Bone Morphogenetic Proteins Protect Pulmonary Microvascular Endothelial Cells From Apoptosis by Upregulating α-B-Crystallin

Mariana Ciumas,* Mélanie Eyries,* Odette Poirier,* Svetlana Maugenre, France Dierick, Natalia Gambaryan, Kevin Montagne, Sophie Nadaud, Florent Soubrier

Objective—To investigate the role of bone morphogenetic proteins (BMPs) on α-B-crystallin (CRYAB) expression and its physiological consequences on endothelial cells (ECs).

Approach and Results—We report that the gene encoding for the small heat shock protein, CRYAB, is a transcriptional target of the BMP signaling pathway. We demonstrate that CRYAB expression is upregulated strongly by BMPs in an EC line and in human lung microvascular ECs and human umbilical vein ECs. We show that BMP signals through the BMPR2-ALK1 pathway to upregulate CRYAB expression through a transcriptional indirect mechanism involving Id1. We observed that the known antiapoptotic effect of the BMPs is, in part, because of the upregulation of CRYAB expression in EC. We also show that cryab is downregulated in vivo, in a mouse model of pulmonary arterial hypertension induced by chronic hypoxia where the BMP pathway is downregulated.

Conclusions—We demonstrate a cross-talk between BMPs and CRYAB and a major effect of this regulatory interaction on resistance to apoptosis. (Arterioscler Thromb Vasc Biol. 2013;33:2577-2584.)

Key Words: ACVRL1 protein, human ■ anoxia ■ BMP2 protein, human ■ Id1 protein, human ■ RNA, small interfering ■ Smad proteins

Bone morphogenetic proteins (BMPs) belong to the transforming growth-factor-β superfamily of extracellular signaling proteins and play crucial roles in embryonic development, adult tissue homeostasis, and in the pathogenesis of a variety of diseases.1 The highly conserved BMP signaling pathway comprises the BMP ligands, 2 types of receptors (type I: ALK1, 2, 3, and 6 and type II: BMPR2, ACTR2A, and ACTR2B), and signal transducers, the Smad proteins. Once activated, the receptor complex phosphorylates the carboxy terminus of the receptor-regulated Smad proteins, Smad1, 5, and 8. Activated receptor-regulated Smad proteins interact with the common partner Smad, Smad4, and translocate to the nucleus, where the Smad complex directly binds defined DNA sequences and regulates target gene expression.2

BMPs were reported to promote survival of human pulmonary arterial endothelial cells (ECs) and circulating endothelial progenitor cells isolated from normal subjects. This antiapoptotic effect induced by the BMP signaling pathway on vascular EC is, in part, because of the decrease in caspase-3 activity.1 CRYAB is a soluble cytoplasmic protein and a member of the small heat shock protein (sHSP) family. CRYAB was identified initially as an abundant structural eye lens protein but seemed later to be expressed in the heart, skeletal muscle, central nervous system, kidney, and lung of mice.4 It is induced by heat shock, proinflammatory cytokines, and oxidative stress and plays a role in cellular growth, migration, differentiation, and development.5,7 CRYAB has chaperone-like properties, binds to both desmin and cytoplasmic actin, and helps to maintain cytoskeletal integrity.6 As in the case of other chaperones and sHSPs, CRYAB can bind to unfolded proteins and can prevent their denaturation and aggregation.9 CRYAB has been shown to protect cells against apoptosis induced by different factors, such as DNA-damaging agents, tumor necrosis factor-α, tumor-necrosis-factor related apoptosis inducing ligand, hydrogen peroxide, etc.10–12 CRYAB is known to protect vascular EC from apoptosis, in part, by binding to caspase-3 and blocking its autoproteolytic maturation.10,13,14 CRYAB is also considered as an oncogenic protein, by protecting cancer cells against apoptosis induced by chemotherapeutic agents.

