Mouse CD36 Has Opposite Effects on LDL and Oxidized LDL Metabolism In Vivo

Vilayphone Luangrath, Mathieu R. Brodeur, David Rhainds, Louise Brissette

Objective—The cluster of differentiation-36 (CD36) is a multifunctional protein which is recognized for its in vitro ability to take up oxidized low-density lipoproteins (oxLDL) in macrophages and is therefore considered atherogenic. It also binds LDL. Our objective was to define the physiological role of CD36 in both native LDL and oxLDL metabolism in mice.

Methods and Results—Clearance studies of labeled LDL and oxLDL were conducted in wild-type, CD36 knockout (KO), scavenger receptor class B, type I (SR-BI) KO, and SR-BI/CD36 double KO mice. We found that CD36 impedes the disappearance of native LDL and favors that of oxLDL. This was confirmed by association and degradation assays with primary cultures of hepatic cells from wild-type and CD36 KO mice. In addition, our in vivo work indicates that neither SR-BI nor CD36 plays a significant role in cholesteryl esters (CE) selective uptake (SU) from oxLDL, whereas CD36, in absence of SR-BI, can selectively take CE from LDL.

Conclusion—Our investigation showed for the first time that CD36 plays a significant role in oxLDL uptake in vivo in the mouse. As CD36 also retards LDL clearance, its atherogenic character may also relate to its negative effect on LDL catabolism. (Arterioscler Thromb Vasc Biol. 2008;28:1290-1295)

Key Words: CD36 ■ SR-BI ■ LDL ■ mouse ■ cholesterol

It is well established that high levels of plasma cholesterol associated with LDL increase the risk of developing atherosclerosis. The level of LDL in blood circulation is related to the synthesis rate of their precursors and their uptake by the liver. A large part occurs through LDL-receptors (LDLR) which take up the entire LDL particles and lead to their complete degradation, a mechanism referred to as the holoparticle pathway. LDL can also selectively transfer their CE without concomitant degradation of their apolipoproteins, a process termed selective uptake (SU).

SR-BI is a cell surface receptor recognized for its ability to selectively take up CE from high-density lipoproteins (HDL) and LDL. Our in vivo studies showed that ablation of SR-BI expression in mice leads to the complete loss of LDL-CE SU early after injection. However, after an hour, an acceleration of CE disappearance was obvious in SR-BI KO mice, indicating that a SR-BI-independent pathway can rescue SR-BI deficiency. A potential candidate for the SR-BI-independent pathway is CD36, another class B scavenger receptor. This is supported by studies showing that COS cells overexpressing either SR-BI or CD36 are equipotent toward LDL-CE uptake. However the physiological importance of CD36 in LDL-CE SU remains to be established in vivo.

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LDL can undergo oxidative modification, and these are detectable in circulation. Many studies have used standard conditions generating fully oxidized LDL (oxLDL), which we refer simply as to oxLDL. Although oxLDL cannot interact with LDLr, it is known that injected oxLDL disappear faster than native LDL in rodents and that this uptake involves the liver. In vivo studies by Ling et al showed that the scavenger receptor class A type I/II (SR-A) is not implicated. Endemann et al have revealed the importance of CD36 as an oxLDL receptor in vitro. Moreover we have demonstrated that CD36 is partially responsible for oxLDL uptake in the human hepatic parenchymal cell model: HepG2. However, the role of CD36 in oxLDL clearance in vivo was never addressed. Inasmuch as it is generally accepted that mildly oxLDL are rather present in blood circulation, these particles also deserve consideration.

Little work was conducted on CE-SU from oxLDL. Whereas our group has reported this pathway in HepG2 cells, using SR-BI-transfected Chinese hamster ovary (CHO) cells, Gillotte-Taylor et al did not, but demonstrated greater oxLDL degradation compared to control cells. Recently, we have shown by in situ experimentations that both mouse
parenchymal and nonparenchymal liver cells selectively take CE from mildly oxLDL and oxLDL. However, we could determine a role of SR-BI only for oxLDL by nonparenchymal cells. Thus another receptor or pathway needs to be defined for mildly oxLDL-CE SU in both parenchymal and nonparenchymal cells as well as for parenchymal cell CE-SU from oxLDL.

