

## Lipoprotein(a) An Elusive Cardiovascular Risk Factor

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**Abstract**—Lipoprotein (a) [Lp(a)], is present only in humans, Old World nonhuman primates, and the European hedgehog. Lp(a) has many properties in common with low-density lipoprotein (LDL) but contains a unique protein, apo(a), which is structurally different from other apolipoproteins. The size of the apo(a) gene is highly variable, resulting in the protein molecular weight ranging from 300 to 800 kDa; this large variation may be caused by neutral evolution in the absence of any selection advantage. Apo(a) influences to a major extent metabolic and physicochemical properties of Lp(a), and the size polymorphism of the apo(a) gene contributes to the pronounced heterogeneity of Lp(a). There is an inverse relationship between apo(a) size and Lp(a) levels; however, this pattern is complex. For a given apo(a) size, there is a considerable variation in Lp(a) levels across individuals, underscoring the importance to assess allele-specific Lp(a) levels. Further, Lp(a) levels differ between populations, and blacks have generally higher levels than Asians and whites, adjusting for apo(a) sizes. In addition to the apo(a) size polymorphism, an upstream pentanucleotide repeat (TTTTA<sub>n</sub>) affects Lp(a) levels. Several meta-analyses have provided support for an association between Lp(a) and coronary artery disease, and the levels of Lp(a) carried in particles with smaller size apo(a) isoforms are associated with cardiovascular disease or with preclinical vascular changes. Further, there is an interaction between Lp(a) and other risk factors for cardiovascular disease. The physiological role of Lp(a) is unknown, although a majority of studies implicate Lp(a) as a risk factor. (*Arterioscler Thromb Vasc Biol.* 2004;24:2219-2226.)

**Key Words:** atherosclerosis ■ genetics ■ blacks ■ lipids

Lipoprotein(a) [Lp(a)] was first described ≈40 years ago, and interest in this entity is largely derived from its putative role as a cardiovascular risk factor.<sup>1-3</sup> Underlying this concept is the realization that Lp(a) has many properties in common with low-density lipoprotein (LDL), a well-established atherogenic factor for coronary artery disease.<sup>2-5</sup> Thus, the composition of the lipid moiety of Lp(a), including its cholesteryl ester-rich core, is similar to that of LDL, and the density distribution of the lipid moiety of Lp(a) in a given subject closely mirrors that of LDL.<sup>6</sup> Furthermore, like LDL, each particle of Lp(a) has 1 molecule of apolipoprotein B-100 (apo B-100); both apolipoprotein B (apoB) and the lipid core are pro-atherogenic.<sup>7</sup> Also, Lp(a) clearance rates are similar to those for LDL.<sup>8,9</sup> However, Lp(a) contains a unique protein, apolipoprotein(a) [apo(a)], which is structurally different from other apolipoproteins, having a hydrophilic, carbohydrate-rich structure with no amphipathic helices.<sup>5,10,11</sup> Apo(a) is linked to apoB through a single disulfide bond connecting their C-terminal regions<sup>12-15</sup> (Figure 1).

The presence of apo(a) influences to a major extent metabolic and physicochemical properties of Lp(a).<sup>16-18</sup> Notably, the cysteine residue in apoB involved in the covalent bond between apoB and apo(a) is close to the postulated LDL

receptor-binding region of apoB.<sup>19,20</sup> It appears from many clinical studies that Lp(a) levels are not affected by LDL receptor activity,<sup>16,21,22</sup> suggesting that the large, carbohydrate-rich apo(a) protein introduces a charge and/or steric interaction affecting the binding potential of apoB in Lp(a) for the LDL receptor. This could at least partly explain why Lp(a) plasma levels are mainly determined by the synthetic rate in contrast to LDL in which catabolism through the LDL receptor is an important regulator of plasma levels, although overall clearance rates are similar for the 2 lipoprotein fractions.<sup>3,8,9</sup> Lp(a) levels are particularly affected by apo(a) synthetic rate, which is subject to strong genetic regulation. Because of this strong genetic impact, Lp(a) plasma levels are affected only to a minor extent by age, sex, and environmental factors.<sup>3</sup>

### Lp(a): Molecular Properties

Lp(a) is very heterogeneous and the underlying reasons for this heterogeneity were uncovered by the elegant work on the gene structure of apo(a) by Lawn, Scanu, and their collaborators.<sup>23</sup> They reported an analogy between the apo(a) and plasminogen genes; both genes have coding sequences for loop structures stabilized by intrachain disulfide bonds, so-

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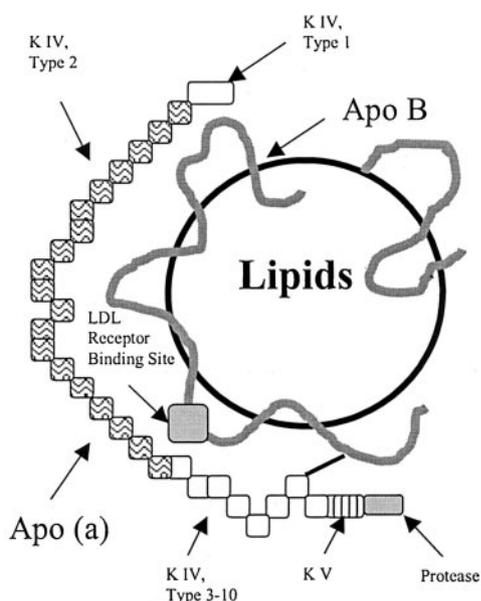
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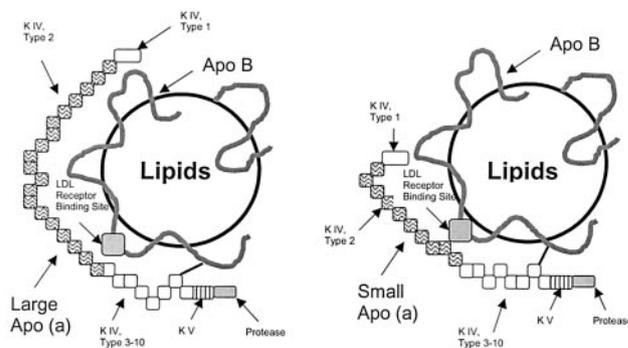
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**Figure 1.** Model of Lp(a). The LDL-like moiety consists of a lipid core of cholesteryl esters and triglycerides surrounded by a surface layer of phospholipid and free cholesterol. In addition to lipids, it also contains one molecule of apolipoprotein B, which is linked to apolipoprotein(a) through a single disulfide bond. The putative LDL receptor binding domain of apoB is shown. The apo(a) moiety consists of a single copy of kringles KIV, types 1 and 3 to 10, kringle V, and a protease domain analogous to plasminogen. In addition, it contains multiple copies of kringle IV, type 2.

called kringle (K) domains. The plasminogen gene contains coding sequences for 5 different K domains (K1 to K5), and 2 of these are present in the apo(a) gene, K4 and K5. Interestingly, the sequence coding for one of these K domains, K4, is repeated many-fold in the apo(a) gene.<sup>3,23–26</sup> Altogether, the apo(a) gene has 10 different types of plasminogen-like K4 domains, referred to as K4 type 1 through 10. K4 types 1 and 3 to 10 are present as single copies, whereas K4 type 2 is present as multiple copies, varying in number from 3 to >40 copies.<sup>16,23–26</sup> Each kringle contains ≈80 to 85 amino acids and has a molecular weight of ≈10 kDa, and the K4 repeat unit is thus unusually large. This heterogeneity in apo(a) gene size corresponds to a size variation in the apo(a) protein and apo(a) size isoforms containing from 12 to ≈50 K4 motifs have been reported, corresponding to a protein molecular weight ranging from 300 to 800 kDa.<sup>16,27</sup> The size variability of apo(a) impacts on Lp(a) levels; there is a general inverse relation between apo(a) size and Lp(a) levels.<sup>13,27,28</sup> Thus, smaller apo(a) sizes tend to correspond to higher plasma Lp(a) levels; however, this pattern is complex. For a given apo(a) size, there is a considerable variation in Lp(a) levels across individuals.

