding protein homologous protein and 78kDa glucose regulated protein. GW1516 intervention normalized mitogen-activated protein kinase activation, insulin signaling, and endoplasmic reticulum stress.

Conclusions—Intervention with a PPARδ agonist inhibits aortic inflammation and attenuates the progression of pre-established atherosclerosis. (Arterioscler Thromb Vasc Biol. 2014;34:52-60.)

Key Words: atherosclerosis ■ inflammation ■ insulin resistance ■ lipids

The principal cause of mortality in patients with type 2 diabetes mellitus is atherosclerosis, a chronic inflammatory disease that is the primary precursor underlying most cardiovascular events. Although the molecular and pathophysiological links between type 2 diabetes mellitus and atherosclerosis are not fully understood, a crucial factor is likely insulin resistance. This is, in part, because of the promotion of multiple independent risk factors associated with cardiovascular disease, including obesity, hypertension, and dyslipidemia. Dyslipidemia associated with insulin resistance is characterized by increased plasma triglyceride (TG)-rich very-low-density lipoprotein (VLDL) and cholesteryl ester–rich low-density lipoprotein (LDL), both of which can permeate a compromised endothelium and initiate atherogenesis. Therapeutic strategies to reduce plasma LDL have proven effective in reducing cardiovascular events. However, a significant unmet medical need persists, making VLDL-lowering strategies an attractive therapeutic target.

Subendothelial retention of atherogenic lipoproteins leads to a series of maladaptive immune responses culminating in the development of macrophage foam cells. Foam cells play a critical role in the progression of fatty streaks toward more advanced lesions. In particular, M1 macrophages secrete inflammatory effector cytokines such as interleukin (IL)-1β and tumor necrosis factor-α, driven predominantly by mitogen-activated protein kinase (MAPK) and nuclear factor

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(NF)-κB signaling. However, insulin signaling, namely the Akt/forkhead box O1 (FoxO1) pathway, may also contribute to arterial inflammation. In vitro, IIIβ is a FoxO1 target gene. Despite 1 study reporting the contrary, a growing body of evidence suggests that in vivo, arterial insulin resistance directly promotes atherosclerosis. Global deletion of Akt1 in apolipoprotein E–deficient mice accelerated coronary artery disease and aortic atherosclerosis, concomitant with significant aortic inflammation. Hematopoietic deletion of the insulin receptor in LDL receptor knockout (Ldlr−/−) mice amplified atherogenesis, an effect attributed to impaired macrophage Akt signaling. Furthermore, increased areas of apoptotic macrophages and necrotic core have been visualized in atherosclerotic lesions from patients with type 2 diabetes mellitus. Collectively, these studies support the concept that arterial insulin resistance promotes inflammation and atherogenesis.

Peroxisome proliferator–activated receptors (PPARs) are a class of ligand-dependant transcription factors involved in the regulation of metabolic and inflammatory signaling. Three isoforms exist (α, γ, δ), which exhibit overlapping but distinct patterns of tissue distribution and function. Although PPARδ has been considered the most enigmatic PPAR, this receptor has emerged as an important regulator of cellular lipid homeostasis and inflammatory responses. In cultured macrophages, PPARδ activation inhibits both macrophage lipid accumulation and pro-inflammatory cytokine expression in response to human VLDL. Furthermore, TG accumulation was decreased via angiopoietin-1/2 and activation of Akt/FoxO1 signaling. In vivo, Lee et al demonstrated that macrophage deletion of Pparδ in Ldlr−/− mice paradoxically decreased atherogenesis. This effect was attributed to the suppression of atherogenic inflammation by liberation of the inflammatory repressor protein B-cell lymphoma-6 because this protein is normally sequestered by the PPAR-retinoid X receptor corepressor complex. These studies highlight that deletion of Pparδ mimics the liganded state of the receptor, suggesting that ligand activation may be atheroprotective. However, studies examining the effects of synthetic PPARδ agonists using prevention protocols in mice have produced a spectrum of results.

In a second study, GW0742 reduced lesion development in female Ldlr−/− mice, although the doses used yielded serum drug levels 2-fold higher than the EC50 values for murine PPARα and PPARγ, raising the possibility that atheroprotection by GW0742 was not PPARδ specific. In Ldlr−/− mice fed a high-fat diet, low doses of GW0742 prevented the development of angiotensin II–accelerated atherosclerosis. The next-generation PPARδ agonist (GW1516) at PPARδ–specific doses prevented the development of atherosclerosis in apolipoprotein E–deficient mice fed a high-fat diet, concomitant with reduced aortic inflammatory cytokine expression. Although on balance these studies indicate that PPARδ–specific agonists prevent the development of atherosclerosis and arterial inflammation, it is unknown whether PPARδ agonists are atheroprotective in an intervention model with pre-established insulin resistance and atherosclerosis. Furthermore, the effect of PPARδ activation on lesion pathology, as well as aortic inflammatory signaling cascades, insulin resistance, and endoplasmic reticulum (ER) stress, has not been examined.