The aim of this study was to define the role of CD36 in native LDL, mildly oxLDL, and oxLDL holoparticle and CE-selective uptake. Clearance studies were therefore conducted with CD36 KO and wild-type mice. Our results show that CD36 impedes the disappearance of native LDL and favors that of mildly oxLDL and oxLDL. Also, our in vivo studies reveal that neither SR-BI nor CD36 plays a significant role in oxLDL-CE SU, whereas CD36 significantly shows LDL-CE SU activity in absence of SR-BI.

Methods

Animals

CD36 wild-type or deficient mice (backcrossed 7 times to C57BL/6J mice) were obtained from Dr. Maria Febbraio (Cleveland, Ohio). Heterozygous B6/129S-Strb+/+ breeding pairs (backcrossed 8 times to C57BL/6J mice) were obtained from Jackson Labs (Bar Harbor, Me). For breeding and genotyping protocols please see supplemental materials (available online at http://atvb.ahajournals.org). Six- to 12-week-old male mice were used. This study was conducted according to protocols approved by the Animal Care Use Committee of Universite du Quebec a Montreal.

Lipoprotein Profile and Cholesterol Assay

Three mice of all genotypes were fasted for 18 hours and bled through their vena cava. Lipoprotein fractionation of pooled plasma (400 μL) was accomplished by fast protein liquid (FPLC) separation on a Superose 6 column HR13/50 (GE Healthcare). Total cholesterol, triglycerides, as well as free fatty acids were measured with commercial kits (Wako, Richmond, Va).

Lipoprotein Isolation, Modification and Labeling

Human LDL (density 1.025 to 1.063 g/mL) were isolated from plasma as described in. oxLDL were prepared as in Lougheed and Steinbrecher. oxLDL typically resulted in a 2.8-fold increase in the electrophoretic mobility relative to native LDL on 0.5% agarose/barbital gels. Mildly oxLDL were obtained by incubation under similar conditions but for 4 hours and show a 1.5-fold increase in the electrophoretic mobility relative to native LDL. The 3 types of LDL were labeled with iodine-125 and in [3H]-cholesteryl oleoyl ether (CEt) as described previously. Detailed characterization is available in supplemental Table I, and supplemental Figures I and II. Overall, labeling does not change the properties of LDL particles.

Serum Decay and Cellular Distribution of LDL and Oxidized LDL in Liver

Mice were injected by the tail vein with a bolus of human LDL (either native or mildly oxLDL or oxLDL) containing 480 μg of nonradiolabeled lipoproteins and 20 μg of lipoproteins radiolabeled with 125I or [3H]CEt in 150 μL of saline. At the indicated time, blood samples were collected in microvette tubes coated with heparin from exposed saphenous veins and centrifuged at 10,000g for 5 minutes at 4°C. [3H]CEt was directly radioassayed from plasma, as it is not hydrolyzable in cells and therefore free [3H]-cholesterol cannot return in plasma. Differently, 125I was measured in the trichloroacetic acid (TCA)-precipitable fraction of plasma to eliminate the contribution of protein degradation. Disappearance curves of LDL were generated by dividing the plasma radioactivity at each point by the radioactivity determined 2 minutes after tracer injection, whereas that of oxLDL were based on the theoretical values.
being that if the CE-SU activity we measured in absence of SR-BI is attributable to CD36, then no SU should be detectable. SR-BI/CD36 double (d) KO mice were therefore generated. Plasma cholesterol, triglyceride, and free fatty acid concentrations of wild-type, CD36 KO, SR-BI KO, and SR-BI/CD36 dKO mice are shown in the Table and their lipoprotein cholesterol patterns in Figure 2. Our cholesterol results for SR-BI KO mice are very similar to those of Rigotti et al, as a higher plasma cholesterol level and a greater HDL-cholesterol peak were observed. However differently from Feibraio et al, we have not detected a 30% increase in HDL cholesterol in CD36 KO mice, but rather a 27% decrease. Others have seen no difference between the 2 genotypes. Thus we conclude that this modest discrepancy may relate to the starting period or the method of blood collection. Indeed, differently from Feibbraio et al our mice were starved for a longer period. The dKO mice show slightly higher levels of plasma cholesterol and a bigger HDL-cholesterol peak than those of SR-BI KO mice. As seen in the Table, no significant differences were detected between the triglyceride levels of the 4 genotypes. However, as shown by Feibbraio et al, our CD36 KO mice have higher levels of free fatty acids than wild-type mice. This 70% increase is also found in SR-BI/CD36 dKO mice. LDL-protein and CE clearance studies were conducted in wild-type and SR-BI/CD36 dKO mice. Figure 3 reveals that CE-SU is abolished in the dKO mice, indicating that in absence of SR-BI activity, CD36 is able to selectively take up CE from LDL.

