Protein S Is a Cofactor for Platelet and Endothelial Tissue Factor Pathway Inhibitor-α but Not for Cell Surface–Associated Tissue Factor Pathway Inhibitor

Jeremy P. Wood, Paul E. R. Ellery, Susan A. Maroney, Alan E. Mast

Objective—Tissue factor pathway inhibitor (TFPI) is produced in 2 isoforms: TFPIα, a soluble protein in plasma, platelets, and endothelial cells, and TFPIβ, a glycosylphosphatidylinositol-anchored protein on endothelium. Protein S (PS) functions as a cofactor for TFPIα, enhancing the inhibition of factor Xa. However, PS does not alter the inhibition of prothrombinase by TFPIα, and PS interactions with TFPIβ are undescribed. Thus, the physiological role and scope of the PS–TFPI system remain unclear.

Approach and Results—Here, the cofactor activity of PS toward platelet and endothelial TFPIα and endothelial TFPIβ was quantified. PS enhanced the inhibition of factor Xa by TFPIα from platelets and endothelial cells and stabilized the TFPIα/factor Xa inhibitory complex, delaying thrombin generation by prothrombinase. By contrast, PS did not enhance the inhibitory activity of TFPIβ or a membrane-anchored form of TFPI containing the PS-binding third Kunitz domain (K1K2K3) although PS did function as a cofactor for K1K2K3 enzymatically released from the cell surface.

Conclusions—The PS–TFPI anticoagulant system is limited to plasma TFPIα and TFPIα released from platelets and endothelial cells. PS likely functions to localize solution-phase TFPIα to the cell surface, where factor Xa is bound. PS does not alter the activity of membrane-associated TFPI. Because activated platelets release TFPIα and PS, the PS–TFPI anticoagulant system may act physiologically to dampen thrombin generation at the platelet surface. (Arterioscler Thromb Vasc Biol. 2014;34:169-176.)

Key Words: blood coagulation • hemorrhage • thromboplastin • thrombosis

Protein S (PS)1 is a vitamin K–dependent anticoagulant protein that functions as a cofactor for activated protein C in the inactivation of factors Va (FVa) and VIIIa. Inherited deficiency of PS results in greatly increased risk for venous thrombosis.1 Aside from its role as a cofactor for activated protein C, PS also inhibits thrombin generation in an activated protein C–independent manner,2 an anticoagulant function attributed to either its enhancement of the inhibition of factor Xa (FXa) by tissue factor pathway inhibitor (TFPI)3,4 or direct inhibition of FXa and FVa, components of the thrombin-generating prothrombinase complex.5,6 The plasma concentration and anticoagulant activity of TFPI correlate with the plasma PS concentration, suggesting that these 2 proteins have a physiologically important interaction in vivo.7,8

TFPI is a reversible, slow, tight-binding inhibitor of FXa, which is produced in 2 isoforms: TFPIα and TFPIβ. TFPIα contains 3 Kunitz domains (K1, K2, and K3) and a highly basic C terminus,9 which interact for optimal inhibition of FXa. K1 is necessary for formation of the tight inhibitory complex; K2 directly binds the active site of FXa; and K3 and the C-terminal region enhance the initial binding.10–14 Circulating platelets store TFPIα and release it on activation.15,16 In addition, TFPIα circulates in plasma,17 but only 10% to 30% is fully active, full-length protein.18,19 Plasma TFPIα rapidly increases 2- to 4-fold after heparin infusion,19,20 suggesting that some TFPIα may be nonspecifically associated with glycosaminoglycans on the endothelium. However, studies of cultured endothelial cells suggest that the majority of endothelial TFPIα is stored within discrete intracellular granules and released after stimulation with heparin21,22 or thrombin.23 Surface-expressed TFPI on the endothelium seems to be primarily TFPIβ,24,25 in which the K3 domain and basic C terminus of TFPIα are replaced with a glycosylphosphatidylinositol (GPI)-anchor addition sequence, which localizes it to the cell surface.26–28

PS enhances the initial and the final (steady state) rates of inhibition of FXa by TFPIα.3,4,13 The interaction between PS and TFPIα is dependent on the presence of a membrane surface3,13 and K3,13,28 particularly residues Arg-19913 and Glu-226.28 The mechanism by which PS enhances the inhibitory activity of TFPI is not yet clear. The Gla domain of PS may localize TFPI to the membrane surface,29 thereby enhancing...
the interaction between TFPI and FXa, which also binds the membrane through a Gla domain. If so, one would predict that PS will serve as a cofactor only for soluble forms of TFPI. Alternatively, binding of PS may cause a conformational change in TFPI, which increases its affinity for FXA. If so, then PS may also serve as a cofactor for cell surface–expressed forms of TFPI, and membrane binding may promote the interaction between PS and TFPI.