In the present study, we use C57BL/6 Ldlr−/− mice fed a high-fat, high-cholesterol (HFHC) diet, a model of diet-induced dyslipidemia and insulin resistance. After a diet induction phase, intervention with the addition of the PPARδ–specific agonist GW1516 to the HFHC diet resulted in reversal of metabolic dysregulation, including reduced plasma lipids, glucose, and insulin and improved glucose and insulin tolerance. Intervention with GW1516 inhibited aortic MAPK and NF-κB signaling, attenuated aortic inflammation, improved indices of aortic inflammation, reduced aortic ER stress, and collectively attenuated the progression of pre-established atherosclerosis.

Materials and Methods
Materials and Methods are available in the online-only Supplement.

Results
GW1516 Improves HFHC-Induced Metabolic Dysregulation in Ldlr−/− Mice
Male C57BL/6 Ldlr−/− mice were administered an HFHC diet for 4 weeks. The metabolic effects of intervention with the PPARδ agonist GW1516 were evaluated after an additional 8 weeks (Figure 1A). GW1516 at 3 mg/kg per day resulted in GW nonfasting serum concentrations at the end of the dark and light cycles of 604±72 and 369±26 nmol/L, respectively, for a mean concentration of 487±50 nmol/L. This plasma concentration is below the EC50 for murine PPARα (1 μmol/L) and well below the EC50 for murine PPARα (2.5 μmol/L). GW1516 intervention significantly attenuated HFHC-induced weight gain without affecting caloric intake (Figure 1A and IB in the online-only Data Supplement). GW1516 decreased fasting plasma cholesterol, TG, and nonesterified fatty acids compared with 4-week baseline levels, whereas dyslipidemia in mice remaining on the HFHC diet alone continued to progress (Figure 1B). Fast protein liquid chromatography analyses demonstrated that reduced plasma cholesterol in GW1516-treated mice was because of a significant reduction in VLDL cholesterol and a modest but not statistically significant reduction in LDL cholesterol (Figure 1C). GW1516 increased high-density lipoprotein cholesterol by 35% (Figure 1C). The GW1516-mediated
reduction in plasma TG was because of a substantial 63% reduction in VLDL-TG (Figure 1D). GW1516 intervention decreased epididymal fat by 11% compared with 4-week baseline and by 35% compared with mice remaining on the HFHC diet alone (Figure IC in the online-only Data Supplement), demonstrating attenuation of adipose tissue accumulation.

GW1516 prevented the increase in fasting blood glucose (from week 4 to week 12) and completely normalized impaired glucose tolerance induced by the HFHC diet at 12 weeks (Figure IIA and IIC in the online-only Data Supplement). GW1516 intervention reversed both fasting hyperinsulinemia and impaired insulin tolerance induced by the HFHC diet, demonstrating normalization of whole-body insulin sensitivity (Figure IIB and IID in the online-only Data Supplement).

This is consistent with the improved glucose tolerance in high-fat diet–fed C57BL/6J mice and improved insulin tolerance in db/db mice after a 2-week intervention with GW1516.\textsuperscript{21}

GW1516 Attenuates Aortic Sinus Atherosclerosis and Aortic Inflammation in HFHC-Fed Ldlr\textsuperscript{−/−} mice

Examination of aortic sinus atherosclerosis revealed that oil red-O–stained lesion area of HFHC-fed mice at 4 weeks progressed approximately 6-fold by week 12 (Figure 2A and 2C). In contrast, although lesion area continued to increase, the area was significantly attenuated in GW1516 intervention mice by ≈33% compared with animals remaining on HFHC alone (Figure 2C).

GW1516 intervention influenced lesion composition. As a percent of total area, lesions of HFHC-fed animals at either 4 or 12 weeks displayed accumulation of monocyte and macrophage antibody–2–positive macrophages, which was significantly attenuated by intervention with GW1516 (representative images are shown in Figure III in the online-only Data Supplement, and quantification is shown in Figure 2D). No appreciable smooth muscle (SM) α-actin or collagen deposition was observed in lesions of HFHC-fed mice at 4 weeks, and values were low in 12-week chow-fed mice (Figure 2E; Figure III in the online-only Data Supplement). However, SM α-actin occupied 40% of lesion area in HFHC-fed mice at 12 weeks, which was similar to that of GW1516 intervention mice. As assessed by trichrome staining, the HFHC diet at 12 weeks increased collagen deposition to 25% of lesion area, which was further increased (to 35%) in GW1516 intervention mice (data not shown), despite no further effect on percent lesion SM cell content (Figure 2E; Figure III in the online-only Data Supplement).

