Dynamic Tyrosine Kinase-Regulated Signaling and Actin Polymerisation Mediate Aggregate Stability Under Shear

Jocelyn M. Auger, Steve P. Watson

Objective—Aggregate formation on collagen at arteriolar rates of shear is mediated by coordinated signaling between tyrosine kinase–linked and G protein–coupled receptors. We have investigated the role of these receptors and the actin cytoskeleton in maintaining aggregate stability under shear.

Methods and Results—Platelet aggregates are rapidly formed when blood is flowed over collagen at 1000 s⁻¹ and remain stable over 20 minutes. A novel fibrin-independent mechanism of retraction against the direction of flow occurs at the aggregate front and recruits platelets into the main aggregate. Stable aggregates are not observed in the presence of cytochalasin D, which blocks de novo actin polymerization. When exposed to the Src family kinase inhibitor, PD0173952, preformed aggregates spread in the direction of flow and rounded platelets appear within the aggregate body and are lost in the direction of flow. A similar set of observations is observed in the presence of latrunculin A, which disrupts preexisting actin filaments, but not in the combined presence of inhibitors of ADP and thromboxane A₂ formation.

Conclusions—Maintenance of stable aggregates at high shear is a dynamic process mediated by Src kinases and actin polymerization. These signals maintain aggregates in a compact structure and prevent continuous streaming of platelets. (Arterioscler Thromb Vasc Biol. 2008;28:1499-1504)

Key Words: platelet aggregation ■ collagen ■ cytoskeleton ■ Src kinase ■ shear

Collagen is a key component of the vascular basement membrane that surrounds blood vessels and the deeper vessel wall. On injury, the interaction of platelets with collagen plays a critical role in initiating thrombus formation and preventing excessive blood loss. Collagen is also a major component of atherosclerotic plaques and mediates the massive platelet activation that occurs on plaque rupture, leading to thrombotic disorders such as myocardial infarction and stroke.

The events underlying aggregate formation on collagen at arteriolar rates of shear in vitro have been studied extensively. Platelet tethering or capture is mediated by binding of collagen-adherent von Willebrand factor (vWF) to the glycoprotein (GP) Ib-IX-V complex. Subsequent activation via the collagen-adherent von Willebrand factor (vWF) to the glyco-

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Methods and Materials

Collection of blood, fluorescent labeling with DiOC6, and measurement of aggregate formation on collagen at 1000 s⁻¹ was as described. Measurement of protein tyrosine phosphorylation in aggregates was achieved by SDS-PAGE and Western blotting with the antiphosphotyrosine antibody, 4G10. Image analysis was performed using ImagePro Software and confocal microscopy. Results are from a minimum of 3 experiments performed on different donors. For more extensive procedures, see the online supplement, available at http://atvb.ahajournals.org.

Results

Identification of a Fibrin-Independent Mechanism of Aggregate Retraction on Collagen at Arteriolar Shear

When P-PACK-anticoagulated human blood is flowed over collagen at 1000 s⁻¹, which is representative of an intermediate arteriolar rate of shear, platelets adhere to the fibers and form aggregates in a time-dependent manner. This process can be followed in real-time by fluorescently labeling the platelets as illustrated in Figure 1a (i). “Islands” of aggregated platelets can be detected within 60s of blood flow and increase steadily in size over time. These aggregates are stable when washed for several minutes with Tyrode’s buffer (Figure 1a, ii).

Real-time monitoring of aggregate formation using fluorescently labeled platelets reveals retraction of platelets at the aggregate front into the main body of the aggregate, against the direction of blood flow (supplemental Movie I). Aggregate retraction was observed in all 22 donors used in this study (Figure 1b, i and ii). Moreover, all newly formed aggregates, independent of size and shape, undergo retraction against the direction of flow. Retraction is most marked within the first 30 seconds of recording but continues for several minutes (Figure 1b, ii). The degree of retraction over 2 minutes was calculated as 30.7±9.8% (SD; n=22) of the aggregate size, 15 seconds after the onset of formation (Figure 1b, ii). Aggregate retraction can also be seen using DIC microscopy (supplemental Movie II), demonstrating that it is not a consequence of fluorescent labeling. The quality of the DIC recording is limited by the presence of erythrocytes in the blood stream, and so fluorescent images were used for quantitative analyses.