Role of CD36 in oxLDL Metabolism In Vivo in the Mouse

Experiments were conducted to define the role of CD36 in oxLDL metabolism in vivo. Clearance was followed for 1 hour, as oxLDL disappear rapidly. The comparison of protein to CE disappearance curves reveals that in wild-type mice mildly oxLDL are subjected to CE-SU (Figure 4A) but not oxLDL (Figure 4B). Thus in the mouse, the extent of LDL oxidation is negatively correlated with the magnitude of CE-SU. Importantly, CD36 deficiency does not accelerate decay as for LDL but delays protein and CE disappearances of both mildly oxLDL (Figure 4A) and oxLDL (Figure 4B). As both protein and CE clearances are delayed, we conclude that CD36 plays a significant role in the holoparticle uptake of both types of oxLDL. These data are supported by the half-life data shown in supplemental Figure V. The effect of CD36 being incomplete, we looked for an implication of SR-BI by conducting the same experiment but with wild-type and SR-BI KO mice. The results presented in supplemental Figure V and VI are clearly against a role of SR-BI in both types of oxLDL holoparticle or selective uptake.

Role of Mouse Hepatic Cell CD36 in Native LDL and oxLDL Metabolism In Vitro

To assess the involvement of hepatic CD36, experiments were conducted with primary cultures of hepatic cells from wild-type and CD36 KO mice. Supplemental Figure IV shows that these liver cells, which contain 90% hepatocytes, express CD36. Figure 5 reveals that although CD36 deficiency reduces LDL association, it increases LDL degradation by 110%. Thus, CD36 also impairs LDL uptake/degradation in primary cultures of mouse hepatic cells. Interestingly, mildly oxLDL degradation was raised by 55%, whereas oxLDL degradation was reduced by 60% in accordance with clearance studies. Differently, CE association and CE-SU were not significantly affected by the CD36 deficiency, except for oxLDL, where for an unknown reason a reduction by half of the CE association and CE-SU was detectable. It is however likely that this apparent discrepancy is simply linked to the fact that oxLDL disappear so quickly from mouse blood circulation that CE-SU does not occur at a significant level in vivo or is barely detectable. If true this would mean that in vitro, hepatic CD36 can selectively take up CE from oxLDL.

Discussion

Although the role of SR-BI in HDL-CE SU is well accepted, in vitro assays using CHO and Y1 2/3 adrenal cells have rather suggested SR-BI–independent pathways for LDL-CE SU. The first study showed the importance of lipoprotein lipase, and the last of LDLr-related protein (LRP) and apoE. Later, we showed4 by in vivo clearance studies in wild-type and SR-BI KO mice that SR-BI is responsible for LDL-CE SU but that a SR-BI–independent pathway can rescue SR-BI deficiency. The clearance studies in SR-BI/CD36 dKO mice

| Table. Plasma Lipid Levels of Wild-Type, CD36 KO, SR-BI KO, and SR-BI/CD36 dKO Mice |
|---------------------------------|-----------------|-----------------|
| Cholesterol (mg/dl) | Triglycerides (mg/dl) | Free Fatty Acids (mM) |
| Wild-type | 120.6±2.9 (10) | 34.9±4.4 (8) | 0.67±0.058 (6) |
| CD36 KO | 88.2±4.2* (11) | 32.4±3.0 (8) | 1.21±0.039* (4) |
| SR-BI KO | 208.3±10.7† (4) | 35.3±2.2 (4) | 0.62±0.065† (4) |
| CD36/SR-BI dKO | 236.5±2.4* (4) | 35.8±1.1 (4) | 1.37±0.22* (2) |

Values are mean±SEM from the No. of mice indicated in parenthesis. *Statistically significant difference (P<0.005) from wild-type mice. †Statistically significant difference (P<0.0001) from CD36 KO mice.
Figure 3. Clearance of \(^{125}\)I- and \(^3\)H-CET-labeled LDL in wild-type or SR-BI/CD36 double KO mice. 480 \(\mu g\) of nonradiolabeled human LDL and 20 \(\mu g\) of LDL labeled with \(^{125}\)I or with \(^3\)H-CET were injected in wild-type (A) or SR-BI/CD36 double KO (B) mice. Each point represents the mean±SEM from 6 mice.