To further elucidate the effect of GW1516 intervention on lesion collagen, picrosirius red–stained mice sections were imaged with circular polarization microscopy to detect fibrillar collagen\textsuperscript{22} (Figure 2B). The collagen area fraction of lesions was 24% in HFHC-fed animals and 31% in lesions of mice subjected to GW1516 intervention (P<0.05; Figure 2F). This technique also revealed that a collagen-containing fibrinous cap existed in lesions in both circumstances (Figure 2B). To determine whether collagen fiber integrity in the cap was affected by GW1516 intervention, collagen birefringence was quantified (Figure 2B and 2G). This revealed a significant 36% increase in the retardation of polarized light by lesion cap collagen in mice subjected to GW1516 intervention (Figure 2G), denoting more densely packed and aligned collagen fibrils (Figure 2B, arrows). Lesion apoptosis, assessed by cleaved caspase-3 staining, was detected in 1.7% of cells within lesions from 12-week HFHC-fed mice (Figure 2H; Figure IV in the online-only Data Supplement). GW1516 intervention significantly reduced cleaved caspase-3 staining to 1.0% of cells (P<0.05), indicating a reduction in

**Figure 1.** GW1516 intervention improves diet-induced dyslipidemia. Low-density lipoprotein receptor knockout (Ldlr\textsuperscript{−/−}) mice were fed standard chow or a high-fat, high-cholesterol (HFHC) diet for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516); 3 mg/kg per day. A, Experimental timeline for all studies performed. B, Plasma cholesterol, triglyceride, and nonesterified fatty acid (NEFA) concentrations were measured at weeks 0, 4, and 12 (8–12/group). Arrows indicate the time of GW1516 intervention. C and D, Plasma was subjected to fast protein liquid chromatography (FPLC) analysis at week 12, and cholesterol and triglyceride were measured in the eluted fractions (n=3–5/group). Data are presented as mean±SEM. In B, different letters indicate significant differences (P<0.05). In C and D, *indicates significant difference vs HFHC (12 weeks; P<0.05). HDL indicates high-density lipoprotein; LDL, low-density lipoprotein; and VLDL, very-low-density lipoprotein.
GW1516 intervention attenuates high-fat, high-cholesterol (HFHC)-induced atherosclerosis. A, Representative photomicrographs of aortic sinus sections stained with oil red-O and counterstained with hematoxylin. Scale bar, 500 μm. B, Representative photomicrographs of aortic sinus sections stained with picrosirius red and imaged using circularly polarizing light and liquid crystal compensation. Scale bar, 250 μm. Arrows depict birefringent collagen fibers on the surface of atherosclerotic lesions, consistent with fibrous caps of varying organization. Color encoding of light retardation (nm) is depicted in the gradient map (blue: low; red: high). C, Quantification of neutral lipid area (oil red-O). D, MOMA-2 (macrophages) and E, smooth muscle (SM) α-actin (smooth muscle cells) expressed as lesion area (oil red-O) or % lesion area (MOMA-2, SM α-actin; n=6–9/group). Representative photomicrographs are available in Figure III in the online-only Data Supplement for MOMA-2 and SM α-actin. F and G, Quantification of collagen expressed as % lesion area (F) and mean collagen fibril light retardation (G), determined from picrosirius red–stained sections (B) and visualization using circularly polarized light (n=8–12/group). H, Quantification of lesion apoptosis determined by % caspase-3 act–positive cells relative to total cells in aortic sinus lesions (n=5–11/group). Representative photomicrographs are available in Figure IV in the online-only Data Supplement. Staining for collagen, SM α-actin, and caspase-3 act–positive cells was undetectable in lesions from HFHC 4-week sections. Values from individual mice are represented by symbols, and the mean is indicated by a single horizontal line. Different letters indicate significant differences (P<0.05).
NF-κB signaling in HFHC-fed mice at 12 weeks, as demonstrated by increased phosphorylated inhibitor of nuclear factor κ-B kinase and phosphorylated inhibitor of nuclear factor of κ light chain gene enhancer in B-cells, α (Figure 4B). In contrast, GW1516 intervention attenuates NF-κB activation (Figure 4B). This suggests that GW1516 diminishes aortic inflammation, in part, by attenuating diet-induced activation of inflammatory signaling cascades.

GW1516 Intervention Corrects Diet-Induced Aortic Insulin Signaling and ER Stress and Exerts PPARδ-Specific Vessel Wall Effects

Genetic manipulations resulting in impaired insulin signaling in hematopoietic cells exacerbate atherosclerosis, partly because of increased aortic inflammation and ER stress.4,9,10 We, therefore, hypothesized that impaired aortic insulin signaling contributed to the proinflammatory phenotype of aortae in HFHC-fed mice. Aortic phosphorylated Akt and phosphorylated FoxO1 in fasted and acutely refed mice were examined after the 8-week intervention phase. Compared with chow-fed mice, phosphorylated Akt and phosphorylated FoxO1 were higher in aortae from fasted HFHC-fed mice, but in contrast to chow-fed mice were not further increased by refeeding (Figure 5A). GW1516 intervention completely restored the fasting-to-feeding dynamic regulation of phosphorylated Akt and phosphorylated FoxO1 to patterns observed in chow-fed controls (Figure 5A). The Src homology 2 domain–containing tyrosine phosphatase-1 is primarily expressed...
by hematopoietic cells and is a known negative regulator of hepatic insulin signaling. Aortae from HFHC-fed mice at 12 weeks were significantly enriched for the Src homology 2 domain–containing tyrosine phosphatase-1 transcript (Ptpn6; 5-fold) and Src homology 2 domain–containing tyrosine phosphatase-1 protein (30-fold), both of which were strongly attenuated by GW1516 intervention (Figure 5B and 5C).