The presence of the anticoagulant thrombin inhibitor P-PACK suggests that the process of retraction is unrelated to fibrin formation. In line with this, we have been unable to detect fibrin formation in aggregates using a specific antibody (not shown). Furthermore, aggregate formation and retraction was not significantly altered in the presence of an inhibitor of fibrin polymerization, GPRP (n=3), relative to controls (not shown). GPRP was used in these studies at a concentration that prevents thrombin-induced coagulation in platelet-rich plasma (not shown).

Treatment of blood with the actin polymerization inhibitor, cytochalasin D, inhibits aggregate retraction and prevents formation of stable aggregates (Figure 1a and supplemental Movie III). In the presence of cytochalasin D, platelets are able to adhere to the collagen fibers and generate small aggregates, but rapidly embolize in the direction of flow with no evidence of retraction (n=10). Thus, after several minutes of rinsing with buffer, subsequent to flowing over collagen, there is a high level of adhesion of single unspread platelets and small groups of no more than 2 to 3 platelets. In comparison, the Src kinase inhibitor, PD0173952, or a combination of indomethacin and apyrase, completely block aggregate formation as measured in real-time. This is illustrated by the DIC images taken at the end of the recording in Figure 1a (ii).

These results demonstrate that actin polymerization is essential for formation of stable aggregates at an arteriolar rate of shear and reveal a novel mechanism of aggregate retraction at the aggregate front that may contribute to aggregate stability.
Sustained Src Kinase Signaling Is Required for Maintenance of Aggregate Stability

We have previously shown that aggregate formation on collagen is dependent on Src kinases, because when whole blood is treated with a Src kinase inhibitor before flow over collagen, aggregate formation is abolished (Figure 1a, ii). This observation, however, does not address whether continuous signaling through Src kinases is required to maintain stability of newly formed aggregates, which is of particular significance in light of the observation that signaling through the collagen receptor GPVI is sustained.11 To address this, blood was flowed over collagen for 4 minutes before washing with Tyrode buffer for 5 minutes (to allow clear visualization of aggregates using DIC microscopy as well as analysis of protein phosphorylation), and then washing with Tyrode buffer containing either vehicle (0.1% DMSO) or the Src kinase inhibitor PD0173952 (25 μmol/L). The above changes were analyzed quantitatively at the end of the 20-minute perfusion by measurement of surface area, thrombus height, platelet number, and number of visibly detaching platelets. PD017395 induced a significant (P<0.05) increase in surface area of the aggregates of approximately 30% compared with the controls Figure 3ci, although surprisingly there was no significant difference in the amount of platelets as measured by Western blotting for the integrin subunit, αIIb, or change in height of the aggregates (Figure 3cii and iii). These results can be accounted for by the analysis of the DIC movies, which reveals streaming away of approximately 15 to 20 rounded platelets per field/focal plan of view during 10 minutes rinsing in the presence of the inhibitor, which is similar to the number lost in controls. This represents an exceedingly low proportion of the number of platelets that are present in the aggregates. The results therefore indicate that the increase in surface area represents a looser packing of platelets in the horizontal axis and thereby emphasize the role of Src kinase in maintaining the compact structure of the aggregates over time.

Sustained Secondary Mediator Signaling Is Not Required for Maintenance of Aggregate Stability

Several studies have demonstrated that the feedback agonists, ADP and TxA2, play a critical role in the formation of platelet aggregates on collagen at arteriolar rates of shear. In confirmation of this, we have observed that the presence of apyrase and indomethacin in whole blood before flow over collagen leads to abolition of aggregate formation (Figure 1a, v). To investigate whether ADP and TxA2 are required for maintenance of the compact aggregate structure, we perfused newly formed aggregates on collagen with buffer containing apyrase and indomethacin for 20 minutes. The inclusion of the two inhibitors had no significant effect on the appearance of
single platelets at the edge of the aggregates (Figure 3a and supplemental Movie VI) or on platelet surface area and platelet number (supplemental Figure Ia through Ic). Analysis of DIC movies reveals no difference in the number of detachment platelets compared with the control (15 to 20 platelets per field of view over a period of 10 minutes). Moreover, a similar set of results was obtained after perfusion with the P2Y₁₂ antagonist AR-C69931MX (10⁻⁵ M) & apyrase (2U/ml). Band intensity was measured by densitometry and levels of adherent platelets were calculated, relative to control. A representative Western blot is shown. n=3–5. (iii) Adherent platelets were imaged by confocal microscopy and stacks were used to calculate aggregate height. n=3, *P<0.05.