It is currently unknown which pools of TFPI are involved in the PS–TFPI anticoagulant system. To date, all studies have used soluble TFPIα or plasma TFPI. However, the majority of intravascular TFPI is not circulating free in plasma and is instead either expressed on the surface of or released from the endothelium or released from platelets. We undertook studies to define the mechanism by which PS acts as a cofactor for TFPI and to define the physiological pools of TFPI involved in this anticoagulant system. The experimental results demonstrate that PS is a cofactor for TFPIα released from endothelial cells or platelets that acts by localizing TFPIα to a membrane surface. PS is not a cofactor for cell surface–associated forms of TFPI, regardless of the presence or absence of the K3 domain.

Materials and Methods

Materials and Methods are available in the online-only Supplement.

Results

PS Stabilizes the TFPIα–FXa Inhibitory Complex

PS enhances both the initial and final rates of FXa inhibition by TFPIα but not TFPI-160, in a dose-dependent manner, as measured using a chromogenic FXa substrate, confirming previously published results (Figure I in the online-only Data Supplement). Physiologically, FXa acts in prothrombinase (FXa/FVa/Ca2+/phospholipids) to generate thrombin. TFPIα is an effective inhibitor of free FXa, but a poor inhibitor of thrombin generation by prothrombinase assembled with thrombin-activated FVa.11 Ndonwi et al11 reported that TFPIα remains a poor prothrombinase inhibitor in the presence of PS. We extended this observation by quantifying the TFPIα-mediated inhibition of prothrombinase in the presence or absence of 80 nmol/L PS (Figure 1A). PS enhanced the inhibitory activity of TFPIα (50% inhibition observed at 6.5 versus 31.4 nmol/L in the absence of PS), but the concentration necessary for 50% inhibition was still higher than the circulating concentration of TFPIα (<1 nmol/L), suggesting that this is not a physiological inhibitory function. In the absence of TFPIα, PS had little effect on prothrombinase activity, with 10% inhibition observed in the presence of 500 nmol/L PS (Figure II in the online-only Data Supplement).

We have demonstrated recently that TFPIα is an efficient inhibitor of prothrombinase assembled with FXa-activated FVa.6 Because this inhibition is dependent on both K2 and the C terminus, regions flanking the PS-binding K3 domain, we hypothesized that PS may enhance this function of TFPIα. However, the inhibition of prothrombinase containing FXa-activated FVa by TFPIα in the presence of 80 nmol/L PS was
nearly identical to that reported in the absence of PS (IC₅₀=0.9 nmol/L for each; Figure 1B), indicating that PS has no effect on the kinetics of this reaction.

These data suggest that PS does not have a physiologically relevant impact on prothrombinase inhibition by TFPIα. However, the TFPIα–PS system delays thrombin generation in plasma-based assays, an effect which may be explained if PS stabilizes the inhibitory TFPIα–FXa complex that forms before prothrombinase assembly. To test this hypothesis, assays were performed in 2 ways: (1) FVa and TFPIα were preincubated for 30 minutes with prothrombin, cephalin, and a thrombin chromogenic substrate, with or without PS, and reactions initiated by addition of FXa; and (2) FXa and TFPIα were preincubated for 30 minutes with prothrombin, cephalin, and substrate, with or without PS, and reactions initiated by addition of FVa (Figure 1C). This assay was absolutely dependent on prothrombinase assembly, as no significant thrombin generation was observed in the absence of FVa (Figure 1C), and inhibition was dependent on TFPIα as PS had minimal effect on thrombin generation in this system in the absence of TFPIα (Figure III in the online-only Data Supplement). When initiating with FXa, the inclusion of PS resulted in a slight delay in thrombin generation. When initiating with FVa, thrombin generation was substantially delayed, reflecting the need for TFPIα to dissociate from FXa before prothrombin may be cleaved. Inclusion of PS in these reactions further delayed thrombin generation, suggesting that PS further stabilizes the TFPIα–FXa inhibitory complex. The delays in prothrombinase function observed in Figure 1C were quantified by measuring initial rates of thrombin generation (Figure 1D). TFPIα reduced the initial rate of thrombin generation >4-fold in reactions initiated with FVa compared with reactions initiated with FXa (1.22±0.12 and 0.28±0.06 nmol/L per second, respectively). TFPIα in the presence of PS prevented any detectable thrombin generation during the first 2 minutes in reactions initiated with FVa, and also produced an ≈2-fold decrease in the initial rate of thrombin generation in reactions initiated with FXa (0.67±0.06 versus 1.22±0.12 nmol/L per second). The difference in thrombin generation observed when FXa was preincubated with TFPIα alone or TFPIα with PS (Figure 1C) may be because of a difference in the amount of FXa inhibited during this incubation step, a difference in the amount of FXa released from inhibition during the assay, or both. To assess this, the initial rates of thrombin generation (Figure 1D) were used to estimate the percentage of uninhibited FXa at time t=0, and reactions were performed in the presence of that amount of FXa (0.023 nmol/L to mimic TFPIα and 0.00014 nmol/L to mimic TFPIα–PS; Figure 1E). Although thrombin generation was dose dependently related to FXa concentration, as anticipated, thrombin generation in the presence of low FXa concentrations was substantially slower than in the presence of preinhibited FXa (particularly when comparing 0.00014 nmol/L FXa to 0.1 nmol/L FXa inhibited with TFPIα and PS). Thus, FXa must be released from inhibition during the assay to explain the thrombin generation kinetics observed.