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by interacting with proapoptotic proteins, or inhibiting caspase-3 activation.\textsuperscript{10,12,15} In cardiomyocytes, it was shown that CRYAB silencing promoted cell death after exposure to H\textsubscript{2}O\textsubscript{2}, and that CRYAB has a protective role against apoptosis, by a mechanism involving translocation of CRYAB into mitochondria and by interacting with different proteins, including voltage-dependent anion channels and caspase-3.\textsuperscript{16}

Here, we show that BMPs induce CRYAB expression by a transcriptional, indirect molecular mechanism, which requires the BMPR2 and ALK1 receptors and the transcription factor Id1. BMPs protect EC from serum starvation and hypoxia-induced apoptosis and the protecting effect is, in part, because of a strong upregulation of the antiapoptotic protein CRYAB. CRYAB silencing promoted cell death after exposure to H\textsubscript{2}O\textsubscript{2}, and CRYAB has a protective role against apoptosis, by a mechanism involving translocation of CRYAB into mitochondria and by interacting with different proteins, including voltage-dependent anion channels and caspase-3.\textsuperscript{16}

We confirmed, by real-time polymerase chain reaction (PCR), the upregulation of CRYAB expression in HMEC-1 treated by 10 ng/mL of BMP4 for 24, 48, and 72 hours (4.3±4.1, 8.6±0.7, and 7.5±1, respectively; Figure 1A). CRYAB mRNA was also increased by other members of the BMP growth factors family, BMP7 (fold increase at 4.8±0.9 at 24 hours, 4.8±1.8 at 48 hours, and 3.4±0.06 at 72 hours) and BMP9 (fold increase at 86.5±6.3 at 24 hours, 25.1±0.8 at 48 hours, and 15.1±1.5 at 72 hours), similar to BMP4 (Figure 1A). In human lung microvascular ECs (HLMEC), BMP7 and BMP9 increased CRYAB expression, whereas BMP4 had no effect on CRYAB expression (Figure 1 in the online-only Data Supplement). BMP9 also induced CRYAB expression in human umbilical vein ECs (HUVEC; Figure I in the online-only Data Supplement). By Western blot, we observed that CRYAB levels were increased strongly in HMEC-1 treated with BMP4, BMP7, and BMP9 compared with control cells (Figure 1B).
BMP treatment, pre-mRNA levels were strongly induced by BMP4 or BMP7 stimulation (6.5±2- and 6.1±1-fold respectively; Figure 2B). These data show that CRYAB is induced at the transcriptional level by BMPs. Altogether, these results indicate that CRYAB induction by BMPs is dependent on a transcriptional indirect mechanism and does require de novo synthesis of proteins.

**CRYAB Uptregulation by BMPs Is Mediated by the BMPR2–ALK1–Id1 Pathway**

We knocked down the expression of BMPR2 using a BMPR2-targeting siRNA (Figure II in the online-only Data Supplement). CRYAB upregulation at the mRNA level by BMPs was nearly abolished in HMEC-1 transfected with BMPR2-directed siRNA compared with those transfected with nonspecific siRNA (19.4±8.9% of control siRNA value) compared with those transfected with nonspecific siRNA (Figure 3A). In contrast, inhibiting ALK3 by siRNA did not significantly decrease CRYAB mRNA upregulation (Figure 3A). Altogether, these results demonstrate that the upregulation of CRYAB expression by BMPs is mediated by BMPR2 and ALK1, but not by ALK3. ALK6 was not tested because it is not expressed in EC (data not shown).

The main intracellular signaling pathway activated by BMPs is the Smad pathway. We observed an induction of Smad1/5/8 phosphorylation after BMP treatment of HMEC-1 in a time-dependent manner concomitantly with BMP-induced CRYAB expression (Figure III in the online-only Data Supplement). We tested the implication of the transcription factors Id1, Id2, and Id3, which are strongly induced by BMPs. After silencing these mRNA targets using specific siRNAs (Figure II in the online-only Data Supplement), we observed that ID1 expression knockdown reduces CRYAB mRNA induction in HMEC-1 by 55±14%, whereas ID2 and ID3 knockdown did not modify significantly this induction (Figure 3B). These data corroborate the actinomycin D and cycloheximide experiments, which suggested the transcription and translation of an inducer factor.

**Role of CRYAB in the Antiapoptotic Effect of BMPs on Serum-Starved HLMEC**

To determine whether the upregulation of CRYAB mRNA by BMPs plays a role in the antiapoptotic effect induced by BMPs on serum-starved HLMEC, we inhibited CRYAB expression using a specific siRNA targeting CRYAB mRNA (Figure II in the online-only Data Supplement). In cells transfected with a nonspecific siRNA, we observed an antiapoptotic effect of BMP7 in serum-free conditions. This effect was abolished in cells whose CRYAB expression is knocked down by a specific siRNA (Figure 4A). This observation was confirmed by flow cytometry because the percentage of annexin V–positive cells is higher when CRYAB-targeting siRNA is transfected in cells treated with BMP7 (6.9±2.2% in siCT-transfected cells versus 13.5±4.52% in siCRYAB-transfected cells) and BMP9 (7.2±2.4% in siCT-transfected cells versus 13.7±4.7% in siCRYAB-transfected cells) compared with cells transfected with a control siRNA (Figure 4B and 4C).