Lp(a) is found only in humans, in Old World nonhuman primates, and in the European hedgehog.<sup>3,29,30</sup> The hedgehog version of apo(a) appears to have evolved separately from the primate and human apo(a) versions, because it contains the plasminogen K3 domain instead of the K4 domain and is not subject to size heterogeneity.<sup>29</sup> Thus, although apo(a) is novel from an evolutionary standpoint, it appears nevertheless to have emerged twice.<sup>30</sup> Despite this, there is currently a



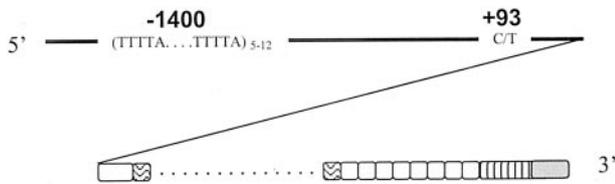
**Figure 2.** Two Lp(a) particles with different apo(a) sizes. Symbols as in Figure 1.

profound lack of understanding of any physiological function for Lp(a).

### Population Studies: Apo(a) Genetics and Predictors of Lp(a) Levels

Considerable efforts have been devoted to characterizing Lp(a) levels and the frequency distribution of apo(a) sizes for different populations.<sup>31–37</sup> Along with the aforementioned general inverse relationship between apo(a) size and Lp(a) plasma levels, differences between populations have been noted. The most profound differences have been noted between blacks and other populations, including Asians and whites.<sup>31–37</sup> Notably, as a group, blacks have substantially higher Lp(a) levels than do whites or Asians, and this difference is not explained by differences in apo(a) size distribution or, so far, by other genetic factors.<sup>38–40</sup> This apparent paradox was addressed by Marcovina et al, who demonstrated that blacks had higher mean Lp(a) levels than whites adjusting for apo(a) sizes.<sup>41</sup> In particular, the interethnic difference was considerable over mid-size apo(a) ranges. This finding has been confirmed in other studies, although the reasons for this difference remain unknown.<sup>39,40</sup> Apart from clarifying the role of apo(a) size variation across populations, these results demonstrate the importance of assessing apo(a) allele-specific Lp(a) levels. Because homozygosity for apo(a) size is rare, most subjects have Lp(a) particles with 2 different size apo(a), and the plasma Lp(a) level is the sum of Lp(a) carried in the 2 Lp(a) populations (Figure 2). Although smaller size apo(a) frequently is associated with higher Lp(a) levels, this relation is far from absolute, and for a given subject, the relative contribution of each apo(a) size isoform to the overall plasma Lp(a) level can vary substantially.<sup>37</sup> Therefore, assessment of apo(a) size isoform-specific Lp(a) levels provides additional information beyond plasma Lp(a) levels.

In most previous studies, Lp(a) levels have been expressed in mass units, usually in mg/dL. When comparing Lp(a) and LDL cholesterol in assessing cardiovascular risk, it should be noted that although LDL is expressed as cholesterol levels, Lp(a) level measurements reflect particle concentrations, including both lipid and protein components. The use of mass units for Lp(a) has also required an assumption of a particular apo(a) mass, ie, ignoring apo(a) size variation. The use of molar units (eg, nmol/L) is therefore preferable and this is



**Figure 3.** Upstream polymorphisms of the apo(a) gene. The PNR polymorphism is indicated by the variable presence (n=5 to 12) of TTTTA repeats at position -1400, and the C/T polymorphism represent a single nucleotide polymorphism in the promoter region of the apo(a) gene. The variability of KIV type 2 repeats of the apo(a) gene is indicated by the dotted line. Symbols for kringle repeats as in Figure 1.

facilitated by recent progress in the standardization of Lp(a) measurements.<sup>42</sup>

Because there is profound interindividual variation in Lp(a) levels for a given apo(a) size, suggesting that factors beyond gene size predict plasma levels, interest has focused on other apo(a) gene variations. Several polymorphisms in the apo(a) gene have been reported. Of these, a C/T variation in the promoter region of the apo(a) gene and a pentanucleotide repeat (TTTTA<sub>n</sub>) ≈ 1 kb upstream of the apo(a) gene have been studied in this regard<sup>43–49</sup> (Figure 3). Of note, very few blacks carry the T allele, suggesting that the presence of this allele in the black population reflects white ancestry. Whereas experimental data linked these variations to Lp(a) levels,<sup>43,46</sup> findings limited to a single locus need to be interpreted with caution in the presence of linkage disequilibrium. We and others have found evidence of linkage disequilibrium for both blacks and whites between the C/T, TTTTA<sub>n</sub> polymorphism, and apo(a) size, albeit to a different degree in the 2 ethnic groups.<sup>47–51</sup> When taking the apo(a) size polymorphism into account, the TTTTA<sub>n</sub> polymorphism, but not the C/T polymorphism, was found to affect Lp(a) levels.<sup>47,50,51</sup>

Of note, although many studies have addressed differences between blacks and whites, population differences with regard to Lp(a) and apo(a) size are not limited to these groups. Differences in both Lp(a) levels and apo(a) size distribution have been noted among Asian populations.<sup>52–56</sup> Thus, whereas East Asians have been reported to have a relatively narrow apo(a) size distribution and low Lp(a) levels, South Asians have higher mean Lp(a) levels.<sup>52–56</sup> Further studies of the genetic variability of apo(a) size and Lp(a) levels across different populations are clearly warranted and may help us understand both the evolutionary pattern and regulation of this intriguing molecule, as well as shed light on the contribution of Lp(a) to the risk for cardiovascular disease.

### Cardiovascular Risk Properties: Clinical Studies

Many clinical studies of coronary artery disease, cerebrovascular disease, and peripheral artery disease have been undertaken to assess the association of Lp(a) with disease. In numerous but not in all studies, mainly in white populations, elevations of plasma Lp(a) levels have been significantly correlated with coronary artery disease.<sup>57–70</sup> It has proven more difficult to detect an association between Lp(a) and cardiovascular disease among blacks;<sup>71,72</sup> however, using

assessment of apo(a) size allele-specific Lp(a) levels as discussed, we found an association between cardiovascular disease and Lp(a) among black and white men with small apo(a) sizes.<sup>38</sup> Further, the possibility that Lp(a) levels could be increased secondary to cardiovascular disease has been raised, implying that an increased Lp(a) level could be a result of disease rather than a part of the causal pathway. Studies in model systems demonstrating a stimulation of apo(a) secretion from monkey hepatocytes by interleukin (IL)-6 have provided some support for this concept.<sup>73</sup> However, the role of Lp(a) as an acute phase reactant has not been universally confirmed, and reductions in Lp(a) after estrogen treatment did not correlate with changes in acute phase proteins.<sup>74–76</sup> Recently, data supporting the possibility that Lp(a) might induce IL-6 formation in monocytes were published, suggesting other possible links between Lp(a) and an inflammatory response.<sup>77</sup> Prospective studies offer possibilities to assess more direct evidence for Lp(a) as a risk factor, and recently, 2 meta analyses have demonstrated an association of Lp(a) levels with cardiovascular disease in both retrospective and prospective studies.<sup>78,79</sup> However, because the majority of studies on Lp(a) have been performed in white populations, prospective studies in other populations are needed. Evidence for an interaction between Lp(a) and other established or emerging risk factors for cardiovascular disease, such as LDL cholesterol, high-density lipoprotein cholesterol, and homocysteine, have also been found.<sup>80–83</sup> In support of a pro-atherogenic role, Lp(a) has been detected in the vessel wall, where it appears to be retained more avidly than LDL.<sup>84–86</sup> Further studies are warranted in clarifying the interaction of Lp(a) with elements of the vascular wall. Furthermore, based on the similarity between apo(a) and plasminogen, it has been suggested that Lp(a) may be an interloper in the fibrinolytic system.<sup>87</sup> Recent clinical studies have provided support for this hypothesis because Lp(a) was reported to attenuate fibrinolysis and promote coagulation.<sup>88–90</sup> Collectively, these results support a pro-atherogenic/thrombotic role for Lp(a).