**PS Is a Cofactor for Platelet TFPIα**

PS has been described previously as a cofactor for plasma TFPIα. However, much of intravascular TFPIα is within platelets and endothelial cells. Therefore, we undertook studies to determine the PS cofactor activity toward these pools of TFPIα. Platelets release TFPIα and PS on activation. The role of platelet PS as a cofactor for platelet TFPIα was assessed using lysed human platelets. The platelet lysate, containing a final concentration of 0.01% TX-100, inhibited FXa activity (Figure 2A). FXa activity was restored partially by addition of a polyclonal antibody against PS and almost completely restored by addition of a monoclonal antibody against the K2 domain of TFPI, demonstrating that the inhibitory activity is mediated by TFPIα and enhanced by PS (Figure 2A; Figure IV in the online-only Data Supplement). In the absence of platelet lysate, PS (80 nmol/L), the antibodies, and the TX-100 detergent had no effect on FXa activity in the absence of platelet lysate. B. Reactions performed with increasing concentrations of PS. C. Platelets were activated with calcium ionophore, and FXa activity was restored partially by addition of a monoclonal antibody against the K2 domain or against PS (100 nmol/L each) or in the presence of exogenous PS (80 nmol/L). For all panels, the average progress curves from 3 platelet donors are shown.
PS Is Not a Cofactor for TFPIα

Secreted From Cultured Endothelial Cells

TFPIα is contained within cultured endothelial cells and released on treatment with heparin.20–22 The cofactor activity of PS toward endothelial TFPIα was examined by treating wild type and atherosclerosis-resistant EA.hy926 (AR-EA.hy926) cells with heparin, quantifying the amount of TFPIα released by ELISA (Table I in the only-only Data Supplement), and measuring FXa inhibition (Figure 3). The AR-EA.hy926 cells were studied because they do not express surface TFPI and, therefore, all of the heparin-releasable TFPIα is TFPIα secreted from intracellular stores.34 Heparin increased the TFPIα secreted into the supernatants of EA.hy926 and AR-EA.hy926 cells 3.2- and 5.9-fold, respectively (Table I in the online-only Data Supplement). PS enhanced the initial rate of inhibition by the supernatants of both heparin-treated EA.hy926 cell lines (Figure 3). In control experiments, heparin did not alter PS cofactor activity when added to supernatants of buffer-treated cells (data not shown).

PS Is Not a Cofactor for TFPIβ

TFPIβ is the primary form of TFPI on the endothelial surface,24–25 but PS cofactor activity toward TFPIβ has not been studied previously. Although this activity would circumvent the interaction of PS with K3, it was hypothesized that PS may form a membrane-associated complex with TFPIβ and enhance its inhibitory activity. This hypothesis was tested using Chinese hamster ovary (CHO) cells expressing TFPIβ, EA.hy926 cells, and human umbilical vein endothelial cells. PS did not enhance the initial or final rates of inhibition of FXα by TFPI expressed on any cell line analyzed (Figure 4A). PS also did not alter the initial rate of thrombin generation by prothrombinase containing FVa, in assays performed using any of the cell lines tested (Figure 4B). For each cell line, an inhibitory polyclonal antibody against TFPI reversed the cellular inhibition of FXα, demonstrating that TFPI was responsible for the inhibitory activity observed. Further, CHO cells not expressing TFPIβ and AR-EA.hy926 cells did not inhibit FXα (data not shown). In addition, cells not expressing TFPI did not inhibit FXα in the presence of PS, suggesting that the cellular membrane conditions are sufficient to prevent the PS multimerization-induced FXα inhibition described by Seré et al.35

Because PS did not enhance the inhibitory activity of cell surface–associated TFPI, additional experiments were performed to determine whether cell-associated TFPI can interact with PS after release from the cell surface. Phosphoinositol-specific phospholipase C (PIPLC) was used to release TFPI from the surface of CHO/TFPIβ, EA.hy926, and AR-EA.hy926 cells. TFPIβ, released from PIPLC-treated CHO/TFPIβ cells, was not enhanced by PS (Figure 4C). PIPLC treatment produced a 3.1-fold increase in the amount of total TFPI released from EA.hy926 cells, compared with buffer-only treatment (Table I in the online-only Data Supplement). In contrast, only a small amount of TFPIα was released (1.2-fold over mock treatment), suggesting that the PIPLC-releasable TFPI on the surface of these cells was TFPIα. PIPLC did not release TFPI from the AR-EA.hy926 cells, consistent with the lack of TFPI on the surface of these cells.34 In the presence of 80 nmol/L PS, the supernatants of PIPLC-treated EA.hy926 cells showed slight, statistically significant increases in both the initial and final (steady state) inhibition of FXα, compared with buffer treatment samples (P<0.001; Figure 4D). PS cofactor
activity was reversed by the addition of an antibody against the PS-binding K3 domain of TFPIα (Figure VI in the online Data Supplement), demonstrating that the observed cofactor activity is because of TFPIα secreted during the incubation period, and not PIPLC-released TFPIβ.

