Cyclosporin A Enhances Interleukin-8 Expression by Inducing Activator Protein-1 in Human Aortic Smooth Muscle Cells

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Objective—Cyclosporin A (CsA) and tacrolimus (FK506) are widely used as immunosuppressants. However, their use has been hampered by various adverse effects, such as acceleration of atherosclerosis. Interleukin (IL)-8, a chemotactic cytokine, plays an important role in pathogenesis of atherosclerosis. We thus investigated whether synthesis of IL-8 from primary human aortic smooth muscle cells is influenced by CsA and FK506.

Methods and Results—Northern blot analysis revealed that CsA increased IL-8 mRNA level and enhanced its increase by epidermal growth factor or tumor necrosis factor-α. In contrast, FK506 had no effect on the mRNA level. IL-8 accumulation in culture media was also increased by CsA. Stability of IL-8 mRNA was not affected by CsA, whereas luciferase reporter gene assay using the human IL-8 promoter revealed that CsA significantly augmented the promoter activity. Electrophoretic mobility shift assay showed that binding activity of activator protein (AP)-1 was increased by CsA, and introduction of a mutation into the AP-1 site in the promoter abolished its CsA-dependent activation. The increased AP-1 binding activity was accompanied by c-Fos synthesis.

Conclusions—CsA stimulates synthesis of IL-8 via activation of AP-1 in human aortic smooth muscle cells, providing a novel aspect of biological effects of CsA on the cells. (Arterioscler Thromb Vasc Biol. 2003;23:2034-2040.)

Key Words: Cyclosporin A ■ tacrolimus ■ interleukin-8 ■ activator protein-1 ■ human aortic smooth muscle cells
present study, we therefore investigated whether synthesis of IL-8 from human aortic SMCs is influenced by CsA treatment. Our results show that, in contrast to most previous reports, CsA but not FK506 stimulates synthesis of IL-8 via activation of AP-1. It will be also shown that CsA enhances EGF-dependent and TNF-α-dependent synthesis of IL-8.

**Methods**

**Cell Culture**

Primary human aortic smooth muscle cells (HASMCs; Kurabo Industries Limited) were cultured in a growth medium, HuMedia-SB2 medium (Kurabo) supplemented with 5% FBS, 0.5 ng/mL human EGF, 2 ng/mL human fibroblast growth factor (FGF)-B, 5 μg/mL insulin, 50 μg/mL gentamicin, and 50 ng/mL amphotericin B. Nearly confluent HASMCs were then cultured for 4 days in a differentiation medium consisting of HuMedia-SB2 medium supplemented with 1% FBS, 30 μg/mL heparin, 50 μg/mL gentamicin, and 50 ng/mL amphotericin B. Note that the differentiation medium does not contain EGF nor FGF-B. The cells were treated with 2 μmol/L CsA or 2 μmol/L FK506 for 30 minutes, followed by incubation with 30 ng/mL human EGF or 40 ng/mL human TNF-α. After 3-hour incubation, the cells were harvested for electrophoretic mobility shift assay (EMSA) and Western blot analysis and after 6 hours of incubation, the cells were divided into 2 groups. One was cultured in the presence of CsA or 2 μmol/L FK506, not containing EGF nor FGF-B. The cells were treated with 2 μmol/L CsA or 2 μmol/L FK506 for 30 minutes, followed by incubation with 30 ng/mL human EGF or 40 ng/mL human TNF-α. After 3-hour incubation, the cells were harvested for electrophoretic mobility shift assay (EMSA) and Western blot analysis and after 6 hours of Northern blot analysis. CsA and FK506 were generous gifts from Novartis Pharma and Fujisawa Pharmaceutical, respectively. In an experiment using cycloheximide (CHX), it was added to the media at a concentration of 10 μg/mL 15 minutes before CsA treatment. Jurkat T cells were also used to study the effect of CsA. They were treated with 2 μmol/L CsA for 30 minutes, followed by 1 μmol/L calcium ionophore (A23187) and 30 nmol/L phorbol myristate acetate (PMA) for 6 hours. The cells were then harvested for Northern blot analysis. To analyze stability of IL-8 mRNA, the cells were stimulated with 30 ng/mL EGF for 6 hours. Then a RNA synthesis inhibitor, actinomycin D (ActD), was added to the media at a final concentration of 1 μg/mL. After 1 hour of incubation, the cells were divided into 2 groups. One was cultured in the presence of 2 μmol/L CsA, and the other in its absence. The cells were harvested for Northern blot analysis at 0, 1, 2, 3, and 4 hours after addition of CsA. The effect of 1 μmol/L A23187 and 30 nmol/L PMA on IL-8 mRNA stability was also studied in the same protocol.