### Cardiovascular Risk Properties: Animal Models and In Vitro Studies

The presence of apo(a) only in humans and Old World primates have limited the use of animal models to explore the role of Lp(a) as a risk factor. To address this void, transgenic animal models have been created using mice and rabbits.<sup>91–96</sup> Initial attempts to generate a transgenic model for Lp(a) relied on the formation of this particle from human apo(a) and murine apoB. Although murine apoB did not form an Lp(a) complex with human apo(a), important insights into predictors of Lp(a) formation were gained from these studies.<sup>91</sup> In subsequent studies, animals transgenic for human apo(a) and apoB, allowing in vivo formation of a circulating Lp(a) particle, were used.<sup>92,93</sup> Somewhat contradictory results have been reported from these experiments; an increase in aortic atherosclerosis was reported by Rubin et al,<sup>93</sup> whereas no significant increase was reported in studies by Hobbs et al.<sup>92</sup> A number of factors could have contributed to the discordant results, such as different feeding conditions, varying genetic strains, and the actual apo(a) construct used. In more recent

studies, presence of atherosclerosis in transgenic mice fed a normal diet has been reported.<sup>94</sup> Studies in rabbits transgenic for Lp(a) have also provided support for a pro-atherogenic role.<sup>95,96</sup> In addition, studies in transgenic mice as well as from an arterial injury model in cynomolgus monkeys have provided support for a prothrombotic and/or antifibrinolytic role of Lp(a).<sup>83,97,98</sup>

### **Apo(a) and Cardiovascular Disease: Influence of Apo(a) Size**

Atherogenicity associated with Lp(a) could be caused by risk associated with the LDL moiety, with the apo(a) moiety, or from their unique combination in Lp(a). Because there is a substantial size heterogeneity of apo(a), and smaller apo(a) sizes are associated with higher plasma Lp(a) levels, the possibility that size variation of apo(a) could be associated with cardiovascular disease has been investigated in a number of studies.<sup>99–109</sup> This raises the question whether such an association simply would be caused by higher Lp(a) levels commonly carried by Lp(a) particles with smaller apo(a), or whether the size variation is independently important. In several of the early studies on apo(a) size and cardiovascular disease, the technique available permitted the resolution of a limited number of apo(a) size isoforms preventing comprehensive analysis of this issue.<sup>103,109</sup> This problem was largely resolved by the introduction of a high-resolution technique by Kamboh et al, later refined by Marcovina et al.<sup>110,111</sup> Using these techniques, which allow for detailed apo(a) isoform separation, small apo(a) size has been associated more consistently with cardiovascular disease in men. Studies have shown that total Lp(a) levels in individuals who carry at least 1 small isoform are associated with cardiovascular disease or with preclinical vascular changes.<sup>99,107,112,113</sup> These results suggest strongly that Lp(a) particles that carry small apo(a) molecules are the appropriate risk factor. We have reported that the small apo(a) molecule is not always the dominant form of Lp(a)—quite frequently, the larger apo(a) molecule is present in greater amount.<sup>37</sup> Using pulsed-field electrophoresis of DNA, we found in a number of individuals with a single apo(a) isoform that the smaller allele was not expressed. Thus, use of isoform-specific or allele-specific levels may be more informative in assessing Lp(a) as a risk factor.<sup>38,112</sup> This further implies that determination of predictors of allele-specific apo(a) levels would be of interest in relation to Lp(a) atherogenicity. Recently, we assessed the effect of apo(a) size together with 2 other genetic variants, the upstream C/T and pentanucleotide repeat polymorphisms, on allele-specific apo(a) levels in whites and blacks (Rubin et al, unpublished observations). In both populations, each allele size affected the level of that allele as well as that of the other allele, albeit to a smaller degree. Further, in whites but not in blacks, the pentanucleotide polymorphism affected levels. Overall, the results implicate an interaction between apo(a) size alleles as well as an influence by other genetic polymorphisms on allele-specific apo(a) levels. However, more studies are needed to confirm or refute these findings as well as the hypothesis that small apo(a) molecules convey cardiovascular risk.

Whereas an association with small isoform-specific Lp(a) levels and cardiovascular disease has been found in men, the results are less convincing in women.<sup>38,65,114</sup> Furthermore, although Lp(a) levels have been reported to be increased in women with myocardial infarction,<sup>115</sup> a recent prospective study on cerebrovascular disease demonstrated an association between Lp(a) and stroke in men but not in women.<sup>116</sup> This does not necessarily conflict with an association between plasma Lp(a) levels and cardiovascular disease among women, although it could suggest that any risk carried by specific apo(a) sizes may be subject to modulation by gender-specific factors. Further studies are needed to explore these apparent gender differences.

### **Oxidized Phospholipids: Relation to Lp(a)**

Recently, a new possible mechanism for apo(a) atherogenicity has been suggested. In a series of studies, Witztum et al have demonstrated convincingly that a key oxidized phospholipid is preferentially associated with Lp(a).<sup>117–119</sup> Proinflammatory, oxidized phospholipids are covalently bound to kringle V in apo(a), a portion of apo(a) associated with macrophage IL-8 production.<sup>117</sup> These results suggest that Lp(a) may act as a preferential acceptor that tightly binds oxidized phospholipids transferred from tissues or from other lipoproteins.<sup>119</sup> This could imply that Lp(a) functions as a scavenger absorbing potentially deleterious oxidized lipids, preventing an increased uptake in the vessel wall of other lipoproteins, primarily LDL, containing this factor. However, the presence of oxidized phospholipids in Lp(a), potentially being taken up by the vessel wall, could also accelerate development of atherosclerosis. Because kringle V is present as a single copy in the apo(a) molecule, the results suggest an apo(a) size-independent potential for binding of oxidized phospholipids, although it is possible that apo(a) size can affect the binding site through conformational changes. Notably, Lp(a) levels have been found to be higher among white centenarians, raising the possibility that Lp(a) may serve as a longevity factor, although opposite results have been reported in a Japanese cohort.<sup>120–122</sup>

### **Approaches to Modulate Lp(a) Levels: Challenges and Possibilities**

In assessing appropriate cardiovascular preventive measures, one needs to consider whether intervention to lower Lp(a) is clinically warranted. At present, Lp(a) is not an established cardiovascular risk factor and there are no guidelines recommending intervention.<sup>42,123</sup> Our current level of understanding would suggest that Lp(a) lowering might be beneficial in white men and some other subgroups of patients with high Lp(a) levels, but we still lack enough details on how to define such subgroups with regard to Lp(a) levels, apo(a) size, and presence of other risk factors. Additionally, the lack of knowledge of Lp(a) metabolism both regarding its formation and catabolism raises considerable challenges in devising strategies to lower Lp(a) levels.<sup>16</sup> Because apo(a) synthesis is of major importance in regulating Lp(a) levels, interference in Lp(a) particle formation may offer intervention possibilities.<sup>15,124,125</sup> Further, model studies in transgenic animals and cell cultures have suggested a hepatic elimination pathway.<sup>126</sup>