These results suggest that BMPs protect EC from apoptosis, in part, by upregulating the antiapoptotic factor CRYAB.

**Role of BMP and CRYAB in Hypoxia-Induced Apoptosis in HUVEC**

To corroborate results obtained on HLMEC by serum-starvation–induced apoptosis, we used hypoxia as another inducer of apoptosis in EC. HUVEC were cultured in hypoxic conditions (1% O₂). Apoptosis was measured by Hoechst 33342 staining to quantify condensed pyknotic nuclei in apoptotic cells. In hypoxic conditions, BMP9 induced a significant decrease of apoptotic cells in the presence of a control siRNA (41±7%; Figure 5). This antiapoptotic effect of BMP9 was no more detectable in cells transfected with a siRNA targeted on the CRYAB mRNA. Therefore, these results confirm in a different model of apoptosis (hypoxia-induced apoptosis) that CRYAB participates in the antiapoptotic effects of BMPs in EC.
Decreased BMP Activity Correlates With the Downregulation of CRYAB Expression In Vivo

BMP signaling is known to be altered in pulmonary hypertension. Thus, we studied the BMP signaling pathway and cryab expression in the mouse hypoxia model of pulmonary arterial hypertension (PAH). Mice were exposed to chronic hypoxia for 35 days leading to the development of pulmonary hypertension as assessed by hemodynamic parameter measurements (Figure IV in the online-only Data Supplement). In this model, a significant reduction of bmpr2 expression in mouse lung was observed (Figure 6A). In these conditions, we also observed a decrease of Smad 1/5/8 phosphorylation compared with control conditions (Figure 6C and 6D). These results confirmed that the BMP signaling pathway is downregulated in the mouse hypoxia model of PAH in our experimental conditions.

We observed a significant downregulation of the cryab mRNA and protein in the lung of animals exposed to chronic hypoxia compared with those exposed to normal oxygen conditions (Figure 6B, 6C, and 6D). These results show that cryab expression is downregulated in vivo during chronic hypoxia, and this downregulation correlates with the decreased BMP signaling pathway activity.

We analyzed the expression of cryab in the mouse lung by immunofluorescence staining. The expression of cryab in EC of small vessels was shown by double staining with antibodies against the EC-specific Von Willebrandt factor and CRYAB (Figure 6E). We observed that cryab is expressed in the mouse lung in various types of cells but at different levels. In the lung parenchyma, the highest levels of expression are observed in EC, in smooth muscle cells (from vessels and airways) and in bronchiolar epithelial cells (Figure 6E; Figure VI in the online-only Data Supplement). We confirmed the specificity of the CRYAB antibody by performing a parallel hybridization in similar conditions on mouse kidney. The results show specific expression of cryab in kidney vessels in EC and smooth muscle cells (Figure VII in the online-only Data Supplement).

**Discussion**

We demonstrate in this study a cross-talk between the BMP pathway and the sHSP family regulation, namely the strong induction of the CRYAB gene transcription and expression by BMPs. Experiments in vitro favor an indirect mechanism of transcription induction by BMPs because a protein biosynthesis is required, as shown by cycloheximide experiments. Therefore, a transcription factor upregulated by BMPs is likely to be involved, and our results confirm this conclusion because Id1 knockdown strongly, but not totally, inhibits the induction. The absence of total inhibition could be because of either incomplete mRNA knockdown by siRNAs or the involvement of other intermediate transcription factors. Knockdown experiments of intracellular intermediates of BMP signaling show that BMPR2 and ALK1 are required for the induction.

We initially identified the CRYAB transcriptional upregulation by BMPs in an EC line (HMEC-1), and this result was
extended to HLMEC, and HUVEC, 2 types of human primary culture of EC from different origins. However, we found some contrasted effects between BMPs in these cell types because BMP4, which induces CRYAB in HMEC-1, is not efficient in HLMEC, and BMP9 is clearly the most active inducer in all cell types.