Figure 4. Clearance of \(^{125}\)I- and \(^3\)H-CET-labeled mildly oxLDL or oxLDL in wild-type or CD36 KO mice. 450 \(\mu g\) of nonradiolabeled lipoprotein and 50 \(\mu g\) of the same lipoprotein labeled with \(^{125}\)I or with \(^3\)H-CET-injected in wild-type (+/+) or CD36 KO (-/-) mice. A, Mildly oxLDL. B, oxLDL. Each point represents the mean±SEM from 3 mice.

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(Figure 3) presented in this study directly pointed out CD36 as being responsible for the SR-BI-independent pathway in vivo. As the cholesterol lipoprotein patterns do not differ substantially between SR-BI KO and SR-BI/CD36 dKO mice (Figure 2 and Table), it is unlikely that the complete loss of CE-SU encountered in the dKO mice is a consequence of the endogenous pool of lipoproteins of these mice.

As CD36 plays a role in LDL-CE SU in absence of SR-BI and no SR-BI deficiency has yet been described in humans, the physiological importance of CD36 in LDL clearance is uncertain. On the other hand, if loss-of-function mutations are to be discovered in the SR-BI gene, it will become important to address in more details the effect of CD36-mediated CE-SU on LDL metabolism and to define whether the pathway is anti- or proatherogenic. Indeed, although HDL-CE SU has been for many years considered as beneficial, more investigations are needed before such a conclusion can be drawn for LDL-CE SU either by SR-BI or CD36, as it is possible that small dense LDL are generated by CE depletion and such a subclass of LDL is positively associated with coronary heart diseases (CHD).

Importantly, we found in vivo (Figure 1) and in vitro (Figure 5) that murine CD36 impairs uptake/degradation of LDL-protein. The in vitro study revealed that mildly oxLDL are also more degraded in CD36 deficient cells, but not oxLDL. It is likely that in absence of CD36, the association of mildly oxLDL to the LDLr is favored because these may also interact with LDLr. As the LDLr level is not different between wild-type and CD36 KO mice (supplemental Figure IV) and as both receptors have similar affinities for human LDL, we suggest that CD36, by tethering native LDL particles to the cell surface, reduces the association of LDL to the LDLr and retards their clearance.

We also showed in vivo that the extent of LDL oxidation is negatively correlated with CE-SU (Figure 4). It can be speculated that when oxidized, LDL particles lose their ability to deliver as much CE by the selective uptake pathway as native LDL. Alternatively, the receptor(s) involved may have a lower potential toward them than for native LDL. It is also possible that oxLDL are too readily taken up by hepatic cells for ultimate degradation to allow a CE-SU process. Moreover, we herein demonstrate by in vivo clearance studies that neither CD36 nor SR-BI is a mediator of oxLDL-CE SU (Figure 4 and supplemental Figure VI). Because we have shown that CE-SU can occur from mildly oxLDL, this means that the receptor or the mechanism involved requires identification.

Interestingly, CD36 deficiency, although it does not accelerate decay as for LDL, delays both protein and CE disappearances of mildly oxLDL and oxLDL (Figure 4), indicating that CD36 significantly, although not completely, mediates oxLDL holoparticle uptake. The partial implication of CD36 in oxLDL global uptake was already shown in vitro. Nonetheless, we provide the first evidence of the importance of CD36 in vivo. It is unlikely that SR-AI/II is the other receptor partially involved because Ling et al have compared the plasma clearance of oxLDL in SR-AI/II KO and wild-type mice and found no difference. The same conclusion applies to SR-BI as we have seen no impact in oxLDL clearance in SR-BI KO mice (supplemental Figure VI). Thus, more...
investigations are also required to identify the other mouse oxLDL receptor/mediator leading as CD36 to holoparticle uptake.