However, the current lack of a well-defined metabolic pathway for Lp(a) has prevented any progress regarding agents that might interfere with either formation or catabolism. At present, nicotinic acid is the only major hypolipidemic agent that has proven efficacy in lowering Lp(a) levels.<sup>127</sup>

### Role of Lp(a) Beyond Risk: Need-Driven Evolution or Development by Chance?

The limited distribution of Lp(a) to humans, Old World primates, and, as a parallel phenomenon, to the European hedgehog, has raised considerable interest and speculation regarding a possible physiological role. This has been fueled further by the pronounced genetic variability of the apo(a) gene, which represents an unusual example of a large nucleotide repeat in a coding region. It is well-known that short nucleotide repeats in coding regions can affect mRNA amounts or protein function and result in variable disease expression—examples of this can be seen in trinucleotide repeats for genes affecting Huntington chorea and fragile X.<sup>128,129</sup> However, there is no evidence of evolutionary pressure to favor development of Lp(a); the existence of apo(a) does not necessarily imply a useful function. After all, a considerable number of individuals have undetectable or very low Lp(a) levels without being at any apparent disadvantage. The interesting but so-far-unresolved differences between blacks and nonblacks regarding Lp(a) levels and apo(a) size allele variation might provide a possibility to assess any advantage associated with Lp(a); however, it could also be caused by the apo(a) allele distribution in the subset of the population that left Africa and subsequently gave rise to other population groups.<sup>130</sup> Functions suggested as possible for Lp(a) have been as an adjunct important for wound healing and/or regulation of angiogenesis.<sup>16,55</sup> Although a positive function may have been important during early stages of primate evolution in strengthening possibilities of survival, subsequently the importance of such a function may have decreased. A lack of selection against the gene may have resulted in retention of the apo(a) gene. In a state of neutral evolution, it would have been possible for a variety of sizes to evolve by duplication of the K4 type 2 repeat sequence, with the larger sizes and other as-yet-unidentified changes in the promoter region leading to widely varying levels of apo(a) expression with no evolutionary disadvantage.

Thus, although a majority of studies implicate Lp(a) as a risk factor, the suggestion from some studies of a beneficial effect may suggest either a dual function, perhaps depending on external circumstances, or simply a result of a neutral evolution. Although Lp(a) continues to offer surprises, further studies are needed to resolve the secrets of this mysterious phenomenon.

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### References

1. Berg K. A new serum system in man: the Lp system. *Acta Pathol Microbiol Scand.* 1963;59:362–382.

2. Albers JJ, Cabana VG, Warnick GR, Hazzard WR. Lp(a) lipoprotein: relationship to sinking pre- $\beta$  lipoprotein, hyperlipoproteinemia, and apolipoprotein B. *Metabolism.* 1975;24:1047–1054.
3. Utermann G. The mysteries of lipoprotein(a). *Science (Washington DC).* 1989;246:904–910.
4. Marcovina SM, Koschinsky ML. Lipoprotein (a): Structure, measurement, and clinical significance. In: Rifai N, Warnick GR, Dominiczak MH, eds. *Handbook of Lipoprotein Testing.* AACC Press: Washington, DC; 1997:283–313.
5. Gaubatz JW, Heideman C, Gotto AM Jr, Morrisett JD, Dahlén GH. Human plasma lipoprotein(a): structural properties. *J Biol Chem.* 1983; 258:4582–4589.
6. Rainwater DL, Ludwig MJ, Haffner SM, VandeBerg JL. Lipid and lipoprotein factors associated with variation in Lp(a) density. *Arterioscler Thromb Vasc Biol.* 1995;15:313–319.
7. Rader DJ, Rosas S. Management of selected lipid abnormalities. Hypertriglyceridemia, low HDL cholesterol, lipoprotein(a), in thyroid and renal diseases, and post-transplantation. *Med Clin North Am.* 2000;84: 43–61.
8. Demant T, Seeberg K, Bedynek A, Seidel D. The metabolism of lipoprotein(a) and other apolipoprotein B-containing lipoproteins: a kinetic study in humans. *Atherosclerosis.* 2001;157:325–339.
9. Rader DJ, Cain W, Zech LA, Usher D, Brewer HB Jr. Variation in lipoprotein(a) concentrations among individuals with the same apolipoprotein (a) isoform is determined by the rate of lipoprotein(a) production. *J Clin Invest.* 1993;91:443–447.
10. Utermann G, Weber W. Protein composition of Lp(a) lipoprotein from human plasma. *FEBS Lett.* 1983;154:357–361.
11. Scanu AM, Edelstein C. Learning about the structure and biology of human lipoprotein(a) through dissection by enzymes of the elastase family: facts and speculation. *J Lipid Res.* 1997;38:2193–2206.
12. Gabel B, Yao Z, McLeod RS, Young SG, Koschinsky ML. Carboxyl-terminal truncation of apolipoprotein B-100 inhibits lipoprotein (a) particle formation. *FEBS Lett.* 1994;350:77–81.
13. McCormick SPA, Linton MF, Hobbs HH, Taylor S, Curtiss LK, Young SG. Expression of human apolipoprotein B90 in transgenic mice: demonstration that apolipoprotein B90 lacks the structural requirements to form lipoprotein (a). *J Biol Chem.* 1994;269:24284–24289.
14. Brunner C, Kraft HG, Utermann G, Müller HJ. Cys4057 of apolipoprotein(a) is essential for lipoprotein(a) assembly. *Proc Natl Acad Sci U S A.* 1993;90:11643–11647.
15. Koschinsky ML, Cote GP, Gabel B, van der Hoek YY. Identification of the cysteine residue in apolipoprotein(a) that mediates extracellular coupling with apolipoprotein B-100. *J Biol Chem.* 1993;268: 19819–19825.
16. Hobbs HH, White AL. Lipoprotein(a): Intrigues and insights. *Curr Opin Lipidol.* 1999;10:225–236.
17. Berglund L. Diet and drug therapy for lipoprotein (a). *Curr Opin Lipidol.* 1995;6:48–56.
18. Edelstein C, Italia JA, Klezovitch O, Scanu AM. Functional and metabolic differences between elastase-generated fragments of human lipoprotein[a] and apolipoprotein[a]. *J Lipid Res.* 1996;37:1786–1801.
19. McCormick SPA, Ng JK, Taylor S, Flynn LM, Hammer RE, Young SG. Mutagenesis of the human apolipoprotein B gene in a yeast artificial chromosome reveals the site of attachment for apolipoprotein(a). *Proc Natl Acad Sci U S A.* 1995;92:10147–10151.
20. Callow MJ, Rubin EM. Site-specific mutagenesis demonstrates that cysteine 4326 of apolipoprotein B is required for covalent linkage with apolipoprotein(a) in vivo. *J Biol Chem.* 1995;270:23914–23917.
21. Kostner GM, Gavish D, Leopold B, Bolzano K, Weintraub MS, Breslow JL. HMG CoA reductase inhibitors lower LDL-cholesterol levels without reducing Lp(a) levels. *Circulation.* 1988;80:1313–1319.
22. Vessby B, Kostner G, Lithell H, Thomas J. Diverging effects of cholestyramine on apolipoprotein B and lipoprotein(a). A dose-response study of the effects of cholestyramine in hypercholesterolemia. *Atherosclerosis.* 1982;44:61–71.
23. McLean JW, Tomlinson JE, Kuang W-J, Eaton DL, Chen EY, Fless GM, Scanu AM, Lawn RM. cDNA sequence of human apolipoprotein (a) is homologous to plasminogen. *Nature.* 1987;330:132–137.
24. Lackner C, Cohen JC, Hobbs HH. Molecular definition of the extreme size polymorphism in apolipoprotein(a). *Hum Mol Genet.* 1993;2: 933–940.
25. van der Hoek YY, Wittekoek ME, Beisiegel U, Kastelein JJ, Koschinsky ML. The apolipoprotein kringle IV repeats which differ from the major