We evaluated the physiological effect of BMPs in EC taking into account the known physiological effect of CRYAB. Indeed, CRYAB is a sHSP known to inhibit apoptosis through inhibition of caspase-3 activation. Because BMPs are known to inhibit apoptosis in EC, an effect which contrasts with the documented proapoptotic effects of BMPs during embryogenesis and also in adult vascular smooth muscle cells, we searched for a specific effect of BMPs on EC apoptosis through CRYAB. We observed a strong apoptotic effect induced by serum deprivation of HLMEC, and this effect was antagonized by BMP7 and BMP9 as attested by the lower percentage of annexin V–positive cells measured by flow cytometry after treatment with these peptides. This effect is dependent on the induction of CRYAB expression as the knockdown of CRYAB expression resulted in a nearly complete loss of the BMP anti-apoptotic effect. We confirmed these results using hypoxia as a stress for inducing apoptosis in HUVEC. In this model, BMP9 also significantly decreased apoptosis, an effect suppressed by inhibiting CRYAB expression. To eliminate an indirect effect mediated by VEGF, we measured VEGF mRNA induction by BMP9 in HMEC-1, and we did not observe any induction (Figure V in the online-only Data Supplement).
We looked for a correlation between the activation status of the BMP signaling pathway and bmpr2 expression in pulmonary tissues by investigating the expression and regulation of bmpr2 in the mouse hypoxia model of PAH. In this model, we observed a downregulation of the BMP pathway, as shown by the pSmad1/5/8 decrease after chronic exposure to hypoxia, confirming previously published results. In this experimental condition, bmpr2 is also decreased in the mouse lung as measured by RNA expression and by Western blot. The expression of CRYAB was detected in pulmonary EC, but also localized in other cell types. Therefore, these results show that, in vivo, a BMP signaling decrease is associated with a decrease in bmpr2 expression, which is consistent with the BMP-induced CRYAB regulation found in cultured HMEC-1, HLMEC, and HUVEC.

A few hereditary vascular diseases result from decreased BMP signaling and are linked to genetic mutations abrogating a single allele function of a gene belonging to the BMP pathway. Hereditary hemorrhagic telangiectasia is mainly caused by ALK1 (ACVRL1), Endoglin, and SMAD4 genes mutations, and heritable PAH is linked to BMPR2 mutations but can also complicate ACVRL1-linked hereditary hemorrhagic
telangiectasia. These 4 genes are receptors or coreceptors for the BMP ligands or intermediate signaling molecules. The cross-talk identified in our study between BMP and CRYAB might be of importance for understanding the physiopathology of these diseases because under stress conditions, a deficient BMP signaling might result in increased endothelial apoptosis, at least in part because of decreased CRYAB expression. It has been proposed that, after a stress to pulmonary EC in the lung, the selection of EC resistant to apoptosis and subsequent vascular smooth muscle cell proliferation could explain the remodeling observed in PAH.

The cross-talk between BMP and CRYAB could have implication beyond resistance to apoptosis because CRYAB has wide physiological functions as sHSP. Indeed, CRYAB is known to be involved in resistance to oxidative stress induced by high glucose concentration in EC and to prevent apoptosis in mouse neural cardiomyocytes exposed to H2O2. BMP7 was reported recently to protect mesangial cells from oxidative stress, and it can be speculated that it might be through CRYAB induction.

The BMP-CRYAB cross-talk might also be of importance for the abnormal angiogenesis observed when the BMP signaling pathway is decreased by an ACVR1I or Endoglin inactivating mutation such as in hereditary hemorrhagic telangiectasia because angiogenesis is a balance between vascular cell apoptosis and proliferation. BMP signaling plays a major role on the balance between endothelial tip cells, present at the tip of growing capillaries, and stalk cells, which have a proliferative behavior, form tubes and vessel branching. CRYAB null mice present with decreased retinal vessel density and subtle defects in vascular pattern morphogenesis. After experimental retinopathy, crynab null mice show greater EC apoptosis, reduced neovascular tufts in the retina, and vessel leakage. CRYAB also stabilizes vascular endothelial growth factor and favors neovascularization in retinopathies. Taken together, these results and ours suggest that CRYAB might be one of the BMP targets whose decreased expression can lead to the abnormal angiogenesis observed in hereditary hemorrhagic telangiectasia. We, and others, previously showed a strong inhibition by BMP9 of Apelin gene expression, encoding a peptide highly expressed in endothelial tip cells, which binds to its receptor present on stalk cells, where it promotes cell proliferation and vessel branching through mTOR signaling. Therefore, the expected increase in apelin expression when BMP signaling is decreased might contribute synergistically with CRYAB decrease to the abnormal angiogenesis observed in mutation carriers of the BMP pathway genes.