Concomitant with dysregulated aortic insulin signaling was a significant increase in ER stress markers GRP78 and CHOP in aortae of HFHC-fed mice at 12 weeks (Figure 5D). The CHOP target gene and negative regulator of insulin signaling, tribbles homolog 3 (Trib3),24 was also elevated (Figure 5E). GW1516 intervention restored GRP78, CHOP, and Trib3 to levels observed in chow-fed controls (Figure 5D and 5E).

To determine whether GW1516 exerted effects directly within the aorta, we examined the expression of known PPARδ target genes. Expression of adipose differentiation related protein (Adfp), angiopoietin-like 4 (Angptl4), and carnitine palmitoyltransferase (Cpt)-1α was significantly increased in aortae of GW1516-treated mice compared with HFHC-fed mice or chow-fed controls at 12 weeks (Figure 6A). Expression of target genes specific for PPARα (acyl-CoA oxidase [Acox]) and PPARγ (lipoprotein lipase [Lpl] and fatty acid binding protein 4 [Fabp4]) was unaffected by GW1516 intervention (Figure 6B). Similar results were observed in liver (data not shown). This indicates that GW1516 exerts a direct effect on the arterial wall, which likely contributes to attenuation of aortic inflammation, insulin resistance, and ER stress, as well as...
lesion progression. These data further indicate that with respect to PPARs, the aortic effects of GW1516 are PPARδ specific.

Discussion

The risk of atherosclerosis is elevated approximately 4-fold in adults with type 2 diabetes mellitus.1 Despite this, therapeutic strategies to alleviate atherosclerosis associated with insulin-resistant syndromes remain sparse. Here, we demonstrate in mice that intervention with a synthetic PPARδ agonist, in the context of diet-induced dyslipidemia and insulin resistance, attenuates the progression of early stage lesions to larger plaques. PPARδ activation was associated with beneficial changes in lesion composition, including fewer macrophages, increased expression of M2 markers, less lipid, increased collagen, decreased ER stress, and fewer apoptotic cells, characteristic of lesions with a more stable phenotype. Furthermore, we show that in HFHC-fed mice, the aberrant inflammatory response and impaired insulin signaling within the aorta are reversed by PPARδ activation.

Dyslipidemia associated with insulin resistance is characterized by elevated plasma VLDL and LDL, concomitant with reduced plasma high-density lipoprotein.3 Although statins effectively lower plasma LDL, they do not fully correct other features of atherosclerosis risk, namely elevated plasma VLDL, decreased high-density lipoprotein, insulin resistance, and body fat composition.25 The present study demonstrates that intervention by a PPARδ agonist corrects previously established metabolic disturbances. Although plasma LDL cholesterol was modestly reduced, PPARδ activation primarily targeted plasma VLDL and high-density lipoprotein. This is consistent with human studies demonstrating that PPARδ agonists correct mixed dyslipidemia in patients with metabolic syndrome.26,27 The current study contributes to the plausibility of PPARδ agonists as therapeutic agents for metabolic dysregulation associated with insulin resistance. Whether PPARδ agonists will have an effect in a setting where elevated LDL is a primary determinant of atherosclerosis remains to be determined.

We recently demonstrated in cultured macrophages that PPARδ activation attenuates VLDL-induced TG accumulation and proinflammatory cytokine expression.14 We extend these in vitro findings, demonstrating that GW1516 intervention prevents further increase in aortic TG, in concert with significant induction of the PPARδ target genes Angptl4 and Cpt1a. This suggests that GW1516 may stimulate aortic fatty acid β-oxidation and inhibit aortic lipoprotein lipase activity, thus contributing to reduced atherogenesis. We provide evidence that inflammatory cells within the aorta of HFHC-fed animals were polarized to the proinflammatory M1 phenotype.28,29 Furthermore, GW1516 intervention increased the anti-inflammatory M2 state,28,29 consistent with reports demonstrating that alternative M2 activation of adipose tissue macrophages and hepatic Kupffer cells is, in part, mediated by PPARδ.30,31 M2 macrophages are thought to contribute to tissue remodeling and repair29 and are increased in lesions undergoing regression.32 Although GW1516 did not induce size regression of early lesions, the M2 phenotype was associated with significant slowing of lesion progression. Longer-term studies are required to assess the effect of PPARδ agonists on more advanced stage lesions. Nevertheless, the present study demonstrates that PPARδ activation alleviates aortic lipid accumulation and inflammation, thus contributing to attenuated lesion development.

That GW1516 increased lesion collagen deposition without altering percent SM cell content is possibly as a result of PPARδ activation of the synthesis and deposition of extracellular matrix by lesion SM cells. This hypothesis is consistent with a report that PPARδ activation in cultured vascular SM cells inhibits IL-1β-induced matrix metalloproteinase-2 and matrix metalloproteinase-9 expression.33 Reduced lipid deposition in vascular SM cells restores their capacity to elaborate extracellular matrix.34,35 Thus, the ability of PPARδ agonists to improve the function of lipid-loaded vascular SM cells merits further attention.