- repeat kringle are present in variably-sized isoforms. *Hum Mol Genet.* 1993;2:361–366.
26. Koschinsky ML, Beisiegel U, Henne-Bruns D, Eaton DL, Lawn RM. Apolipoprotein (a) size heterogeneity is related to variable number of repeat sequences in its mRNA. *Biochemistry.* 1990;29:640–644.
  27. Gavish D, Azrolan N, Breslow J. Plasma Lp(a) concentration is inversely correlated with the ratio of Kringle IV/Kringle V encoding domains in the apo(a) gene. *J Clin Invest.* 1989;84:2021–2027.
  28. Kraft HG, Kochl S, Menzel HJ, Sandholzer C, Utermann G. The apolipoprotein[a] gene—a transcribed hypervariable locus controlling plasma lipoprotein[a] concentration. *Hum Genet.* 1992;90:220–230.
  29. Lawn RM, Boonmark NW, Schwartz K, Lindahl GE, Wade DP, Byrne CD, Fong KJ, Meer K, Patthy L. The recurring evolution of lipoprotein(a). Insights from cloning of hedgehog apolipoprotein(a). *J Biol Chem.* 1995;270:24004–24009.
  30. Lawn RM. How often has Lp(a) evolved? *Clin Genet.* 1996;49:167–174.
  31. Guyton JR, Dahlén GH, Patsch W, Kautz JA, Gotto AM Jr. Relationship of plasma lipoprotein Lp(a) levels to race and to apolipoprotein B. *Arteriosclerosis.* 1985;5:265–272.
  32. Parra HJ, Luyeye I, Bouramou C, Demarquilly C, Fruchart JC. Black-white differences in serum Lp(a) lipoprotein levels. *Clin Chim Acta.* 1987;167:27–31.
  33. Gaubatz JW, Ghanem KI, Guevara J Jr, Nava ML, Patsch W, Morrisett JD. Polymorphic forms of human apolipoprotein(a): inheritance and relationship of their molecular weights to plasma levels of lipoprotein(a). *J Lipid Res.* 1990;31:603–613.
  34. Sandholzer C, Hallman DM, Saha N, Sigurdsson G, Lackner C, Czászár A, Boerwinkle E, Utermann G. Effects of the apolipoprotein(a) size polymorphism on the lipoprotein(a) concentration in 7 ethnic groups. *Hum Genet.* 1991;86:607–614.
  35. Gaw A, Boerwinkle E, Cohen JC, Hobbs HH. Comparative analysis of the apo(a) gene, apo(a) glycoprotein, and plasma concentrations of Lp(a) in three ethnic groups: evidence for no common “null” allele at the apo(a) locus. *J Clin Invest.* 1994;93:2526–2534.
  36. Marcovina SM, Albers JJ, Jacobs DR Jr, Perkins LL, Lewis CE, Howard BV, Savage P. Lipoprotein(a) concentrations and apolipoprotein(a) phenotypes in Caucasians and African Americans: the CARDIA study. *Arterioscler Thromb.* 1993;13:1037–1045.
  37. Rubin J, Paultre F, Tuck CH, Holleran S, Reed RG, Pearson TA, Thomas CM, Ramakrishnan R, Berglund L. Apolipoprotein(a) genotype influences isoform dominance pattern differently in African Americans and Caucasians. *J Lipid Res.* 2002;43:234–244.
  38. Paultre F, Pearson TA, Weil HFC, Tuck CH, Myerson M, Rubin J, Francis CK, Marx H, Philbin E, Reed RG, Berglund L. High levels of lipoprotein(a) carrying a small apolipoprotein(a) isoform is associated with coronary artery disease in both African American and Caucasian men. *Arterioscler Thromb Vasc Biol.* 2000;20:2619–2624.
  39. Mooser V, Scheer D, Marcovina SM, Wang J, Guerra R, Cohen J, Hobbs HH. The apo(a) gene is the major determinant of variation in plasma Lp(a) levels in African Americans. *Am J Hum Genet.* 1997;61:402–417.
  40. Barkley RA, Brown AC, Hanis CL, Kardia SL, Turner ST, Boerwinkle E. Lack of genetic linkage evidence for a trans-acting factor having a large effect on plasma lipoprotein(a) levels in African Americans. *J Lipid Res.* 2003;44:1301–1305.
  41. Marcovina SM, Albers JJ, Wijnsman E, Zhang Z, Chapman NH, Kennedy H. Differences in Lp(a) concentrations and apo(a) polymorphs between black and white Americans. *J Lipid Res.* 1996;37:2569–2585.
  42. Marcovina SM, Koschinsky ML, Albers JJ, Skarlatos S. Report of the National Heart, Lung, and Blood Institute Workshop on lipoprotein(a) and cardiovascular disease: recent advances and future directions. *Clin Chem.* 2003;49:1785–1796.
  43. Zysow BR, Lindahl GE, Wade DP, Knight BL, Lawn RM. C/T polymorphism in the 5′ untranslated region of the apolipoprotein(a) gene introduces an upstream ATG and reduces in vitro translation. *Arterioscler Thromb Vasc Biol.* 1995;15:58–64.
  44. Trommsdorff M, Köchl S, Lingenhel A, Kronenberg F, Delpont R, Vermaak H, Lemming L, Klausen C, Faergeman O, Utermann G, Kraft H-G. A pentanucleotide repeat polymorphism in the 5′ control region of the apolipoprotein(a) gene is associated with lipoprotein(a) plasma concentrations in Caucasians. *J Clin Invest.* 1995;96:150–157.
  45. Kraft HG, Windegg M, Menzel HJ, Utermann G. Significant impact of the +93 C/T polymorphism in the apolipoprotein(a) gene on Lp(a) concentrations in Africans but not in Caucasians: confounding effect of linkage disequilibrium. *Hum Mol Genet.* 1998;7:257–264.