In conclusion, we have detected a molecular link between the BMP signaling pathway and a chaperone protein acting in different cell processes, in particular, in resistance to apoptosis, and this relationship might play a major role in the physiopathology of vascular diseases linked to BMP signaling deficiency.

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Disclosures
None.

References


We show the strong induction of CRYAB, a chaperone protein which is a small heat shock protein, by bone morphogenetic proteins. This induction is transcriptional indirect and is dependent on BMPR2, ALK1, and the transcription factor Id1. We show that, in vitro, bone morphogenetic protein inhibits endothelial cell apoptosis induced by serum starvation and by hypoxia, and that this inhibition is dependent on CRYAB induction. We also show in the mouse hypoxia model a decrease in the bone morphogenetic protein pathway activity and a decrease in CRYAB expression. We speculate that the CRYAB decrease observed in this model can favor initial apoptosis and secondary endothelial cell proliferation.
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BMPs up-regulate α-B-crystallin in endothelial cells, and that mediates in part their anti-apoptotic effect.

**Reagents.** Recombinant human BMP4, BMP7 and BMP9 were purchased from R&D Systems (Minneapolis, MN). Cycloheximide and Actinomycin D were obtained from Boehringer Ingelheim GmbH (Ingelheim, Germany). Primary antibodies used were: αB-crystallin: SPA-222, for Western blot and SPA-223 for immunofluorescence, (both from Enzo life science, Villeurbanne, France), pSmad1/5/8, SMAD5 (Cell Signaling Technology, Danvers, MA), β-actin (Sigma-Aldrich, St Louis, MO), Von Willebrandt Factor (#ab11713 from Abcam, Cambridge, UK).

**Cell culture.** The human dermal microvascular endothelial cell line HMEC-1 was obtained from Thomas J. Lawley (Emory University, School of Medicine, Atlanta, GA) and grown as previously described. Human lung microvascular endothelial cells (HLMEC) were obtained from Clonetics (Baltimore, MD) and cultured in endothelial cell Basal Medium 2 (EBM-2) supplemented with EGM-2MV Single Quots (Clonetics, Baltimore, MD). HUVEC (Human Umbilical Vascular Endothelial Cell) were extracted from umbilical cord as previously described and cultured in endothelial cell basal medium 2 (EBM-2) supplemented with EGM-2MV (Lonza, Basel Switzerland).

**siRNA transfection.** Synthetic small interfering RNA (siRNA) targeting the human BMPR2 mRNA, CRYAB mRNA, ALK1 mRNA, ALK3 mRNA, Id1 mRNA, Id2 mRNA and Id3 mRNA were purchased from Dharmacon Research (Lafayette, CO). Non-targeting siRNA 2 (Dharmacon #D-001210-02) was used as a non-specific control. SiRNA were transfected using Dharmafect-1 transfection reagent according to manufacturer’s recommendation. After transfection, cells were starved for 24h then treated with BMPs for indicated time before lysis or staining.

**Real-time RT-PCR.** Real-time RT-PCR assay was performed as previously described. Primers used for real-time RT-PCR are indicated below:

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<th>Gene</th>
<th>Primer sequences</th>
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<td>hCRYAB</td>
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<td>RPL32</td>
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Western Blot. Cells were lysed with 100μl of Ripa buffer (25 mM Tris•HCl pH 7.6, 150 mM NaCl, 1% NP-40, 1% sodium deoxycholate, 0.1% SDS) supplemented with Protease Inhibitor Cocktail, Phosphatase Inhibitor Cocktail 1 and 2 (Sigma-Aldrich, St Louis, MO), snap frozen and stored at -80°C until use. Frozen tissues were homogenized in the same lysis buffer, centrifuged for 5 minutes at 14000 rpm at 4°C in a chilled micro centrifuge and the supernatant was stored at -80°C until use. Samples were subjected (20μg of proteins per lane for cell extracts and 50μg of proteins per lane for tissue extracts) to SDS-polyacrylamide gel electrophoresis (PAGE) in Tris-Glycine gels. The proteins were transferred to the PVDF membrane using XCell II Blot Module (Invitrogen, Carlsbad, CA). Membranes were then incubated overnight with corresponding antibodies (CRYAB 1:1000, pSMAD1/5/8 and SMAD5 1:1000, β-actin 1: 40000) followed by an anti-rabbit IgG or anti-mouse IgG secondary antibody conjugated to horse-radish peroxidase (1:40000, 1h; Jackson ImmunoResearch Laboratories, West Grove, PA). Bands were visualized using an enhanced chemiluminescence substrate system ECL+ detection system (GE Healthcare, Buckinghamshire, UK) using either High Performance Chemiluminescence Films or Ettan DIGE Imager (GE Healthcare, Buckinghamshire, UK). The intensity of each band was analyzed by densitometry using ImageJ software or using ImageQuant software (GE Healthcare, Buckinghamshire, UK) respectively.

Transcription assay of the CRYAB gene. Gene transcription was measured by quantifying pre-mRNA as previously described\textsuperscript{4}. CRYAB pre-mRNA was measured by real-time PCR using the following intron-exon primers: upstream 5'-TTGGTCTCACCTAAGGGGA-3' located in intron 1 and downstream 5'-TGTCATCTGATGGGGTCCTCA-3' located in exon 2. CRYAB pre-mRNA expression was normalized to GAPDH expression measured by real-time PCR using the following primers: upstream 5'-GAAGGTGAAGGTCGGAGT-3' placed on exon 2 and downstream 5'-GAAGATGGTGATGGGATTTC-3' placed on exon 4.

Apoptosis assay. HLMCE were plated at a density of 2.5x10\textsuperscript{5} cells/well in 6-well plates for 24h. Cells were then cultured for 48h in complete medium or SF medium, in the presence of BMP7 (200 ng/ml). Cell lysates were tested for apoptosis by measurement of cytoplasmic nucleosomes using a Cell Death Detection ELISA kit (Roche Diagnostics GmbH, Roche Molecular Biochemicals, Mannheim, Germany) according to the manufacturer's recommendation. This assay quantitatively detects histone-associated DNA fragments in the cytoplasmic fraction of cell lysates.

Annexin assay. Apoptosis was assessed in HLMEC by flow cytometry analysis using the Annexin V:FITC Apoptosis Detection Kit II (BD Pharmingen, San Diego, CA) under control or apoptosis-inducing conditions (SF), in the presence or absence of BMPs and siRNA targeting CRYAB mRNA or a non-targeting siRNA. At the end of the treatment period (72h), cells were suspended in Enzyme Free Cell Dissociation Buffer (Invitrogen, Carlsbad, CA) and washed twice with cold PBS. Annexin assay was performed according to the manufacturer's
Fluorescence was measured on a Beckman Coulter XL analyzer (Beckman Coulter, Fullerton, CA).

**Nuclear chromatin condensation assay.** HUVEC were seeded in 6-well plates at a density of 2×10^5 per well, and cultured in normoxia or hypoxia (1%O2) conditions for 24h. Plates were washed twice with PBS, and cells were fixed using 4% paraformaldehyde for 15 min, followed by another two washes of PBS and incubated in 500 µl of Hoechst 33342 solution (5 µg/ml in PBS) for 15 min in the dark. Analysis was performed by fluorescent microscopy. Blue cells were scored as apoptotic or normal based on morphological criteria: small, round, condensed cells were scored as apoptotic, and all other cells were scored as healthy. For each sample, about 1000 cells were scored in a blinded manner.

**Animals.** Eight weeks-old male C57Bl/6 mice (CERJ, Orléans, France) were used for this study and were divided into two groups: a control group exposed to normoxic conditions, and one group exposed to hypoxic conditions (10% O2) in a normobaric hypoxic chamber for 35 days. After 35 days, mice were euthanized immediately after hemodynamic assessment. Lungs were washed in PBS, immediately frozen in liquid nitrogen and stored at -80° before RNA and protein extraction, or inflated with diluted OCT (1:2 in PBS, 2ml) and frozen in isopentane dipped into liquid nitrogen for immunofluorescence analysis. Mice were treated in accordance with our institutional guidelines (Ministère de l’Agriculture, France; authorization 75-1206) and conformed to the Directive 2010/63/EU of the European Parliament.