The MAPK and NF-κB signaling pathways regulate inflammatory cytokine expression.36 In the aortae of HFHC-fed animals, both signaling cascades were significantly activated. GW1516 intervention substantially blunted these changes. In cultured cardiomyocytes, the PPARδ agonist GW0742 attenuated lipopolysaccharide-induced NF-κB activation through increased IκB expression, thereby inhibiting nuclear translocation of NF-κB.36 We did not observe appreciable changes in total aortic IκB protein. Thus, the mechanism by which PPARδ inhibits NF-κB activation in the context of aortic inflammation remains to be determined. With respect to MAPK activation, GW0742 inhibited angiotensin II–induced phosphorylation of extracellular signal–regulated kinase 1/2 and p38 in cultured mouse macrophages via upregulation of regulator of G-protein signaling (Rgs)4 and Rgs5.19 Consistent with this report, we observed a significant upregulation of both Rgs4 and Rgs5 in aortae of GW1516-treated mice compared with aortae of HFHC-fed mice (Figure V in the online-only Data Supplement). Collectively, our intervention studies suggest that PPARδ activation within the aorta dampens inflammatory signaling, leading to attenuation of inflammatory cytokine expression.

Impaired insulin signaling in the vasculature has emerged as a major contributor to lesion progression.4 In mice, macrophage deletion of the insulin receptor accelerated the development of advanced lesions,10 and loss of Akt1 led to severe atherosclerosis.5 Endothelial cell–specific deletion of 3 FoxO isoforms resulted in atheroprotection, attributed, in part, to an anti-inflammatory effect.37 Although these gene deletion models highlight the significance of vascular insulin signaling in atherogenesis, these studies do not identify whether insulin signaling becomes dysregulated during lesion development.38 Here, we demonstrate that mice with diet-induced atherosclerosis exhibit impaired aortic insulin signaling, as evidenced by loss of dynamic fasting-to-refeeding regulation of both Akt and FoxO1 phosphorylation, coupled to an induction of negative regulators of insulin signaling, Src homology 2 domain–containing tyrosine phosphatase-1123 and Trib3.24 This suggests that the loss of insulin regulation of both Akt and FoxO1 results in FoxO1 target genes such as Il6 being chronically transcribed rather than dynamically regulated. This mechanism may contribute to the accumulation of proinflammatory mediators within the vessel wall, inducing a state of chronic low-grade inflammation. Impaired aortic insulin
signaling was also correlated with elevations in ER stress markers GRP78 and CHOP. Furthermore, activation of aortic PPARδ restored dynamic insulin signaling and attenuated ER stress. It is important to note that the presence of arterial insulin resistance did not impair the ability of GW1516 to attenuate pre-established lesion progression. Thus, although difficult to quantify, it remains possible that improved insulin signaling within GW1516-treated aortae contributes to atheroprotection. In this study, a major factor in the attenuation of lesion development by intervention with GW1516 is reduction of plasma lipids, particularly VLDL/intermediate-density lipoprotein, thereby reducing the atherogenic stimulus. However, we demonstrate that in the aorta, GW1516 stimulates PPARδ-specific target genes, which are known to improve macrophage lipid homeostasis and attenuate the inflammatory response. Although these effects likely contribute to the observed reduction in atherosclerosis, further studies are required to elucidate the extent to which improved metabolic parameters versus direct vessel wall effects contribute to PPARδ-mediated atheroprotection. Nevertheless, the current study provides strong evidence that intervention to an HFHC diet with a PPARδ agonist suppresses and favorably modifies the HFHC diet-induced progression of early lesions. It will be important to determine whether intervention by PPARδ activation improves the pathology of more advanced lesions and whether extended treatment achieves regression. We conclude that PPARδ activation remains a viable therapeutic target for atherosclerosis prevention and treatment.

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Disclosures
None.

References
Vascular insulin resistance has been postulated to accelerate atherogenesis by increasing inflammation and endoplasmic reticulum stress. The peroxisome proliferator–activated receptor (PPAR) δ is a ligand-dependent transcription factor that regulates insulin sensitivity, lipid homeostasis, and inflammation. We demonstrate for the first time that intervention with a PPARδ agonist, in the context of diet-induced dyslipidemia and insulin resistance, attenuates the progression of early stage lesions to larger plaques. PPARδ activation was associated with beneficial changes in lesion composition, including fewer macrophages, increased M2 cytokine expression, less lipid, increased collagen, decreased endoplasmic reticulum stress, and fewer apoptotic cells, characteristic of more stable lesions. Furthermore, we show that in high-fat, high-cholesterol–fed mice, the inflammatory response and insulin signaling within the aorta are impaired, but these abnormalities are reversed by PPARδ activation. Collectively, these findings highlight a role for PPARδ agonists in the prevention and treatment of atherosclerosis, even in a setting of pre-established insulin resistance and atherosclerosis.