  46. Wade DP, Clarke JG, Lindahl GE, Liu AC, Zysow BR, Meer K, Schwartz K, Lawn RM. 5′ control regions of the apolipoprotein(a) gene and members of the related plasminogen gene family. *Proc Natl Acad Sci U S A.* 1993;90:1369–1373.
  47. Mooser V, Mancini FP, Bopp S, Pethö-Schramm A, Guerra R, Boerwinkle E, Müller H-J, Hobbs HH. Sequence polymorphisms in the apo(a) gene associated with specific levels of Lp(a) in plasma. *Hum Mol Genet.* 1995;4:173–181.
  48. Valenti K, Aveyrier E, Leauté S, Laporte F, Hadjian AJ. Contribution of apolipoprotein(a) size, pentanucleotide TTTTA repeat and C/T (+93) polymorphisms of the apo(a) gene to regulation of lipoprotein(a) plasma levels in a population of young European Caucasians. *Atherosclerosis.* 1999;147:17–24.
  49. Rubin J, Pearson TA, Reed RG, Berglund L. A fluorescence-based, non-radioactive method for efficient detection of the pentanucleotide repeat (TTTTA)<sub>n</sub> polymorphism in the apolipoprotein(a) gene. *Clin Chem.* 2001;47:1758–1762.
  50. Kalina A, Csaszar A, Fust G, Nagy B, Szalai C, Karadi I, Duba J, Prohaszka Z, Horvath L, Dieplinger H. The association of serum lipoprotein(a) levels, apolipoprotein(a) size and (TTTTA)<sub>n</sub> polymorphism with coronary heart disease. *Clin Chim Acta.* 2001;309:45–51.
  51. Berglund L, Rubin J, Pearson TA, Holleran S, Ramakrishnan R. The apo(a) size, C/T and PNR polymorphisms in Lp(a): A population genetic study of linkage in African Americans and Caucasians. *Arterioscler Thromb Vasc Biol.* 2004;24:e80–e81.
  52. Geethanjali FS, Luthra K, Lingenhel A, Kanagasaba-Pathy AS, Jacob J, Srivastava LM, Vasisht S, Kraft HG, Utermann G. Analysis of the apo(a) size polymorphism in Asian Indian populations: association with Lp(a) concentration and coronary heart disease. *Atherosclerosis.* 2003;169:121–130.
  53. Hughes K, Aw TC, Kuperan P, Choo M. Central obesity, insulin resistance, syndrome X, lipoprotein(a) and cardiovascular risk in Indians, Malays, and Chinese in Singapore. *J Epidemiol Community Health.* 1997;51:394–399.
  54. Hoogeveen RC, Gambhir JK, Gambhir DS, Kimball KT, Ghazaly K, Gaubatz JW, Vaduganathan M, Rao RS, Koschinsky M, Morrisett JD. Evaluation of Lp(a) and other independent risk factors for CHD in Asian Indians and their USA counterparts. *J Lipid Res.* 2001;42:631–638.
  55. Utermann G. Genetic architecture and evolution of the lipoprotein(a) trait. *Curr Opin Lipidol.* 1999;10:133–141.
  56. Kraft HG, Lingenhel A, Pang RWC, Delpont R, Trommsdorff M, Vermaak H, Janus ED, Utermann G. Frequency distributions of apolipoprotein(a) kringle IV repeat alleles and their effects on lipoprotein(a) levels in Caucasian, Asian, and African populations: the distribution of null alleles is non-random. *Eur J Hum Genet.* 1996;4:74–87.
  57. Rhoads GG, Dahlén G, Berg K, Morton NE, Dannenberg AL. Lp(a) lipoprotein as a risk factor for myocardial infarction. *JAMA.* 1986;256:2540–2544.
  58. Dahlén GH, Guyton JR, Attar M, Farmer JA, Kautz JA, Gotto AM Jr. Association of levels of lipoprotein Lp(a), plasma lipids, and other lipoproteins with coronary artery disease documented by angiography. *Circulation.* 1986;74:758–765.
  59. Kostner GM, Avogaro P, Cazzolato G, Marth E, Bittolo-Bon G, Quinci GB. Lipoprotein Lp(a) and the risk for myocardial infarction. *Atherosclerosis.* 1981;38:51–61.
  60. Zenker G, Koltringer P, Bone G, Niederkorn K, Pfeiffer K, Jürgens G. Lipoprotein(a) as a strong indicator for cerebrovascular disease. *Stroke.* 1986;17:942–945.
  61. Cambillau M, Arnar ASJ, Giral P, Ather V, Segond P, Levenson J, Merli I, Megnien JL, Plainfosse MC, Moatti N, and the PCVMEIRA group. Serum Lp(a) as a discriminant marker of early atherosclerotic plaque at three extracoronary sites in hypercholesterolemic men. *Arterioscler Thromb.* 1992;12:1346–1352.
  62. Bostom AG, Gagnon DR, Cupples LA, Wilson PWF, Jenner JL, Ordovas JM, Schaefer EJ, Castelli WP. A prospective investigation of elevated lipoprotein (a) detected by electrophoresis and cardiovascular disease in women. The Framingham Heart Study. *Circulation.* 1994;90:1688–1695.
  63. Schaefer EJ, Lamon-Fava S, Jenner JL, McNamara JR, Ordovas JM, Davis CE, Abolafia JM, Lippel K, Levy RI. Lipoprotein (a) and risk of coronary heart disease in men: the lipid research clinics coronary primary prevention trial. *JAMA.* 1994;271:999–1003.
  64. Bostom AG, Cupples LA, Jenner JL, Ordovas JM, Seman LJ, Wilson PWF, Schaefer EJ, Castelli WP. Elevated plasma lipoprotein (a) and