The matter of the tissue(s) from which CD36 exerts its effect on native and oxLDL metabolism in vivo deserves some attention, as it is frequently assumed that CD36 is barely present in liver and our results suggest that the effect that we observed is attributable to hepatic CD36. Firstly, it is well accepted that when oxLDL are injected in rodents, they are readily cleared by the liver which is also the major tissue involved in native LDL uptake. Secondly, in the present study we show a detectable level of CD36 in mouse hepatic cells and we were able to detect, with primary cultures of these cells, a very similar impact of CD36. However, this does not completely exclude that part of the effect we observed in vivo is attributable to another tissue.

It is important to discuss our findings in relation to the association of CD36 with CHD. Indeed, as recently reviewed by Febbraio and Silverstein, CD36 can be viewed as “The Good and The Bad”, depending on its very many roles. Very elegant studies were conducted in hyperlipidemic mouse models and all23–25 but one of them26 revealed a proatherosclerotic role of CD36. Based on our results, CD36 by retarding LDL disappearance in the circulation can be regarded as proatherosclerotic. In opposition, it can be viewed as antiatherosclerotic by being involved in oxLDL clearance by the liver. As oxLDL are usually found at a much lower concentration than LDL in blood circulation, our data rather suggest a proatherosclerotic role of murine CD36. How can our results be extrapolated to humans? Human CD36 deficiency exists and has been divided into 2 subgroups according to the phenotypes. In type I deficiency, neither platelets nor monocytes express CD36. CD36 is expressed in monocytes but not in platelets of type II deficient individuals. This type is very rare in Whites (0.3% of the population) but more frequent in Asians and African Americans (3 to 4%).27 Interestingly, evidence is growing that CD36 might be beneficial in humans. Indeed, abnormalities of glucose and lipid metabolism were shown in CD36 deficiency including increased plasma triglycerides, decreased HDL-cholesterol, increased LDL-cholesterol, impaired glucose tolerance, and delayed response of insulin secretion.28–29 Moreover, the frequency of CD36 deficiency is higher in patients with CHD than in control subjects.30 The apparent difference between conclusions reached in humans and rodents may be a consequence of their lipoprotein profiles, as the LDL/HDL ratio is higher in humans than in rodents. Furthermore, it is possible that the effect of a receptor such as CD36 on atherosclerosis depends on the plasma lipoprotein concentrations and compositions (normo- versus hyperlipidemic).

In conclusion, our study has demonstrated that CD36: (1) can lead to LDL-CE SU in absence of SR-BI; (2) retards LDL clearance; and (3) is responsible for a significant fraction of oxLDL particle clearance.

Acknowledgments
The authors wish to thank Dr Maria Febbraio for generously providing the CD36 null mice and Dr Lise Bernier for helpful assistance in lipoprotein fractionation.

Sources of Funding
This work was supported by the Canadian Institute for Health Research and Quebec Heart Foundation grants attributed to L.B. D.R. was the recipient of a scholarship from the Fonds de la Recherche en Santé du Québec.

Disclosures
None.

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Arterioscler Thromb Vasc Biol. 2008;28:1290-1295; originally published online April 24, 2008; doi: 10.1161/ATVBAHA.107.161653
Arteriosclerosis, Thrombosis, and Vascular Biology is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
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Print ISSN: 1079-5642. Online ISSN: 1524-4636

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Mouse CD36 has opposite effects on LDL and oxidized LDL metabolism in vivo

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Online publication information

Supplementary Material and Methods

Material

Anti-SR-BI and anti-C36 (NB-144) antibodies were obtained from Novus Biologicals (Littleton, CO). Anti-LDLr antibodies were from Research Diagnostics (Flanders, NJ) and goat anti-rabbit IgG coupled to horseradish peroxidase were from Chemicon (Temecula, CA). Lipid hydroperoxide assay kit came from EMD Bioscience (San Diego, CA), while the oxLDL ELISA assay kit was purchased from Medicorp Inc. (Montreal, Que). Enhanced chemiluminescence substrate was from Roche Diagnostics (Laval, Que.).