**Figure 3.** Inhibition of Src kinases or actin polymerization leads to instability in preformed aggregates on collagen. Blood was flowed over collagen at 1000 s⁻¹ for 4 minutes, and aggregates were rinsed with Tyrode for 5 minutes and then for further 20 minutes in the presence of inhibitors as described in Figure 2. a, DIC images (of a single field of view) were recorded during rinsing. b, DIC images of adherent platelets were recorded after 20 minutes of rinse (higher magnification). c, (i) Fluorescent images, recorded subsequent to rinse, were used to calculate surface area coverage. n=3–7, *P<0.05. (ii) Platelet lysates from capillaries were western blotted with anti-GPⅡb mAb. Band intensity was measured by densitometry and levels of adherent platelets were calculated, relative to control. A representative Western blot is shown. n=3–5. (iii) Adherent platelets were imaged by confocal microscopy and stacks were used to calculate aggregate height. n=3, *P<0.05.

### The Actin Cytoskeleton Contributes to Aggregate Stability

The contribution of the actin cytoskeleton to the stability of newly formed aggregates was investigated using the above protocol and the inhibitors cytochalasin D and latrunculin A. These 2 inhibitors differ in that the former prevents further growth of existing actin filaments but does not affect their breakdown, whereas the latter promotes a more rapid breakdown of filamentous actin through binding to monomeric actin. The rapid streaming of single rounded platelets from the newly formed aggregates in the direction of flow can be seen throughout the 20 minutes wash in the presence of latrunculin A (supplemental Movie VII) whereas this is less apparent with cytochalasin D (not shown) and minimal in controls. Additionally, many of these platelets remain attached to the aggregates by long tethers before either release or, occasionally, their return to the main body of the aggregate as illustrated in supplemental Movie VII and in Figure 3b (iii). These tethers are similar in appearance to those that have been previously described for platelets flowing over vWF, which have also been shown to increase on inhibition of actin polymerization. Analysis of DIC movies reveals approximately 4 times as many platelets visibly detaching from the rinsing aggregates over the control (60 to 80 per 10 minutes rinse with latrunculin A), although a significant number of these can be seen to reaggregate downstream of their point of release. The relatively low number of released platelets and the fact that a proportion of these are caught by aggregates lower down in the direction of flow accounts for the lack of a significant change in the overall platelet mass as measured by Western blotting for integrin Ⅱb (supplemental Figure Ic). In line with this, there was no increase in surface area (supplemental Figure Ia) despite the change in shape. However, analysis of aggregate height revealed a significant increase in the presence of latrunculin A of approximately 50% of the control size (supplemental Figure Ic), representing aggregate destabilization and spreading out of platelets in the vertical plane (P<0.05). However, in contrast to the effects of PD0173952, latrunculin A did not inhibit tyrosine phosphorylation of platelet proteins, indicating that the collagen signaling pathway was unaffected by the loss of actin polymerization (Figure 2).

### Discussion

The current study demonstrates that maintenance of stable compact platelet aggregates on collagen at arteriolar rates of flow is a highly dynamic process that is regulated by Src family kinases and the actin cytoskeleton, but not by the secondary mediators, ADP and thromboxanes. This reveals a fundamental difference between the mechanisms that mediate aggregate formation and those that maintain ag-
aggregate stability at arteriolar rates of flow. In addition, the present study reports the novel observation that platelets captured at the front of an aggregate are brought into the main body by a process that we have termed “aggregate retraction.” This retraction is mediated by the actin cytoskeleton and is independent of fibrin formation. Continuous signaling through Src family kinases and dynamic actin polymerization are required to limit the loss of platelets from the front of the aggregate and maintain platelets in a fully spread morphology. These events may play a crucial role in vivo in limiting the loss of platelets from aggregates in the arteriolar system.