- coronary heart disease in men aged 55 years and younger: a prospective study. *JAMA*. 1996;276:544–548.
65. Wild SH, Fortmann SP, Marcovina SM. A prospective case-control study of lipoprotein (a) levels and apo (a) size and risk of coronary heart disease in Stanford Five-City Project participants. *Arterioscler Thromb Vasc Biol*. 1997;17:239–245.
  66. Nguyen TT, Ellefson RD, Hodge DO, Bailey KR, Kottke TE, Abu-Lebdeh HS. Predictive value of electrophoretically detected lipoprotein (a) for coronary heart disease and cerebrovascular disease in a community-based cohort of 9936 men and women. *Circulation*. 1997;96:1390–1397.
  67. Berg K, Dahlén G, Christophersen B, Cook T, Kjekshus J, Pedersen T. Lp(a) lipoprotein level predicts survival and major coronary events in the Scandinavian Simvastatin Survival Study. *Clin Genet*. 1997;52:254–261.
  68. Jauhiainen M, Koskinen P, Ehnholm C, Frick MH, Mänttari M, Manninen V, Huttunen JK. Lipoprotein (a) and coronary heart disease risk: a nested case-control study of the Helsinki Heart Study participants. *Atherosclerosis*. 1991;89:59–67.
  69. Rosengren A, Wilhelmsen L, Eriksson E, Risberg B, Wedel H. Lipoprotein (a) and coronary heart disease: a prospective case-control study in a general population sample of middle aged men. *BMJ*. 1990;301:1248–1251.
  70. Cremer P, Nagel D, Labrot B, Mann H, Mueche R, Elster H, Seidel D. Lipoprotein (a) as predictor of myocardial infarction in comparison to fibrinogen, LDL cholesterol and other risk factors: results from the prospective Gottingen Risk Incidence and Prevalence Study (GRIPS). *Eur J Clin Invest*. 1994;24:444–453.
  71. Sorrentino MJ, Vielhauer C, Eisenbart JD, Fless GM, Scanu AM, Feldman T. Plasma lipoprotein(a) concentration and coronary artery disease in black patients compared with white patients. *Am J Med*. 1992;61:402–417.
  72. Moliterno DJ, Jokinen EV, Miserez AR, Lange RA, Willard JE, Boerwinkle E, Hillis LD, Hobbs HH. No association between plasma lipoprotein(a) concentrations and the presence or absence of coronary atherosclerosis in African Americans. *Arterioscler Thromb Vasc Biol*. 1995;15:850–855.
  73. Ramharack D, Barkalow D, Spahr M. Dominant negative effect of TGF-beta1 and TNF-alpha on basal and IL-6-induced lipoprotein(a) mRNA expression in primary monkey hepatocyte cultures. *Arterioscler Thromb Vasc Biol*. 1998;18:984–990.
  74. Noma A, Abe A, Maeda S, Seishima M, Makino K, Yano Y, Shimokawa K. Lp(a): an acute-phase reactant? *Chem Phys Lipids*. 1994;67/68:411–417.
  75. Tuck CH, Holleran S, Berglund L. Hormonal regulation of lipoprotein (a) levels: effects of estrogen replacement therapy on lipoprotein (a) and acute phase reactants in post-menopausal women. *Arterioscler Thromb Vasc Biol*. 1997;17:1822–1829.
  76. Andreassen A, Berg K, Torsvik H. Changes in Lp(a) lipoprotein and other plasma proteins during acute myocardial infarction. *Clin Genet*. 1994;46:410–416.
  77. Buechler C, Ullrich H, Aslanidis C, Bared SM, Lingenhel A, Ritter M, Schmitz G. Lipoprotein(a) downregulates lysosomal acid lipase and induces interleukin-6 in human blood monocytes. *Biochim Biophys Acta*. 2003;1642:25–31.
  78. Stein JH, Rosenson RS. Lipoprotein Lp(a) excess and coronary heart disease. *Arch Intern Med*. 1997;157:1170–1176.
  79. Danesh J, Collins R, Peto R. Lipoprotein(a) and coronary heart disease. Meta-analysis of prospective studies. *Circulation*. 2000;102:1082–1085.
  80. Maher VM, Brown BG, Marcovina SM, Hillger LA, Zhao XQ, Albers JJ. Effects of lowering elevated LDL cholesterol on the cardiovascular risk of lipoprotein(a). *JAMA*. 1995;274:1771–1774.
  81. Cantin B, Gagnon F, Moorjani S, Despres JP, Lamarche B, Lupien PJ, Dagenais GR. Is lipoprotein(a) an independent risk factor for ischemic heart disease in men? The Quebec Cardiovascular Study. *J Am Coll Cardiol*. 1998;31:519–525.
  82. Foody JM, Milberg JA, Robinson K, Pearce GL, Jacobsen DW, Sprecher DL. Homocysteine and lipoprotein(a) interact to increase CAD risk in young men and women. *Arterioscler Thromb Vasc Biol*. 2000;20:493–499.
  83. von Eckardstein A, Schulte H, Cullen P, Assman G. Lipoprotein(a) further increases the risk of coronary events in men with high global cardiovascular risk. *J Am Coll Cardiol*. 201;37:434–439.
  84. Dangas G, Mehran R, Harpel PC, Sharma SK, Marcovina SM, Dube G, Ambrose JA, Fallon JT. Lipoprotein(a) and inflammation in human coronary atheroma: association with the severity of the clinical presentation. *J Am Coll Cardiol*. 1998;32:2035–2042.
  85. Nielsen LB. Atherogenicity of lipoprotein(a) and oxidized low density lipoprotein: insight from in vivo studies of arterial wall influx, degradation and efflux. *Atherosclerosis*. 1999;143:229–243.
  86. Beisiegel U, Niendorf A, Wolf K, Reblin T, Rath M. Lipoprotein(a) in the arterial wall. *Eur Heart J*. 1990;11(Suppl E):174–183.
  87. Miles LA, Plow EF. Lp(a): an interloper into the fibrinolytic system? *Thromb Haemost*. 1990;63:331–335.
  88. Marcovina SM, Koschinsky ML. Evaluation of lipoprotein(a) as a prothrombotic factor: progress from bench to bedside. *Curr Opin Lipidol*. 2003;14:361–366.
  89. von Depka M, Nowak-Göttl U, Eisert R, Dieterich C, Barthels M, Scharrer I, Ganser A, Ehrenforth S. Increased lipoprotein(a) levels as an independent risk factor for venous thromboembolism. *Blood*. 2000;96:3364–3368.
  90. Nowak-Göttl U, Junker R, Kreuz W, von Eckardstein A, Kosch A, Nohe N, Schobess R, Ehrenforth S, Childhood Thrombophilia Study Group. Risk of recurrent venous thrombosis in children with combined prothrombotic risk factors. *Blood*. 2001;97:858–862.
  91. Chiesa G, Hobbs HH, Koschinsky ML, Lawn RM, Maika SD, Hammer RE. Reconstitution of lipoprotein (a) by infusion of human low density lipoprotein into transgenic mice expressing human apolipoprotein (a). *J Biol Chem*. 1992;34:24369–24374.
  92. Sanan DA, Newland DL, Tao R, Marcovina S, Wang J, Mooser V, Hammer RE, Hobbs HH. Low density lipoprotein receptor-negative mice expressing human apolipoprotein B-100 develop complex atherosclerotic lesions on a chow diet: no accentuation of apolipoprotein(a). *Proc Natl Acad Sci U S A*. 1998;95:4544–4549.
  93. Callow MJ, Verstuyft J, Tangirala R, Palinski W, Rubin EM. Atherogenesis in transgenic mice with human apolipoprotein B and lipoprotein(a). *J Clin Invest*. 1995;96:1639–1646.
  94. Berg K, Svindland A, Smith AJ, Lawn RM, Djurovic S, Aleström A, Aleström P, Eliassen K. Spontaneous atherosclerosis in the proximal aorta of LPA transgenic mice on a normal diet. *Atherosclerosis*. 2002;163:99–104.
  95. Fan J, Shimoyama H, Sun H, Marcovina S, Honda K, Watanabe T. Transgenic rabbits expressing human lipoprotein(a) develop more extensive atherosclerotic lesions in response to a cholesterol-rich diet. *Arterioscler Thromb Vasc Biol*. 2001;21:88–94.
  96. Sun H, Unoki H, Wang X, Liang J, Ichikawa T, Arai Y, Shiomi M, Marcovina SM, Watanabe T, Fan J. Lipoprotein(a) enhances advanced atherosclerosis and vascular calcification in WHHL transgenic rabbits expressing human apolipoprotein(a). *J Biol Chem*. 2002;277:47486–47492.
  97. Palabrica TM, Liu AC, Aronovitz MJ, Furie B, Lawn RM, Furie BC. Antifibrinolytic activity of apolipoprotein(a) in vivo: human apolipoprotein(a) transgenic mice are resistant to tissue plasminogen activator-mediated thrombolysis. *Nat Med*. 1995;1:256–259.
  98. Williams JK, Bellinger DA, Nichols TC, Griggs TR, Bumol TF, Fouts RL, Clarkson TB. Occlusive arterial thrombosis in cynomolgous monkeys with varying plasma concentration of lipoprotein(a). *Arterioscler Thromb*. 1993;13:548–554.
  99. Sandholzer C, Saha N, Kark JD, Rees A, Jaross W, Dieplinger H, Hoppichler F, Boerwinkle E, Utermann G. Apo(a) isoforms predict risk for coronary heart disease: a study in six populations. *Arterioscler Thromb*. 1992;12:1214–1226.
  100. Kraft HG, Lingenhel A, Kochl S, Hoppichler F, Kronenberg F, Abe A, Muhlberger V, Schonitzer D, Utermann G. Apolipoprotein(a) kringle IV repeat number predicts risk for coronary artery disease. *Arterioscler Thromb Vasc Biol*. 1996;16:713–719.
  101. Gazzaruso C, Garzaniti A, Buscaglia P, Bonetti G, Falcone C, Fratino P, Finardi G, Geroldi D. Association between apolipoprotein(a) phenotypes and coronary heart disease at a young age. *J Am Coll Cardiol*. 1999;33:157–163.
  102. Bowden J-F, Pritchard PH, Hill JS, Frohlich JJ. Lp(a) concentration and apo(a) isoform size: relation to the presence of coronary artery disease in familial hypercholesterolemia. *Arterioscler Thromb*. 1994;14:1561–1568.
  103. Seed M, Hoppichler F, Reveley D, McCarthy S, Thompson GR, Boerwinkle E, Utermann G. Relation of serum lipoprotein(a) concentration and apolipoprotein(a) phenotype to coronary heart disease in patients with familial hypercholesterolemia. *N Engl J Med*. 1990;322:1494–1499.