**PS Is Not a Cofactor for Cell Surface–Associated TFPI Containing the K3 Domain**

Because TFPIβ lacks the K3 domain, the absence of PS cofactor activity may be related to its inability to bind TFPIβ. PS cofactor activity toward cell surface–associated TFPI containing K3 was assessed using CHO cells expressing TFPIβ (Figure 5A) was assessed using CHO cells expressing TFPIβ, and CHO cells expressing TFPIβ, EA.hy926 cells, or AR-EA.hy926 cells, in the absence or presence of PS (80 nmol/L). The averages of ≥4 progress curves are shown. B. Discontinuous thrombin generation assays were performed in the presence of CHO cells not expressing TFPI (No TFPI), CHO cells expressing TFPIβ, EA.hy926 cells, or AR-EA.hy926 cells, in the absence or presence of PS (80 nmol/L). Shown are the ratios of the thrombin generation rates in the presence and absence of PS (means±SD; n=3). C and D. CHO/TFPIβ (C) or EA.hy926 (D) cells were treated with (gray) or without (black) phosphoinositol-specific phospholipase C (PIPLC; 1 U/mL) for 1 hour, and FXa amidolytic assays were performed in the presence of the supernatant and in the absence (lines) or presence (dashes) of PS (80 nmol/L). The averages of 6 progress curves are shown.

**Discussion**

The ability of PS to enhance the inhibition of FXa by plasma TFPI and by recombinant TFPIα produced in *Escherichia coli* was originally demonstrated by Hackeng et al. Subsequent studies have investigated the interaction of PS and TFPIα in plasma, finding that patients with PS deficiency have decreased plasma TFPIα, that immunodepletion of PS depletes plasma TFPIα as well, and that plasma TFPIα correlates with free PS, rather than with C4bp-bound PS. Thus, it seems that there is a physiologically relevant PS–TFPI anticoagulant system functioning in vivo, yet much remains unclear. The majority of intravascular TFPI is TFPIβ on the endothelium, which does not have the PS-binding K3 domain. In addition, the repercussions of FXa inhibition by PS–TFPIα are not apparent, as TFPIα is a poor inhibitor of thrombin production by the prothrombinase complex assembled with FXa and thrombin-activated FVa, and PS does not enhance this inhibitory function to physiologically relevant rates (Figure 1D).

The current study was designed to further characterize the human PS–TFPI anticoagulant system by quantifying PS cofactor activity directed toward physiological pools of TFPI that have not been examined previously. Our results demonstrate that PS enhances the inhibition of FXa by solution-phase TFPIα, including platelet TFPIα and TFPIα released from cultured endothelial cells. In contrast, PS has no effect on the inhibition of FXa by surface-associated TFPIβ on transfected CHO cells or on cultured endothelial cells; or by solution-phase TFPIα released from the cell surface by PIPLC. These findings suggest that PS exerts its cofactor
activity by localizing TFPIα to the cell surface, where it can readily interact with membrane-associated FXa, and that cell surface TFPI is not affected by PS. This notion is supported by experiments using an altered form of TFPI expressed in CHO cells that contains the 3 Kunitz domains attached to the cell surface via a GPI-anchor. Inhibition of FXa by this form of TFPI was unaffected by PS when localized to the cell surface but was enhanced by PS after removal from the cell surface with PIPLC. However, these experimental results do not absolutely rule out a PS-induced conformational change in TFPIα that may contribute to the enhanced inhibitory activity because PS may not have been able to bind K3 when the GPI-anchored protein was bound to the cell surface. Regardless, the data presented in Figure 4 demonstrate that PS has no significant cofactor activity toward forms of TFPI endogenously expressed on the surface of endothelial cells.

Recent data from our laboratory have demonstrated that TFPIα is a potent inhibitor of prothrombinase assembled with FXa-activated FVa.32 This inhibition requires (1) an interaction between the K2 domain and the FXa active site and (2) an interaction between the basic TFPIα C terminus and an acidic region within the FV B-domain, which is retained after activation by FXa but removed after activation by thrombin. As such, this TFPIα inhibitory activity is relevant only during the initiation phase of thrombin generation. We hypothesized that PS would enhance this inhibitory activity. However, PS had no effect (Figure 1B) on the ability of TFPIα to inhibit prothrombinase assembled with FXa-activated FVa.

In contrast to the results with purified prothrombinase, the TFPIα–PS anticoagulant system inhibits thrombin generation in plasma-based assays.3 We assessed the impact of PS on the preformed TFPIα–FXa inhibitory complex in an effort to explain this observation, finding that PS stabilizes the inhibitory complex. This PS activity was observed in thrombin generation experiments in which the TFPIα–FXa or PS–TFPIα–FXa complexes were allowed to form before the addition of thrombin-activated FVa. A significant delay in thrombin generation occurred in reactions containing the PS–TFPIα–FXa complex compared with those lacking PS. If the inhibitory complex was not formed before initiation, then TFPIα was a poor inhibitor of prothrombinase, even in the presence of PS as previously described.13,31 These results suggest that the PS–TFPIα anticoagulant system may regulate prothrombinase function, inhibiting FXa and sequestering it from FVa or prothrombin. As TFPIα effectively inhibits cleavage of chromogenic substrates by prothrombinase,37 the more likely explanation is that dissociation of TFPIα from FXa allows prothrombinase to bind prothrombin.