An inhibitor of Src kinases or an agent that promotes rapid disruption of the actin cytoskeleton promotes the appearance of single rounded platelets at the edge of the aggregate and causes spreading of the aggregate in the direction of flow or into the capillary lumen. This suggests loosening of the packing of the aggregate, presumably attributable to loss in strength of platelet-platelet interactions. This could be a result of reduced activation of integrin αIIbβ3 (or other receptors that mediate attachment of platelets to each other), or attributable to a change in platelet morphology forcing a greater distance between adjacent platelets in an aggregate. The dependence on Src family kinases indicates that these changes are mediated by loss of tyrosine kinase-dependent signals from the major GP receptors, most notably integrin αIIbβ3 and GPVI. The stabilizing effects of Src kinases could also involve α2β1, which has previously been shown to be important in the early formation of stable aggregates on collagen under shear. These findings are consistent with the recent observation that dynamic signaling through Src kinases is required for sustained lamellipodia formation on collagen under static conditions. Interestingly, signaling by ADP and TxA2 is dispensable for aggregate stability, most likely because of emptying of dense granules and reduced activation of phospholipase A2, respectively.

We have previously reported that Rac1-deficient mouse platelets, which are unable to form lamellipodia, generate unstable aggregates that rapidly embolise at arteriolar rates of shear. The current study demonstrates that blockade of actin polymerization (and the corresponding inability of platelets to spread) leads to a loss in aggregate retraction and subsequent aggregate instability, raising the possibility that lamellipodia formation plays a critical role in the retraction process. The observation that loss of aggregate stability in the presence of either Src kinase blockade or actin disruption is associated with retraction of lamellipodia and the appearance of single rounded platelets leads us to speculate that Src kinases contribute to aggregate stability through the actin cytoskeleton, especially bearing in mind the wealth of evidence for direct regulation of actin polymerization by integrin αIIbβ3 and GPVI.

Several group have reported stable thrombus formation at arteriolar rates of shear in vitro, as observed in the present study. On the other hand, over the past few years, the use of various in vivo thrombosis models has revealed unstable aggregate formation in the absence of many platelet receptors or receptor mutants, including the α2β1-adrenergic receptor, SLAM receptors, CD40L, and the diYF mutant of the integrin β3 subunit. In many cases, the molecular mechanism underlying the thrombus instability is unknown and could even be mediated through a nonplatelet mechanism. For example, in the case of the tetraspanin TSSC6 and Gas6 receptors, aggregate instability may be related to involvement of these proteins in fibrin clot retraction. On the other hand, the discovery that absence of either coagulation factor XI or XII leads to increased embolization in vivo reinforces the importance of coagulation in thrombus stability. A recent study by Goto et al demonstrated that inhibition of P2Y1 or P2Y12 receptors leads to breakup of the platelet aggregate. The probable explanation for the difference with the findings of the present current study is that Goto et al administered the two P2 receptor antagonists much earlier after aggregate formation, when presumably dense granules are continuing to release ADP. Together these studies emphasize the importance of understanding the molecular mechanisms that underlie aggregate stability, the way in which these change, and their interaction with the coagulation system.

An important consideration based on the present study is whether inhibiting Src kinases or actin polymerization in vivo could facilitate the breakup of life-threatening thrombi in occluded arteries and arterioles, without generating emboli that themselves would cause major clinical problems. Certainly, the streaming away of individual rounded platelets is unlikely to have a major effect on blood vessel patency, but on the other hand, the rate of dissolution would appear to be too slow to have a significant effect on thrombus size over short periods of time, although it could be of potential benefit over longer periods. Promoting thrombus breakdown in this way could also be of benefit in cases of small thrombi that have formed on damaged vessels, including atherosclerotic plaques, and which in turn might lead to the generation of larger, life-threatening thrombi.

**Note Added During Revision**

During the course of revision of this study, Ono et al also described fibrin-independent contraction in platelets and further demonstrated that this also occurs in vivo.

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**Disclosures**

None.

**References**


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**Methods and Materials**

**Materials**

Fibrillar-type I Horm collagen was from Nycomed UK (Bucks, UK). PD0173952 was a gift from Pfizer Global Research and Development (Ann Arbor, MI, USA). GPRP (Gly-Pro-Arg-Gly) was from Chromatide (Runcorn, UK). AR-C69931MX (Cangrelor) was from Omega PharmaServices, Inc. (Milford, MA, USA). DiOC₆ (3,3’-dihexyloxacarbocyanine iodide) and PP1 were from Invitrogen (Paisley, UK). P-PACK (D-Phe-Pro-Arg-chloromethylketone, HCl) and AEBSF were from Merck Biosciences Ltd (Nottingham, UK). Anti-αIIb mAb, SZ22, was from Beckman Coulter (Bucks, UK). Anti-phosphotyrosine mAb, 4G10, was from Millipore (Watford, UK). Other materials were from Sigma (Poole, UK) or previously described sources [19, 20].