Significance
Peroxisome Proliferator–Activated Receptor δ Agonist GW1516 Attenuates Diet-Induced Aortic Inflammation, Insulin Resistance, and Atherosclerosis in Low-Density Lipoprotein Receptor Knockout Mice

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SUPPLEMENTAL FIGURES

PPARδ agonist GW1516 attenuates diet-induced aortic inflammation, insulin resistance and atherosclerosis in \( \text{Ldlr}^{-/-} \) mice

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Supplemental Figure I: GW1516-treatment attenuates rate of body weight gain and epididymal fat mass. Ldlr−/− mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). A, Body weight. Arrow indicates time of GW1516 intervention. B, Caloric intake expressed as kcal per gram body weight per day of study. C, Epididymal fat mass in grams. Data is presented as mean +/- SEM (n=8-12/group). Different letters indicate significant differences, (P<0.05).
Supplemental Figure II: GW1516 improves diet-induced dysregulation of metabolic indices.

Ldlr−/− mice were fed standard chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). A, Fasting blood glucose levels. B, Fasting plasma insulin concentrations. C, Intraperitoneal glucose tolerance test at 12 weeks. Inset graph, absolute area under the curve (AUC), (glucose mmol/Lx120min). D, Intraperitoneal insulin tolerance test at 12 weeks. Inset graph, absolute AUC (glucose mmol/Lx60min). Data is presented as mean +/- SEM (n=8-12/group). Different letters indicate significant differences, (P<0.05).
Supplemental Figure III: GW1516 improves atherosclerotic lesion morphology. *Ldlr<sup>−/−</sup>* mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). Mice were fasted for 4h prior to sacrifice. **A, and B**, Photomicrographs of serial sections of the aortic sinus stained with MOMA-2 (monocyte/macrophage antibody-2), antibody to smooth muscle (SM) α-actin, or trichrome. The bar indicates 500 μm (A) or 250 μm (B). Arrows indicate positive staining for macrophages (MOMA-2), smooth muscle cells (SM α-actin) or collagen (trichrome).
Supplemental Figure IV: GW1516 prevents lesional cell apoptosis

Ldlr−/− mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). Mice were fasted for 4h prior to sacrifice. Representative photomicrographs of aortic sinus sections stained with antibody to activated caspase-3 and counterstained with haematoxylin. M indicates media; L, lumen; Arrows, positive staining for activated caspase-3; Bar 125 μm.
Supplemental Figure V: GW1516 stimulates aortic expression of regulators of G-protein coupled receptor signaling *Rgs4* and *Rgs5*. *Ldlr*−/− mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). Mice were fasted for 4h prior to sacrifice. Analyses were performed on samples of full length aortae dissected free of fat and connective tissue. The indicated cytokines were measured by qRT-PCR. Target genes were normalized to *Gapdh*. Expression relative to chow depicted for each cytokine as mean +/- SEM (n=4-6/group). Different letters indicate significant differences, (P<0.05).
Materials and Methods

PPARδ agonist GW1516 attenuates diet-induced aortic inflammation, insulin resistance and atherosclerosis in Ldlr<sup>−/−</sup>mice

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Animals and Diets

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Plasma Lipid, Blood Glucose and Plasma Insulin Determinations

Plasma TG (Roche TG/GB, glycerol blanked, Roche Diagnostics, Laval, QC), total cholesterol (TC) and non-esterified fatty acids (NEFA) (Waco, VWR, Mississauga, ON) were measured using enzyme reagents as described previously.<sup>8</sup> Blood glucose was measured in whole blood using the Bayer Contour Blood Glucose Monitoring System (Bayer Healthcare, Etobicoke, ON).<sup>8</sup> Plasma insulin concentrations were determined by ELISA (Alpco Diagnostics, Salem, NH) on EDTA-plasma as per manufacturer’s instructions. Fast Performance Liquid Chromatography (FPLC) was performed on unfrozen EDTA-plasma using an AKTA purifier and Superose 6 column.<sup>8</sup>

Tissue Lipids

Lipids from full-length aortae dissected free of fat and connective tissue were extracted using the method of Folch et al., from samples stored at -80°C, as described previously.<sup>10</sup> 1α,2α(n)-[<sup>3</sup>H]Cholesteryl oleate and glycerol tril[<sup>14</sup>C]oleate (GE Healthcare, Piscataway, NJ) were added to assess recovery. Total triglyceride (TG) and total cholesterol (TC) were quantitated in samples solubilized in a 1% Triton X-100 solution in chloroform, dried under N<sub>2</sub>, and resolubilized in deionized water, prior to analysis by enzymatic, colorimetric assays using reagents obtained from Roche Diagnostics (TG) or Waco Chemicals (TC).<sup>10,11</sup>
Liquid Chromatography-Tandem Mass Spectrometry of GW501516