104. Ruiz J, Thillet J, Huby T, James RW, Erlich D, Flandre P, Froguel P, Chapman J, Passa P. Association of elevated lipoprotein(a) levels and coronary heart disease in NIDDM patients: relationship with apolipoprotein(a) phenotypes. *Diabetologia*. 1994;37:585–591.
105. Parlavecchia M, Pancaldi A, Taramelli R, Valsania P, Galli L, Pozza G, Chierchia S, Ruotolo G. Evidence that apolipoprotein(a) phenotype is a risk factor for coronary artery disease in men <55 years of age. *Am J Cardiol*. 1994;74:346–351.
106. Lindén T, Taddei-Peters W, Wilhelmson L, Herlitz J, Karlsson T, Ullström C, Wiklund O. Serum lipids, lipoprotein(a) and apo(a) isoforms in patients with established coronary artery disease and their regulation to disease and prognosis after coronary by-pass surgery. *Atherosclerosis*. 1998;137:175–186.
107. Longenecker JC, Klag MJ, Marcovina SM, Powe NR, Fink NE, Giaculli F, Coresh J. Small apolipoprotein(a) size predicts mortality in end-stage renal disease. The CHOICE Study. *Circulation*. 2002;106:2812–2818.
108. Holmer SR, Hengstenberg C, Kraft HG, Mayer B, Poll M, Kurzinger S, Fischer M, Lowel H, Klein G, Riegger GAJ, Schunkert H. Association of polymorphisms of the apolipoprotein(a) gene with lipoprotein(a) levels and myocardial infarction. *Circulation*. 2003;107:696–701.
109. Mølgaard J, Klausen IC, Lassvik C, Færgeman O, Gerdes LU, Olsson AG. Significant association between low-molecular-weight apolipoprotein(a) isoforms and intermittent claudication. *Arterioscler Thromb*. 1992;12:895–901.
110. Kamboh MI, Ferrell RE, Kottke BA. Expressed hypervariable polymorphism of apolipoprotein(a). *Am J Hum Genet*. 1991;49:1063–1074.
111. Marcovina SM, Hobbs HH, Albers JJ. Relation between number of apolipoprotein(a) kringle 4 repeats and mobility of isoforms in agarose gel: basis for a standardized isoform nomenclature. *Clin Chem*. 1996;42:436–439.
112. Wu HD, Berglund L, Dimayuga C, Jones J, Sciacca RR, DiTullio MR, Homma S. High lipoprotein(a) levels and small apolipoprotein(a) sizes are associated with endothelial dysfunction in a multiethnic cohort. *J Am Coll Cardiol*. 2004;43:1828–1833.
113. Kronenberg F, Kronenberg MF, Kiechl S, Trenkwalder E, Santer P, Oberhollenzer F, Egger G, Utermann G, Willeit J. Role of lipoprotein(a) and apolipoprotein(a) phenotype in atherosclerosis. Prospective results from the Bruneck Study. *Circulation*. 1999;100:1154–1160.
114. Paultre F, Tuck CH, Boden-Albala B, Kargman DE, Todd E, Jones J, Paik MC, Sacco RL, Berglund L. Relation of apo(a) size to carotid atherosclerosis in an elderly, multiethnic population. *Arterioscler Thromb Vasc Biol*. 2002;22:141–146.
115. Orth-Gomér K, Mittleman MA, Schenck-Gustafsson K, Wamala SP, Eriksson M, Belkic K, Kirkeeide R, Svane B, Rydén L. Lipoprotein(a) as a determinant of coronary heart disease in young women. *Circulation*. 1997;95:329–334.
116. Ariyo AA, Thach C, Tracy RP. Lp(a) lipoprotein, vascular disease and mortality in the elderly. *N Engl J Med*. 2003;349:2108–2115.
117. Edelstein C, Pfaffinger D, Hinman J, Miller E, Lipkind G, Tsimikas S, Bergmark C, Getz GS, Witztum JL, Scanu AM. Lysine-phosphatidylcholine adducts in kringle V impart unique immunological and potential pro-inflammatory properties to human apolipoprotein(a). *J Biol Chem*. 2003;278:52841–52847.
118. Silaste ML, Rantala M, Alftan G, Aro A, Witztum JL, Kesäniemi YA, Hörkkö S. Changes in dietary fat intake alter plasma levels of oxidized low-density lipoprotein and lipoprotein(a). *Arterioscler Thromb Vasc Biol*. 2004;24:498–503.
119. Tsimikas S, Bergmark C, Beyer RW, Patel R, Pattison J, Miller E, Juliano J, Witztum JL. Temporal increases in plasma markers of oxidized low-density lipoprotein strongly reflect the presence of acute coronary syndromes. *J Am Coll Cardiol*. 2003;41:360–370.
120. Thillet J, Doucet C, Chapman J, Herbeth B, Cohen D, Faure-Delanef L. Elevated lipoprotein(a) levels and small apo(a) isoforms are compatible with longevity: evidence from a large population of French centenarians. *Atherosclerosis*. 1998;136:389–394.
121. Pepe G, Di Perna V, Resta F, Lovecchio M, Chimienti G, Colacicco AM, Capurso A. In search of a biological pattern for human longevity: impact of apo IV genetic polymorphisms on lipoproteins and the hyper-Lp(a) in centenarians. *Atherosclerosis*. 1998;137:407–417.
122. Matsubara M, Akita H, Shibuya H, Chiba H. Decreased longevity in Japanese men, associated with low-molecular-weight apolipoprotein(a) phenotypes. *Ann Clin Biochem*. 2001;38:120–123.
123. Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults: Executive summary of the third report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (ATP III). *JAMA*. 2001;285:2486–2497.
124. White AL, Lanford RE. Biosynthesis and metabolism of lipoprotein (a). *Curr Opin Lipidol*. 1995;6:75–80.
125. Steyrer E, Durovic S, Frank S, Gieβauf W, Burger A, Dieplinger H, Zechner R, Kostner GM. The role of lecithin: cholesterol acyltransferase for lipoprotein (a) assembly. *J Clin Invest*. 1994;94:2330–2340.
126. Hrzanjak A, Frank S, Wo X, Zhou Y, Van Berkel T, Kostner GM. Galactose-specific asialoglycoprotein receptor is involved in lipoprotein (a) catabolism. *Biochem J*. 2003;376:765–771.
127. Carlson LA, Hamsten A, Asplund A. Pronounced lowering of serum levels of lipoprotein Lp(a) in hyperlipidaemic subjects treated with nicotinic acid. *J Intern Med*. 1989;226:271–276.
128. Aylward EH, Li Q, Stine OC, Ranen N, Sherr M, Barta PE, Bylsma FW, Pearlson GD, Ross CA. Longitudinal change in basal ganglia volume in patients with Huntington's disease. *Neurology*. 1997;48:394–399.
129. Tassone F, Hagerman RJ, Loesch DZ, Lachiewicz A, Taylor AK, Hagerman PJ. Fragile X males with unmethylated, full mutation trinucleotides repeat expansions and elevated levels of FMR1 messenger. *Am J Med Genet*. 2000;94:232–236.
130. Tishkoff SA, Williams SM. Genetic analysis of African populations: Human evolution and complex disease. *Nat Genet*. 2002;3:611–621.

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