**Platelet adhesion on collagen under shear**

Human blood, from subjects who gave informed consent, was collected into 40μM P-PACK and further supplemented with 10μM P-PACK every two hours. All work was carried out in accordance to Birmingham University guidelines under approval of the institutional review committee. Glass capillary tubes (Camlab, Cambridge, UK) were coated with 100μg/ml Horm collagen for 1h before blocking with phosphate-buffered saline containing 5mg/ml BSA for 1h at room temperature. Capillaries were mounted onto microscope slides, connected to silicone tubing at both ends in a 37°C chamber and flushed with Tyrode’s buffer (134mM NaCl, 2.9mM KCl, 0.34mM...
Na$_2$HPO$_4$.12H$_2$O, 12mM NaHCO$_3$, 20mM HEPES, 1mM MgCl$_2$, 5mM glucose, pH 7.3). Whole blood, pre-incubated at 37°C for 10min, was perfused through the capillaries for 4min at a wall shear rate of 1000s$^{-1}$ using a syringe pump (Harvard Apparatus Ltd, Kent, UK), followed by washing for 5min at the same flow rate with Tyrode’s buffer to remove non-adherent platelets and other blood cells. Stably adherent platelets and aggregates were rinsed with Tyrode’s buffer containing various inhibitors or vehicle for 20min, at the same flow rate. Images of adherent platelets were recorded before, during and after flow using DIC optics on a Zeiss Axiovert 200M microscope (Herts, UK) equipped with a Hamamatsu Orca 285 digital camera (Hamamatsu Photonics UK Ltd, Herts, UK). In some experiments, blood was pre-incubated with DiOC$_6$ (2μM) for 10min at 37°C prior to perfusion over collagen. Fluorescent images were recorded either during the 4min blood flow (after prefocussing on the collagen fibres and maintaining this focal plane throughout the recording), or before and after the 20min rinse with a Leica DM IRB microscope (Milton Keynes, UK) equipped with a CoolSNAP ES digital camera (Photometrics UK, Bucks, UK). Adherent platelets were fixed with 10% formalin for 30min and imaged using a Leica DM IRE2 confocal microscope.

Platelet surface area coverage was calculated from fluorescent images using ImagePro software (Media Cybernetics Europe, Bucks, UK). A minimum of 10 different fields of view were averaged per experiment. Confocal stacks, captured at 0.5μm intervals, were used to quantify the height of aggregates within a field of view. The maximum height of every aggregate fully visible within a field of view of 240x240μm (approximately 20 aggregates per field) from each of 5 fields was averaged. Aggregate retraction was quantified in all experiments by measuring the distance
moved by a single platelet that had become newly attached to the aggregate front. For these analyses, aggregates were allowed to form for approximately 15 sec and the analysis was then performed on the outermost platelet. Measurements were taken from all aggregates within a field of view, excluding those on the very edge.

**Analysis of protein concentration and phosphorylation**

Blood was flowed over collagen and adherent platelets were washed with Tyrode’s buffer for 20 min as described above. Adherent platelets were then lysed by passing 50 μl NP-40 lysis buffer (300 mM NaCl, 20 mM Tris-HCl, 2 mM EGTA, 2 mM EDTA, 2 mM Na₃VO₄, 1 mM AEBSF, 10 μg/ml leupeptin, 10 μg/ml aprotinin, 1 μg/ml pepstatin, 2% NP-40) through the capillary. For phosphorylation, care was taken to avoid air passing through the capillary prior to lysis.