Murine plasma TFPI contains only K1 and K2,25,38 and, consequently, PS does not serve as its cofactor.39 Because mice do not experience a hypercoagulable state, the key components of the PS–TFPIα anticoagulant system may be mediated through interactions of PS with TFPIα released from activated platelets, which secrete TFPIα15,16 and PS.33,40 In the studies presented here, using human platelets, platelet PS was a cofactor for platelet TFPIα inhibitory activity present either in platelet lysates or ionophore-activated platelets. Platelet PS was not saturating in the platelet lysates but was near saturating at the surface of activated platelets, suggesting that it may be released at sufficient localized concentration to saturate TFPIα within a platelet plug in vivo. The in vivo significance of the interaction among platelet PS, platelet TFPIα, and FXa will depend on the conditions required for PS and TFPIα release. If they are both released early in the hemostatic response, then they may be regulating initial prothrombinase. If they are only released after stimulation with thrombin, then they likely do not regulate initial prothrombinase but may serve a role in either spatially containing or downregulating the coagulant response after formation of the fibrin clot. In contrast to the work of Stavenuiter et al,41 we did not observe direct inhibition of FXα by platelet PS in the absence of TFPIα in our assay systems. The reason for these contrasting data is not apparent but may result from the different antibody reagents used in the 2 studies.

Our laboratory has demonstrated that hematopoietic cell TFPI, most likely from platelets, dampens thrombus growth in a murine model of large vessel vascular injury and directly modulates bleeding in a murine hemophilia model.42,43 In addition, it has been demonstrated that degradation of platelet TFPIα by neutrophil proteases increases platelet procoagulant activity both in vitro and in vivo.44,45 Although these actions of platelet TFPIα may result from inhibition of circulating forms of TF, platelet PS and platelet TFPIα may also assemble as a physiologically relevant anticoagulant complex on the surface of activated platelets where they act to prevent development of occlusive intravascular thrombi.
by slowing thrombin generation through the mechanism described above. It is important to note that the TFPIα-PS system is most effective at inhibiting prothrombinase when the inhibitory complex is allowed to form before introduction of FVα and prothrombin. If all components are present, then the IC₅₀ for inhibition (6.5 nmol/L) suggests an unfavorable interaction. However, many factors may alter these kinetics, including (1) the potential for differential release of TFPIα, PS, and FVα from platelet storage granules, (2) the high local concentrations of these proteins within a platelet plug, and (3) the presence of Zn²⁺ bound to plasma PS, which is necessary for its direct inhibition of FXα.⁶

Platelets are thought to be the primary site of prothrombinase assembly and thrombin generation; however, other cell types may also support this function, including monocytes, lymphocytes, neutrophils, erythrocytes, and endothelial cells.⁴⁻⁶ Endothelial cells also release TFPIα from intracellular stores in response to agonists such as heparin and thrombin.²⁰⁻²³ Data presented here support this, demonstrating the release of TFPIα into the supernatants of cultured AR-EA.hy926 cells, which do not express surface TFPI. Thus, it is tempting to speculate that plasma PS may interact with plasma TFPIα or TFPIα released from activated endothelium to regulate thrombin production on vascular cells other than platelets. Collectively, these data demonstrate that PS exerts anticoagulant cofactor activity with TFPIα from any physiological pool, likely by localizing TFPIα to membrane surfaces, stabilizing its interaction with membrane-bound FXα, and slowing thrombin generation. However, PS has no effect on TFPIβ or other forms of membrane-associated TFPI. We hypothesize that a physiological function of the PS–TFPIα anticoagulant system is to slow the generation of thrombin, on platelets and other vascular cells, by inhibiting FXα before FVα and prothrombin become available.

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Disclosures
None.

References
38. First time demonstrate that this system can slow prothrombinase assembly.