Methods

Mice colonies and genotyping

As female SR-BI KO mice are infertile (1), the colony was maintained by breeding male homozygous (−/−) KO mice with heterozygous (+/−) females. To obtain SR-BI/CD36 dKO mice, male homozygous (−/−) SR-BI were crossbred with female homozygous (−/−) CD36. Littermates from that generation (SR-BI+/− and CD36+/−) were then intercrossed to obtain
SR-BI/CD36 dKO mice. SR-BI genotypes were determined by the PCR method of Rigotti et al. (2). CD36 genotyping was done by PCR analysis using specific primers for the targeted allele (5’- CAGCTCATACATTGCTGTTTATG CATG- and 3’- CGCTTCCTCGTCTTACGGTATC). PCR cycle conditions were 94°C for 1 minute, 65°C for 1 minute and 70°C for 2 minutes for 30 cycles with a final extension of 2 minutes at 70°C. The PCR product (~750-800 bp) was size fractionated on 1.2% agarose gels and visualized with ethidium bromide.

**Oxidative status of LDL preparations**

LDL oxidation was measured by a lipid hydroperoxide assay kit and by an OxLDL ELISA kit as described by the manufacturers. Oxidation was also checked by measuring the electrophoretic mobility of LDL on a 0.5% agarose/barbital gel.

**HepG2 cell culturing and binding studies**

HepG2 cells were grown and LDL binding studies were conducted as previously described in (3).

**LDLr, SR-BI and CD36 protein estimation by immunoblotting**

Total cell proteins of hepatic cells from mice were extracted by Triton X-100 1% solubilization (4). Proteins were separated by 8% reducing SDS/PAGE and transferred on nitrocellulose paper according to the method of Burnette (5). The blots were incubated with anti-SR-BI (1:4000) or with anti-LDLr (1:500), or with anti-CD36 (1:1000) antibodies, and
thereafter with anti-rabbit IgG coupled to horseradish peroxidase (1:4000) followed by enhanced chemiluminescence detection on Kodak Biomax ML film. Protein expression was measured by densitometric scanning and analyzed with ImageQuant 5.2 Software (Molecular Dynamics, Sunnyvale, CA).

Other methods

Plasma fractional catabolic rates (FCR) were calculated using a two-compartment model according to the model of Matthews (6). Half-life data were generated as described in (7). Protein determination was done by the method of Lowry et al. (8) with BSA as standard. Student's \( t \)-test was used to obtain statistical comparison of the data. Differences were considered significant at \( p < 0.05 \).

Supplementary Results

Characterization of unlabeled and labeled LDL and oxLDL

The agarose/barbital gel electrophoresis of LDL, mildly oxLDL and oxLDL unlabeled or labeled with \( ^{125}\text{I} \) or \( ^{3}\text{H}-\text{CEt} \) shown in Figure I reveals that the different types of labeling do not modify significantly the migration of the labeled particles compared to their unlabeled counterpart. The competition binding studies appearing in Figure II show that the two different types of labeling do not modify the binding properties of either LDL, mildly oxLDL or oxLDL. Moreover, we found no significant differences in the lipid hydroperoxide levels
between unlabeled LDL, ^{125}\text{I}-\text{LDL} and ^3\text{H}-\text{CE} labeled LDL (mean value: 4.29 ± 0.20 (s.e.m.) nmoles/40 µg LDL-protein). LDL particles either labeled or not were also subjected to an oxLDL ELISA. Table I shows that there are 33-fold and 341-fold more oxLDL epitopes in mildly oxLDL and oxLDL than in LDL, respectively. Moreover, none of the labeling procedures affects the LDL scores. Thus, our preparations of native LDL cannot be considered as significantly oxidized. Furthermore our labeled LDL preparations do not differ from their unlabeled counterparts. This validates our clearance studies.

**Supplementary Statistical Analyses of Figures 1, 3, 4**

Clearance curves of Figures 1, 3, 4 were compared by regression analysis for significant differences, using the average and standard deviation for each point with a two-way ANOVA. The results are:

Figure 1: The clearance of ^{125}\text{I}-\text{LDL} is significantly slower (p < 0.0001) than the clearance of ^3\text{H}-\text{CEt-LDL} in wild-type mice. The clearance of ^{125}\text{I}-\text{LDL} is significantly slower (p < 0.0001) than the clearance of ^3\text{H}-\text{CEt-LDL} in CD36 KO mice. Thus LDL-CE SU is detectable in both genotypes. The clearance of ^{125}\text{I}-\text{LDL} in wild-type mice is significantly slower (p < 0.0001) than the clearance of ^{125}\text{I}-\text{LDL} in CD36 KO mice. The clearance of ^3\text{H}-\text{CEt-LDL} in wild-type mice is significantly slower (p < 0.03) than the clearance of ^3\text{H}-\text{CEt-LDL} in CD36 KO mice. Thus, CD36 impedes LDL clearance.
Figure 3: The clearance of $^{125}$I-LDL in wild-type mice is significantly slower ($p < 0.005$) than the clearance of $^{125}$I-LDL in SR-BI/CD36 KO mice. The clearance of $^3$H-CEt-LDL in wild-type mice is significantly slower ($p < 0.01$) than the clearance of $^3$H-CEt-LDL in SR-BI/CD36 KO mice. The clearance of $^{125}$I-LDL is significantly slower ($p < 0.002$) than the clearance of $^3$H-CEt-LDL in wild-type mice. The clearance of $^{125}$I-LDL is not significantly different ($p = 0.43$) than the clearance of $^3$H-CEt-LDL in SR-BI/CD36 KO mice. Thus LDL-CE SU is not detectable in SR-BI/CD36 KO mice.

Figure 4: The clearance of $^{125}$I-mildly oxLDL is significantly slower ($p < 0.0001$) than the clearance of $^3$H-CEt-mildly oxLDL in wild-type mice, as well as in CD36 KO mice. Thus mildly oxLDL-CE SU is detectable in both genotypes. Differently, no statistical evidence for oxLDL-CE SU was found. The clearance of $^{125}$I-mildly oxLDL in wild-type mice is significantly slower ($p < 0.0001$) than the clearance of $^{125}$I-mildly oxLDL in CD36 KO mice. The clearance of $^3$H-CEt-mildly oxLDL in wild-type mice is significantly slower ($p < 0.0001$) than the clearance of $^3$H-CEt-mildly oxLDL in CD36 KO mice. Identical findings were obtained for oxLDL clearances. Thus, CD36 participates to mildly oxLDL and oxLDL clearance.

Supplementary references


TABLES

Table I: OxLDL levels in unlabeled and labeled LDL preparations in comparison with the levels in mildly oxLDL and oxLDL.

<table>
<thead>
<tr>
<th></th>
<th>Unlabeled (μg oxLDL protein/mg LDL protein)</th>
<th>Labeled with $^{125}$I (μg oxLDL protein/mg LDL protein)</th>
<th>Labeled with $^{3}$H-CE (μg oxLDL protein/mg LDL protein)</th>
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<tr>
<td>LDL batch 1</td>
<td>0.82 ± 0.060 (2)</td>
<td>0.65 ± 0.065 (2)</td>
<td>0.63 ± 0.075 (2)</td>
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<td>LDL batch 2</td>
<td>0.71 ± 0.012 (3)</td>
<td>0.63 ± 0.065 (2)</td>
<td>0.57 ± 0.045 (2)</td>
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<tr>
<td>Mildly oxLDL</td>
<td>22.4 ± 2.46* (3)</td>
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<td>ND</td>
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<td>OxLDL</td>
<td>228.7 ± 67.8*† (2)</td>
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Two different human serum lots were used to prepare two different batches of LDL (1 & 2). Each batch was labeled with $^{125}$I or $^{3}$H-CE. All LDL preparations as well as mildly oxLDL and oxLDL were assayed by an oxLDL ELISA in duplicate. Values are given as mean ± s.e.m of the number of assays indicated in parenthesis. *Statistically different (p < 0.05) from LDL, †Statistically different (p < 0.05) from mildly oxLDL. N.D.: Not determined.
Table II. Fractional catabolic rates of $^{125}$I- and $^3$H-CET-labeled LDL from plasma of wild-type (+/+) and CD36 KO (-/-) mice.