To quantify the level of platelet adhesion on capillaries, lysates were resolved on 1D 10% SDS PAGE gels, transferred to PVDF membranes and immunoblotted with anti-GPIIb antibody SZ22, using an immunoblotting method described previously [19, 21]. SZ22 was used at 1:500 dilution. Very low blot exposures were taken (1s, 2s and 5s) to avoid overexposure on the film. Densitometry was used to compare the mean band intensity for each lysate, using Quantity One Software (BioRad, Hemel Hempstead, UK). To investigate tyrosine phosphorylation of adherent platelets, lysates were resolved on SDS PAGE gels and transferred to PVDF, then blotted with anti-phosphotyrosine antibody 4G10, at 1:1000 dilution. Membranes were stripped as previously described [20], then reprobed with anti-actin antibody AC-40 at 1:500 dilution.
**Statistical Analysis**

All experiments were performed at least 3 times and data are presented as geometric mean±standard error of the mean (SEM). Statistical analysis was performed using a Mann-Whitney test for unpaired values with GraphPad Prism software (GraphPad Software Inc, San Diego, CA, USA). P values less than 0.05 were considered significant.
Supplementary Figure Legends

Supplementary Figure I – Inhibition of Src kinases or actin polymerisation leads to increase in surface coverage and aggregate height, respectively

Blood was flowed over collagen at 1000s\(^{-1}\) for 4min and aggregates rinsed with Tyrode’s for 5min. Aggregates were further rinsed with Tyrode’s containing 0.1% DMSO (control), PD0173952 (25\(\mu\)M), indomethacin (10\(\mu\)M) & apyrase (2U/ml), or latrunculin A (3\(\mu\)M) for 20min. Lysates from capillaries were western blotted for phosphotyrosine (upper) and reprobed for actin (lower).

(a) Fluorescent images, recorded subsequent to rinse, were used to calculate surface area coverage. n=3-7, *P<0.05.

(b) Platelet lysates from capillaries were western blotted with anti-GPIIb mAb. Band intensity was measured by densitometry and levels of adherent platelets were calculated, relative to control. A representative Western blot is shown. n=3-5.

(c) Adherent platelets were imaged by confocal microscopy and stacks were used to calculate aggregate height. n=3, *P<0.05.
Supplementary figure I

(a) % surface area coverage

- Control
- PD0173952
- Indo & apyrase
- Lat A

(b) Relative intensity (% control)

- Control
- PD0173952
- Indo & apyrase
- Lat A

100 103.8 104.5 83.1

(c) Aggregate height (μm)

- Control
- PD0173952
- Indo & apyrase
- Lat A

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Supplementary Movie Legends

**Supplementary movie I – Aggregate retraction on collagen visualised using fluorescently-labelled platelets**

Blood was pre-incubated with 2μM DiOC₆, 0.1% DMSO and 1% PBS for 10min and then flowed over collagen at a shear rate of 1000s⁻¹ for 4min. Real time adhesion and aggregate formation was monitored via recording fluorescent images (of a single field of view) at a rate of 30 frames per minute. Time from start of blood flow is shown. Image size is 100x100μm. Flow is from right to left and is shown by the arrow. The movie is representative of results from 22 donors.

**Supplementary movie II – Aggregate retraction on collagen visualised using DIC microscopy**

Blood was flowed over collagen at a shear rate of 1000s⁻¹ for 4min. Real time adhesion and aggregate formation was monitored using DIC microscopy at a rate of 30 frames per minute. Time from start of blood flow is indicated. Image size is 100x100μm. Flow is from right to left and is shown by the arrow. The movie is representative of results from 3 donors.

**Supplementary movie III – Aggregate retraction on collagen in the presence of cytochalasin D**

Blood was pre-incubated with 2μM DiOC₆, 50μM cytochalasin D and 1% PBS for 10min and then flowed over collagen at a shear rate of 1000s⁻¹ for 4min. Real time adhesion and aggregate formation was monitored via recording fluorescent images (of a single field of view) at a rate of 30 frames per minute. Time from start of blood
flow is indicated in the bottom left corner. Image size is 100x100μm. Flow is from right to left and is shown by the arrow. The movie is representative of results from 3 donors.

**Supplementary movies IV-VII – Aggregate stability on collagen**

Blood was flowed over collagen at a shear rate of 1000s⁻¹ for 4min and aggregates were rinsed with Tyrode’s buffer for 5min. Stable aggregates on collagen were rinsed with Tyrode’s containing:

**Avi IV**: 0.1% DMSO,

**Avi V**: 25μM PD0173952,

**Avi VI**: 10μM indomethacin & 2U/ml apyrase or

**Avi VII**: 3μM latrunculin A.

A sequence of DIC images (of a single field of view) were recorded during rinse, at a rate of 60 frames per minute. The first 10min of rinsing is shown. Images size is 75x75μm. Flow is from right to left.