**Significance**

Protein S (PS) is a cofactor for factor Xa inhibition by tissue factor pathway inhibitor (TFPI). Previous studies with PS have assessed its effect as a cofactor for TFPI, using plasma or purified recombinant protein although the majority of vascular TFPI is TFPIβ. Here, we demonstrate that PS is a cofactor for TFPIβ released from platelets or endothelial cells, and likely acts by localizing soluble TFPIβ to a membrane surface, thereby stabilizing its interaction with membrane-bound factor Xa, sequestering it from prothrombin, and slowing thrombin generation. By contrast, PS has no cofactor activity toward TFPIβ. These data define the physiological scope of the TFPI/PS anticoagulant system and for the first time demonstrate that this system can slow prothrombinase assembly.
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Supplemental Figure I. Protein S is a cofactor for the inhibition of FXa amidolytic activity by TFPIα. TFPIα (A) or TFPI-160 (B) (1 nM) was incubated with cephalin (1/500), FXa (0.1 nM), PS (indicated concentrations), and FXa chromogenic substrate (0.5 mM). The average of 6 progress curves is shown.
Supplemental Figure II. Protein S does not inhibit prothrombinase in the absence of TFPIα. Protein S (1-500 nM) was incubated with phospholipid vesicles (20 μM), FVa (0.5 nM), and the thrombin inhibitor DAPA (3 μM) for 30 min. Reactions were initiated by the addition of prothrombin (1.4 μM) and FXa (0.1 nM), aliquots quenched by addition of EDTA (33 mM), and thrombin generation measured by addition of a thrombin chromogenic substrate (0.32 mM). Shown are the thrombin generation rates (% of control) (mean±SEM; n=3).
Supplemental Figure III. Protein S does not inhibit prothrombinase assembly or function in the absence of TFPIα. Reactions containing cephalin (1/500), prothrombin (1.4 µM), thrombin chromogenic substrate (0.5 mM) and either FXa (0.1 nM) (solid lines) or FVa (5 nM) (dashed/dotted lines) were performed either in the presence (grey) or absence (black) of PS (80 nM). Reactions incubated with FVa were initiated by the addition of FXa and vice versa. The average of ≥3 progress curves is shown.
Supplemental Figure IV. Protein S cofactor activity may be blocked with a polyclonal anti-PS antibody. Factor Xa (0.1 nM) was incubated with cephalin (1/500) and a chromogenic substrate (0.5 mM), and the absorbance at 405 nm was monitored. In some experiments, the FXa and cephalin were pre-incubated with TFPIα (0.5 nM); TFPIα and PS (2.5 nM); or TFPIα, PS, and rabbit anti-PS antibody (100 nM). Shown are the average progress curves from 3 experiments.
Supplemental Figure V. Protein S enhances the initial rate of inhibition of FXa by platelet TFPIα. FXa inhibition reactions were performed with increasing concentrations of PS as described in Figure 2B, and the initial (○) and final (■) free FXa concentrations were determined from the progress curves (mean ± SEM; n≥4).
Supplemental Figure VI. Protein S is a cofactor for TFPIα in the supernatants of PIPLC-treated EA.hy926 cells. EA.hy926 cells were treated either with PIPLC (blue) or buffer (red), as described in Figure 4, and the supernatants were tested for FXa inhibitory activity in the absence (solid lines) or presence of PS (10 nM) (dashed lines) or PS and an antibody against the K3 domain of TFPIα (100 nM) (dashed/dotted lines). Also shown is a FXa control (black). All data are presented as the average progress curves from 3 experiments.
Table I. Release of TFPI from EA.hy926 and AR-EA.hy926 cells by treatment with heparin.

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<tr>
<th></th>
<th>Condition</th>
<th>Total TFPI (pM)</th>
<th>TFPIα (pM)</th>
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<tbody>
<tr>
<td>EA.hy926</td>
<td>mock</td>
<td>235 ± 13</td>
<td>224 ± 36</td>
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<td></td>
<td>Heparin</td>
<td>1307 ± 137</td>
<td>713 ± 80</td>
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<td></td>
<td>PIPLC</td>
<td>718 ± 81</td>
<td>260 ± 22</td>
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<td>AR-EA.hy926</td>
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<td>Heparin</td>
<td>655 ± 61</td>
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<tr>
<td></td>
<td>PIPLC</td>
<td>137 ± 11</td>
<td>109 ± 16</td>
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*Data are shown as mean ± SEM (n=3).
Materials and Methods

Materials. Dulbecco’s Modified Eagle’s Medium (DMEM), F-12K medium, G418, and penicillin/streptomycin were from Mediatech (Manassas, VA). Heparin, phenylmethylsulfonyl fluoride (PMSF), polymyxin B, the cysteine protease inhibitor \(\text{trans-epoxysuccinyl-L-leucylamido(4-guanidino)butane} \) (E-64), and the calcium ionophore A23187 were from Sigma-Aldrich (St. Louis, MO). Hygromycin B was from Invitrogen (Carlsbad, CA). Prostaglandin E1 (PGE1) was from MP Biomedicals (Solon, OH). The thrombin inhibitor dansylarginine-N-(3-ethyl-1,5-pentanediyl)amine (DAPA) was from Haematologic Technologies (Essex Junction, VT). The FXa chromogenic substrate (Spectrozyme Xa) and the thrombin chromogenic substrate (Spectrozyme TH) were from American Diagnostica (Stamford, CT). Arg-Gly-Asp-Ser was from Genscript (Piscataway, NJ).