**Immunofluorescence staining and microscopy.** Frozen mouse lung sections (6 µm) were fixed in 4% PFA for 15 minutes and washed in PBS. Sections were blocked in 3% BSA for 1h at room temperature followed by incubation with primary antibodies (SPA-223 1:100, VWF 1:1000) in blocking solution over night at 4°C. Sections were washed in PBS and incubated with Alexa-conjugated secondary antibodies (Invitrogen, Carlsbad, CA) in PBS buffer for 1h, followed by nuclear staining with DAPI. Sections were mounted using Fluorescent Mounting Medium (Dako, Denmark A/S). Slices were analyzed using an Olympus XL microscope and images were then deconvoluted, using MetaMorph softwear (MDS Analytical Technologies, Downingtown, PA).

**Statistical Analysis-** Statistical analysis was performed with XLSTAT (Addinsoft, Paris, France) using a one-way analysis of variance for all data, followed by a Student Newman-Keuls post-hoc analysis for multiple tests. Pairwise comparisons were analysed using the non parametric Mann-Whitney test. The null hypothesis was rejected for p<0.05. Data are expressed as mean ± S.D.

**References**


Supplementary Figure I. BMPs up-regulate CRYAB in endothelial cells. (A) Serum-restricted HLMEC were stimulated with 200ng/ml of BMP7 or BMP9 for 24, 48 or 72h. (B) Serum-restricted HUVEC were stimulated with 10ng/ml of BMP9 for 24h. Expression of CRYAB was quantified by real-time RT-PCR analysis. The relative CRYAB mRNA level was normalized to RPL32 mRNA level and expressed as a fold change relative to untreated cells. Results are mean ± S.D. values of 3 independent experiments. * P<0.05 relative to untreated cells.
Supplementary figure II. Knock-down expression of targeted genes by specific siRNA. HMEC-1 were transfected with a non specific siRNA (Si CT) or specific siRNA targeting ALK1, ALK3, BMPR2, CRYAB, ID1, ID2 or ID3. Each specific gene expression was quantified by real time RT-PCR analysis. The targeted gene mRNA level was normalized to GAPDH levels and expressed as a fold change of its expression in untreated cells.
Supplementary figure III. SMAD pathway is activated by BMPs in HMEC-1. Serum-restricted HMEC-1 were stimulated with 10ng/ml of BMP4, BMP7 or BMP9 for 24, 48 or 72h. (A) Protein extracts were analyzed by Western blot using specific phospho- SMAD1/5/8 and SMAD5 antibodies. (B) Densitometric analysis of SMAD1/5/8 activation. SMAD activation is defined as the ratio between the phospho-SMAD 1/5/8 signal and SMAD5 signal.
Supplementary figure IV. Hemodynamic parameters in mice exposed to normoxia or hypoxia for 35 days. (A) Hematocrit. (B) Right Ventricule Systolic Pressure (RVSP) (C) Fulton index.
Supplementary Figure V. Absence of VEGF mRNA induction in HMEC-1 treated with BMP9. Serum-restricted HMEC-1 were stimulated with 10 ng/ml BMP9 for 24h. Expression of VEGF mRNA was quantified by real-time RT-PCR analysis. The relative VEGFmRNA level was normalized to GAPDH mRNA level and expressed as a fold change relative to untreated cells. Results are mean ± S.D. values of 5 independent experiments. ns, not significant.
Supplementary Figure VI CRYAB expression in mouse lung exposed to normoxia or hypoxia. (E) Immunofluorescence analysis of endothelial cells (visualized by Von Willebrandt factor (VWF) -green) shows CRYAB expression (in red) in endothelial cells of lung vessels but also in vascular and airway smooth muscle cells and in bronchiolar (br) epithelial cells normal mouse lung; nuclei are stained with DAPI; Expression of CRYAB in endothelial cells of small muscularized (mv) and non muscularized (nmv) lung vessels is indicated by arrows. Scale bar 50µm.
Supplementary Figure VII. CRYAB expression in mouse kidney vessels showing the specificity of the Cryab antibody. Immunofluorescence analysis of endothelial cells (visualized by Von Willebrandt factor (VWF) - green) shows CRYAB expression (in red) in endothelial cells of kidney vessels (v) but not in other kidney cells. Scale bar 50µm.