Serum samples (25 µL) were spiked with 5 µL internal standard (GW0742, Sigma-Aldrich, Oakville, ON, 0.25 ng/µL in methanol), vortexed and precipitated with 75 µL of acetonitrile. Standard curve samples were prepared using control mouse serum with GW501516 (0 – 500 ng/mL final concentration) and spiked with GW0742 (as above). Precipitated protein was pelleted by centrifugation at 14000g at 4°C for 10 min. Formic acid (90 µL of 0.1% formic acid in water) was added to post-precipitation supernatants (60 µL). Samples (25 µL) were injected into the liquid chromatograph (Agilent 1200, San Jose, CA) and analytes were separated using reverse phase chromatography on a Thermo Hypersil Gold C18 column (50 x 5 mm; 5 µm). Mobile phase was delivered at a flow rate of 0.5 mL/min with gradient flow with Mobile Phase A (0.1% formic acid in water) and Mobile Phase B (acetonitrile). For the first 60 sec, the mobile phase consisted of 30% Mobile Phase B then increased linearly to 95% Mobile Phase B over 4 min and held for 1 minute. Thereafter a linear decrease to 30% Mobile Phase B occurred over 1 min which was held for an additional 1 min. Retention times for GW501516 and GW0742 were both 5.0 and 5.2 min, respectively. Analytes were detected by tandem mass spectrometry on a Thermo TSQ Vantage triple quadrupole instrument equipped with heated electrospray ionization probe (HESI-II) set with probe voltage (4000 V), vaporizer temperature (350°C), ion transfer tube temperature (350°C), sheath gas (50 arbitrary units) and auxiliary gas (10 arbitrary units). The following multiple reaction monitoring transitions were used with detection in positive mode and collision gas (1.5 mTorr): GW501516 (454.0 → 257.1 m/z), GW0472 (472.0 → 275.1 m/z) with collision energies at 26 V and 28 V, respectively.

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Histological and morphometric analyses were performed at sacrifice, hearts were mounted in OCT and frozen. Frozen serial sections (70-100 per heart, 10µm) of the aortic sinus, initiating at the origin of the aortic valves, were prepared using a Leica CM 3050S cryostat. For quantitation of lesion area in the aortic sinus, oil red-O (Sigma Aldrich) stained sections were measured. Slides were stained with modified Verhoeff’s and Masson’s trichrome at the Robarts Research Institute Molecular Pathology Core facility. Immunohistochemistry staining for macrophages by MOMA-2 (Accurate #MCA519G, Westbury, NY), smooth muscle α-actin (monoclonal anti-α-smooth muscle actin, Clone 1AH, Sigma) and activated caspase-3 (anti-active caspase 3, ab13847, Abcam, Cedarlane, Burlington, ON) was performed as described. Slides were fixed in acetone and blocked in 2% bovine serum albumin (Sigma-Aldrich). After incubation with primary antibody, a goat biotinylated secondary antibody was used (Vector Laboratories, Burlington, ON). Slides were then incubated in peroxidase blocking reagent, followed by incubation with the ABC reagent (ABC Elite Standard Kit, Vector Laboratories). Slides were exposed to the DAB substrate (Peroxidase substrate kit, Vector Laboratories) followed by counterstain in hematoxylin (Sigma-Aldrich). Photomicrographs were obtained using an Olympus BX50 microscope and a QImaging Retiga EXi FAST camera. In the aortic sinus, lesion area of four serial sections (100µm apart) were quantified using Axiovision computer software. Morphometric analysis of MOMA-2, smooth
muscle α-actin and collagen (trichrome) in lesions from mice from each dietary group was also performed on serial sections. The relative area of the atherosclerotic plaque positive for MOMA-2, smooth muscle α-actin staining or collagen (trichrome) was determined as the area of positive staining divided by the area (from oil red-O-stained sections) of the respective plaque. Quantitation was determined using Axiosvision software. To determine the relative activation of caspase-3, the number of lesional cells per section immunostained for activated caspase-3 were counted and expressed as a percentage of the total lesional cells. To ensure that a standard region was measured in each mouse, lesion analysis began at the origin of the aortic valves.

**Collagen Birefringence Imaging**

Collagen fibrils were assessed using circular polarization microscopy as described previously.13-15 Frozen sections of aortic sinus were stained with picrosirius red (Polysciences, Warrington, PA) and visualized using an Olympus BX51 microscope (Olympus Canada, Inc., Richmond Hill, ON, Canada) equipped with circular polarizer/interference filters, a liquid crystal compensator, a charge-coupled device video camera, and Abrio image-processing software (Abrio LC-PolScope; Cambridge Research and Instrumentation, Inc., Hopkinton, MA). The acquired collagen retardation images are independent of illumination intensity. Collagen fibril content was measured with exported gray-scale images using ImageJ software and expressed as percent lesion area. Birefringence of collagen fibrils at the cap regions of lesions was assessed using Abrio software and expressed as mean retardation (in nanometers).