TFPI\(\alpha\) and TFPI-160 (containing only K1 and K2) were obtained as described previously\(^1\). Bovine FXa was from Enzyme Research Laboratories (South Bend, IN). Human PS, FV, and prothrombin were from Haematologic Technologies. FV was activated by incubation with thrombin (2 nM) or FXa (1 nM) for 10 min at 37°C. When activating with FXa, phospholipid was included in the activation mixture. Mouse anti-TFPI\(_{K2}\) and anti-TFPI\(_{K3}\) monoclonal antibodies were from Novo Nordisk ( Bagsvaerd, Denmark) and rabbit brain cephalin was from Marc Goldford (Sigma-Aldrich). Rabbit polyclonal antibodies against full-length TFPI\(\alpha\) and the TFPI\(\alpha\) C-terminus were as described previously\(^2\). Goat anti-protein S was from Cedarlane Labs (Burlington, NC). Peroxidase-conjugated donkey anti-rabbit IgG was from Jackson ImmunoResearch Laboratories (West Grove, PA). Bovine serum albumin (BSA) was from Sigma-Aldrich, and casein was from G Biosciences (Maryland Heights, MO). Fetal bovine
serum (FBS) was from Biowest USA (Miami, FL). Trypsin-ethylenediaminetetraacetic acid (EDTA) solution and phosphoinositol-specific phospholipase C (PIPLC) were from Invitrogen.

**Cell Culture.** Chinese hamster ovary (CHO) cells, transfected with tissue factor (TF) (CHO-TF) and then co-transfected with TFPIβ or with an altered form of TFPI that contains K1, K2, and K3 with a GPI anchor attachment sequence (K1K2K3)\(^3,4\), were cultured in F-12K media supplemented with FBS (10%), G418 (0.8 mg/mL), hygromycin (0.4 mg/mL), and polymyxin B (50 µg/mL). The EA.hy926 (ATCC, Manassas, VA) and aerolysin-resistant EA.hy926 (AR-EA.hy926)\(^2\) endothelial cell lines were cultured in DMEM media supplemented with FBS (10%) and penicillin/streptomycin (1%). HUVECs were obtained from the Hybridoma Core Facility (Blood Center of Wisconsin, Milwaukee, WI) and used between passage 3 and 6.

**Cell harvesting and standardization.** Cells, seeded onto 100 mm plates, were grown to 90% confluence, washed twice with phosphate-buffered saline (137 mM NaCl, 2.68 mM KCl, 4.76 mM Na\(_2\)HPO\(_4\), 0.73 mM KH\(_2\)PO\(_4\), pH 7.4; PBS), and harvested with PBS containing 5 mM EDTA. Cells were washed with PBS and resuspended in 20 mM HEPES, 150 mM NaCl, pH 7.4 (HBS), containing 5 mM CaCl\(_2\) and 0.1% BSA (HBS/Ca\(^{2+}\)/BSA). An aliquot of the cellular preparation was solubilized (in PBS containing 30 mM CHAPS and 10 mM EDTA) and the total protein concentration measured using a bicinchoninic acid assay (ThermoScientific, Rockford, IL). Total cellular protein concentrations were used to standardize all of the experiments assessing cell surface-associated TFPI, as these were found to be more reproducible than cell counting. The following cellular concentrations were found to express comparable FXa inhibitory activity and were thus used for all assays: CHO/K1K2K3 – 200 µg/mL; CHO/TFPIβ – 50 µg/mL; and EA.hy926 – 50 µg/mL. CHO-TF cells and AR-EA.hy926, which do not express detectable FXa inhibitory activity, were used at 200 µg/mL and 50 µg/mL, respectively.
Treatment of cells with PIPLC and heparin. EA.hy926 or AR-EA.hy926 cells, used at 90% confluence, were washed twice with PBS, then incubated (1 hr, 37°C) with HBS/Ca^{2+}/BSA containing 0.2 mM E-64 and 1 U/mL PIPLC or 10 U/mL heparin. Control cells were incubated in buffer alone. The cell supernatants were saved for analysis.

Preparation of platelet lysates. Studies using human blood were approved by the Blood Center of Wisconsin Institutional Review Board. Informed consent was obtained from all donors. Platelets were isolated from citrated blood as described^5. After the final wash, the platelets were resuspended in HEPES-Tyrode’s buffer (5 mM HEPES, 137 mM NaCl, 2.68 mM KCl, 11.9 mM NaHCO_3, 0.42 mM NaH_2PO_4, 1 mM MgCl_2, 2 mM CaCl_2, 0.2% dextrose, 0.35% BSA, pH 7.4; HTA), and the concentration determined using an ABC Hematology Analyzer (scil Animal Care, Gurnee, IL). Platelets were diluted to 1x10^{10}/mL in HTA containing PMSF (1 mM) and E-64 (0.2 mM), and solubilized in 1% Triton X-100. Insoluble material was removed by centrifugation (20,000xg, 10 min). The lysate was incubated on ice for two hours to allow for PMSF hydrolysis.

Preparation of activated platelets. Washed platelets (3x10^8 platelets/mL in HTA containing 1 mM Arg-Gly-Asp-Ser) were incubated with calcium ionophore (2 µM) for 30 min at 37°C.