<table>
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<th>CD36 genotypes</th>
<th>$^{125}$I-LDL</th>
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<td>$0.143 \pm 0.016$</td>
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<td>$0.183 \pm 0.012^*$</td>
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<td>$0.290 \pm 0.017^*$</td>
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<td>$^3$H-CET-LDL</td>
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Wild-type and CD36 KO mice were injected via the tail vein with a bolus of human LDL (480 μg non-radiolabeled LDL and 20 μg lipoprotein LDL radiolabeled with either $^{125}$I or $^3$H-CET). Blood samples were taken at selected intervals and counted for radioactivity as described in Methods. The FCR were calculated using a two-compartment model and are expressed as pools/h. All values are given as mean ± s.e.m of 6 mice. *Statistically different (p < 0.05) from wild-type (+/+ ) mice.
Figure legends

**Figure I: Agarose/barbital gel electrophoresis of LDL, mildly oxLDL and oxLDL unlabeled or labeled with $^{125}$I or $^3$H-CEt.** A) Migration of unlabeled LDL, mildly oxLDL and oxLDL, respectively lanes 1-3, and LDL, mildly oxLDL and oxLDL labeled with iodine-125, respectively lanes 4-6. B) Migration of unlabeled LDL, mildly oxLDL and oxLDL, respectively lanes 1-3, and LDL, mildly oxLDL and oxLDL labeled with $^3$H-CEt, respectively lanes 4-6.

**Figure II: Competition of the binding of $^{125}$I-labeled lipoproteins to HepG2 cells.** HepG2 cells were incubated at a final concentration of 20 μg/ml of either a mixture of $^{125}$I-LDL and unlabeled LDL or $^{125}$I-LDL and $^3$H-CEt-LDL in the following dilutions: (1/1, 7/8, 3/4, 1/2, 1/4, 1/10, 1/20) (A). The same conditions were followed for mildly oxLDL (B) and oxLDL (C). After incubation and washing, the cell bound gamma radioactivity was measured.

**Figure III: Effect of co-injections of mildly oxLDL or oxLDL on LDL-CEt disappearance in wild-type or SR-BI KO mice.** Mildly oxLDL or oxLDL (480 μg proteins) were injected with $^3$H-CEt-LDL (20 μg) in wild-type (A) or SR-BI KO (B) mice. Lipoprotein disappearance was followed in the blood. Each point represents the mean ± s.e.m. derived from 3 mice. In wild-type mice, only oxLDL reduced significantly (p < 0.001) the disappearance of $^3$H-CEt-LDL. In SR-BI KO mice, both mildly oxLDL and oxLDL reduced significantly (p < 0.0001) the disappearance of $^3$H-CEt-LDL.
Figure IV: Immunoblot analysis of SR-BI, CD36 and rLDL expression in hepatocytes of wild-type or CD36 KO mice. Hepatocytes of wild-type (lanes 1 to 2) and CD36 KO (lanes 3 to 4) mice were isolated and their proteins extracted as described in Methods. Proteins (100 μg) were separated by SDS-PAGE and immunoblotted with anti-rabbit SR-BI polyclonal antibody, or anti-rabbit rLDL, or anti-CD36 antibodies followed by enhanced chemiluminescence detection. Two experiments gave essentially identical results.

Figure V: Half-life data of mildly oxLDL and oxLDL in the plasma of wild-type, CD36 KO or SR-BI KO mice. 450 μg of non-radiolabeled mildly oxLDL and 50 μg of mildly oxLDL labeled with 125I or with 3H-CEt, or 450 μg of non-radiolabeled oxLDL and 50 μg of oxLDL labeled with 125I or with 3H-CEt were injected in wild-type (+/+ or CD36 KO (-/-) mice (top panel) or SR-BI KO (-/-) mice (bottom panel). Lipoprotein disappearance was followed in the blood and half-life data were calculated. aSignificantly different from the wild-type result. bSignificantly different from the result obtained with the iodinated labeled mildly oxLDL or oxLDL.

Figure VI: Turnover studies of 125I and 3H-CEt labeled mildly or oxidized LDL in wild-type or SR-BI KO mice. Mildly oxLDL (A,B) or oxLDL (C,D) labeled with 125I (A,C) or with 3H-CEt (B,D) were injected in wild-type (+/+ or SR-BI KO (-/-) mice and the disappearance of the labeled lipoproteins was followed in the blood. Each point represents the mean ± s.e.m. derived from 3 mice. No statistical difference was found between the clearance curves of the genotypes for either mildly oxLDL or oxLDL. Mildly oxLDL-CE SU was demonstrated in both genotypes (p <0.0001), but not oxLDL-CE SU.
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