**Immunoblotting**

Total cell lysates were isolated from full-length aortae of mice as previously described.16,17 Proteins were separated by SDS-PAGE, transferred to polyvinylidene difluoride (PVDF) membranes and immunoblotted.17 Lysates were probed using antibodies against mouse phospho and total Akt, FoxO1, ERK1/2, p38, IKKα, and IκBα, as well as GRP78, CHOP, SHP-1 and β-actin (Cell Signaling, Danvers, MA). Protein levels were determined by densitometry as described.16,17

**Quantitative real-time PCR**

Total RNA was isolated from full-length aortae of mice using TRizol reagent (Life Technologies, Burlington, ON) as per manufacturer’s standard instructions. Total RNA (2µg) was reverse transcribed using the Applied Biosystems High Capacity cDNA reverse transcription kit (Life Technologies) according to the manufacturer’s protocol. Specific mRNA abundances (Ccl3, Il1b, Icam1, Tnf, Il6, Ccl2, Arg1, iNos, Ptnp6, Trib3, Adfp, Angptl4, Cpt1a, Acox, Lpl, Fabp4, Rgs4, Rgs5, and Gapdh) were measured via quantitative real-time PCR (qRT-PCR) of cDNA (10ng), in triplicate using an ABI Prism (7900HT) Sequence Detection System (Applied Biosystems) as previously described.8,18 Inventarioed TaqMan gene expression assays were obtained from Life Technologies. Abundance of target genes was normalized to Gapdh abundance.

**Statistical Analyses**

Data are expressed as means +/- SEM. One-way ANOVA followed by the Bonferroni test was used to determine significant differences between two groups. One-way ANOVA followed by pair-wise comparisons by the Tukey’s test was used to determine differences between three or more groups. For fasting/re-feeding experiments, two-way ANOVA followed by pair-wise comparisons by the Tukey’s test was used to determine differences and interactions between diet groups and fasted/re-fed states. Significance thresholds were P values less than 0.05 and indicated by different upper case or lower case letters as well as asterisks as indicated in the figure legends.
References


10. Burnett JR, Wilcox LJ, Telford DE, Kleinstiver SJ, Barrett PH, Newton RS, Huff MW. Inhibition of HMG-CoA reductase by atorvastatin decreases both VLDL and LDL


SUPPLEMENTAL FIGURES

PPARδ agonist GW1516 attenuates diet-induced aortic inflammation, insulin resistance and atherosclerosis in Ldlr/- mice

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Supplemental Figure I: GW1516-treatment attenuates rate of body weight gain and epididymal fat mass. *Ldlr*−/− mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). A, Body weight. Arrow indicates time of GW1516 intervention. B, Caloric intake expressed as kcal per gram body weight per day of study. C, Epididymal fat mass in grams. Data is presented as mean +/- SEM (n=8-12/group). Different letters indicate significant differences, (*P*<0.05).
Supplemental Figure II: GW1516 improves diet-induced dysregulation of metabolic indices. 

*Ldlr<sup>-/-</sup>* mice were fed standard chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). A, Fasting blood glucose levels. B, Fasting plasma insulin concentrations. C, Intraperitoneal glucose tolerance test at 12 weeks. Inset graph, absolute area under the curve (AUC), (glucose mmol/Lx120min). D, Intraperitoneal insulin tolerance test at 12 weeks. Inset graph, absolute AUC (glucose mmol/Lx60min). Data is presented as mean +/- SEM (n=8-12/group). Different letters indicate significant differences, (P<0.05).
Supplemental Figure III: GW1516 improves atherosclerotic lesion morphology. *Ldlr*<sup>−/−</sup> mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). Mice were fasted for 4h prior to sacrifice. **A** and **B**, Photomicrographs of serial sections of the aortic sinus stained with MOMA-2 (monocyte/macrophage antibody-2), antibody to smooth muscle (SM) α-actin, or trichrome. The bar indicates 500 µm (A) or 250 µm (B). Arrows indicate positive staining for macrophages (MOMA-2), smooth muscle cells (SM α-actin) or collagen (trichrome).
**Supplemental Figure IV:** GW1516 prevents lesional cell apoptosis

*Ldlr<sup>−/−</sup>* mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). Mice were fasted for 4h prior to sacrifice. Representative photomicrographs of aortic sinus sections stained with antibody to activated caspase-3 and counterstained with haematoxylin. M indicates media; L, lumen; Arrows, positive staining for activated caspase-3; Bar 125 μm.
Supplemental Figure V: GW1516 stimulates aortic expression of regulators of G-protein coupled receptor signaling Rgs4 and Rgs5. Ldlr⁻/⁻ mice were fed a standard lab chow, or a high-fat, high-cholesterol diet (HFHC) for 4 weeks. For a subsequent 8 weeks, chow-fed mice remained on chow; the HFHC-fed mice either remained on HFHC alone or HFHC supplemented with GW501516 (GW1516) (3mg/kg/day). Mice were fasted for 4h prior to sacrifice. Analyses were performed on samples of full length aortae dissected free of fat and connective tissue. The indicated cytokines were measured by qRT-PCR. Target genes were normalized to Gapdh. Expression relative to chow depicted for each cytokine as mean +/- SEM (n=4-6/group). Different letters indicate significant differences, (P<0.05).
Materials and Methods

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