FXa amidolytic activity assay. Reactions were prepared using activated platelets (1x10^8 platelets/mL), platelet lysate (1x10^8 platelets/mL), cell supernatant following PIPLC or heparin treatment (1/3 dilution), varying concentrations of TFPIα or TFPI-160, or intact cells (at the concentrations indicated in “Cell harvesting and standardization”). All reactions were prepared in HBS/Ca^{++}/BSA containing 0.5 mM FXa chromogenic substrate. Reactions performed in the absence of intact cells were supplemented with cephalin (1/500 dilution) to provide a membrane surface. This dilution of cephalin was found to be optimal and to provide sufficient phospholipid to prevent direct FXa inhibition by PS multimers, as was described by Seré and colleagues^6.
Reactions were incubated at 23°C for 30 min, initiated by addition of FXa (0.1 nM), and substrate cleavage monitored at 405 nm on a SpectraMax Plus 384 plate reader (Molecular Devices, Sunnyvale, CA) running SoftMax Pro v4.8 analysis software. For some experiments, PS (varying concentration), mouse anti-TFPI\textsubscript{K2} (100 nM), mouse anti-TFPI\textsubscript{K3} (100 nM), rabbit anti-TFPI (1/400), or goat anti-PS (100 nM) antibody was included in the incubation mixture.

Initial and steady state velocities of substrate cleavage were determined as described\textsuperscript{7}. Briefly, the data were fitted to the equation:

$$A_t = v_s t + \frac{(v_i - v_s)}{k_{obs}}[1 - \exp(-k_{obs}t)] + A_o$$

where $A_t$ and $A_o$ are absorbances at time $t$ and time 0, respectively, $v_s$ and $v_i$ are the steady state and initial velocities, and $k_{obs}$ is the rate of change between $v_i$ and $v_s$. Curve fitting was performed using GraphPad Prism v. 5 (GraphPad Software, La Jolla, CA). The initial and steady state (final) velocities were converted to free (uninhibited) FXa concentrations by comparison to a standard curve.

**Continuous thrombin generation assay.** Reactions containing prothrombin (1.4 µM), cephalin (1/500 dilution), and thrombin chromogenic substrate (0.5 mM) were prepared in HBS/Ca\textsuperscript{2+}/BSA. Either FVa (5 nM) or FXa (0.1 nM) was added, and the reactions were incubated for 30 min at 23°C. Reactions containing FVa were then initiated by the addition of FXa, and vice versa. Thrombin generation was monitored by the change in absorbance at 405 nm, as described for the FXa amidolytic activity assay. For some reactions, TFPI\textsubscript{a} (0.5 nM) and/or PS (80 nM) was included in the incubation mixture.

**Discontinuous thrombin generation assay.** Cells (diluted in HBS/Ca\textsuperscript{2+}/BSA as described for the FXa amidolytic activity assay) were incubated for 30 min at 23°C with prothrombin (1.4 µM), DAPA (3 µM), and either FVa (5 nM) or FXa (0.1 nM). For some experiments, cells were
omitted and TFPIα (0.5 nM) and cephalin (1/500 dilution) were included in the incubation mixture. Protein S (80 nM) was added in some reactions. Reactions pre-incubated with FVa were initiated by addition of FXa and vice versa. Aliquots were removed every 20 s for 2 min and quenched in HBS containing 33.3 mM EDTA and 0.1% PEG-8000. Thrombin generation was determined by measuring cleavage of a thrombin chromogenic substrate (0.4 mM and comparison to a standard curve.

**Determination of IC**$_{50}$ **values.** Reactions were prepared containing thrombin- or FXa-activated FVa (5 nM), DAPA (3 µM), and phospholipid vesicles (20 µM) in HBS containing 5 mM CaCl$_2$ and 0.1% PEG-8000. (Phospholipid vesicles contained 40% phosphatidylcholine, 40% phosphatidylethanolamine, and 20% phosphatidylserine and were prepared as described$^8$.) Either TFPIα or PS (varying concentrations) was added, and the reactions were incubated at room temperature for 20 min. Reactions using TFPIα were incubated in the presence or absence of PS (80 nM). Following incubation, reactions were initiated by the addition of prothrombin (1.4 µM) and FXa (0.5 nM), and aliquots were removed, quenched, and analyzed as described above. The initial rate of thrombin generation at each TFPIα or PS concentration was fit to a four parameter variable-slope curve to determine the IC$_{50}$ value, using GraphPad Prism v. 5.

**Measurement of TFPI antigen.** Concentrations of total TFPI and TFPIα in the supernatants of PIPLC- and heparin-treated cells were measured by enzyme-linked immunosorbertant assay (ELISA) as described$^4$. Briefly, microtiter plate wells were coated with mouse anti-TFPI$_{K2}$ (2 µg/mL) and blocked with PBS containing casein (0.5%) and BSA (0.1%). Total TFPI and TFPIα were detected using rabbit polyclonal antibodies (1/250 dilution each) against the entire TFPIα molecule and the TFPIα C-terminus, respectively. The primary antibodies were detected using peroxidase-conjugated donkey anti-rabbit IgG (0.1 µg/mL), followed by QuantaRed Enhanced
Chemifluorescent HRP Substrate (ThermoScientific), and TFPI concentrations were calculated by comparison to a TFPIα standard curve.

References

1. Lockett JM, Mast AE. Contribution of regions distal to glycine-160 to the anticoagulant activity of tissue factor pathway inhibitor. *Biochemistry*. 2002;41:4989-4997