**Northern Blot Analysis**

Total RNA was extracted by the acid guanidinium thiocyanate-phenol-chloroform method.29 Northern blotting and hybridization were performed as previously described.30 Briefly, 15 μg of total RNA was denatured, electrophoresed, and blotted onto a nylon membrane (Gene Screen Plus; New England Nuclear). Hybridization was carried out using human IL-8 and GAPDH cDNAs as probes.31 After washing the membrane, it was subjected to quantitative analysis using the BAS 2000 bioimage analyzing system (Fuji Film Co).

**EMSA**

Procedures for preparation of nuclear extracts and EMSA were described in our previous reports.23,24 Oligonucleotide sequences for EMSA probes are as follows. AP-1 oligonucleotide, 5'-GTTGATGACTGACTTTGCTC-3', contains a AP-1-binding site (−126 to −20, underlined) in the promoter of human IL-8 gene (DDBJ accession No. M28130). NF-kB oligonucleotide, 5'-GATCTGGAATTTCCTCTC-3', contains a NF-kB-binding site (−81 to −72, underlined). C/EBPβ oligonucleotide, 5'-GCCTACAGTGGAAATGGTCCG-3', contains a C/EBPβ-binding site (−93 to −84, underlined). Canonical AP-1 oligonucleotide, 5'-TGACTGCTAGCTGACTGTCCG-3', contains an AP-1 site in human collagenase promoter (DDBJ accession No. AF007878).34 To identify AP-1, NF-kB, and C/EBP subunits, supershift analysis was performed using antibodies directed against c-Fos, c-Jun, Fra-1, Fra-2, FosB, p50, p52, p65, c-Rel, and C/EBPβ (Santa Cruz). They were added to the binding reaction mixture after addition of the labeled probe and incubated for 1 hour at 4°C.

**Western Blot Analysis**

The cells were washed with PBS and harvested with 50 μL of preheated (85°C) 1× SDS gel loading buffer. After boiling, sonication, and centrifugation, the supernatants were used for Western blot analysis. Samples (100 μg of protein) were subjected to 12% SDS-polyacrylamide gel electrophoresis and transferred onto a nitrocellulose membrane (Hybond-ECL; Amersham Pharmacia). After blocking with 5% skimmed milk, the membrane was incubated with goat anti-c-Fos and c-Jun antibodies (Santa Cruz). After wash, the membrane was incubated with alkaline phosphatase–conjugated rabbit anti-goat IgG (Zymed). The color development was carried out using ready-to-use tablet (Roche).

**Measurement of IL-8 in the Media by ELISA**

The cells were grown to near confluence in a 60-mm dish and cultured in the differentiation medium for 4 days. In 2 mL of fresh medium, the cells were treated for 12 hours with 2 μmol/L CsA, 30 ng/mL EGF, and 100 U/mL TNF-α alone or in combination. Then IL-8 concentration in culture media was measured using an ELISA kit (human IL-8 ANLYZA Immunoassay Kit; Genzyme Techne).

**Luciferase Reporter Gene Assay**

A DNA fragment of the minimum promoter of the human IL-8 gene (DDBJ accession No. M28130) from the nucleotide positions −133 to +46 was amplified by polymerase chain reaction (PCR) and cloned into NheI/HindIII sites of the promoterless luciferase reporter vector pGL3-Basic (Promega). The plasmid was named p-133. Site-directed mutagenesis of the AP-1 binding site (−126 to −120) was performed by PCR using the following primer: 5'-GCTAGCGTGTGAAAGATCTGATTGTCCTGAGGG-3'. The first 6 nucleotides GCTAGC are Nhel site. The AP-1 site, TGACTCA, was mutated to AGATCTA (underlined). This plasmid was named p-133-mutAP1. HASMCs (5 × 10⁴ cells per well) were seeded in 6-well plates and cultured in the growth medium for 2 days. The cells were then transfected with p-133 or p-133-mutAP1 (300 ng/well) and SV40 promoter-driven β-galactosidase plasmid (100 ng/well, pSV-β-Galactosidase Control Vector, Promega) using Effectene transfection reagent (Qiagen). After transfection, the cells were cultured in the growth medium for 6 hours, and then the medium was replaced with the differentiation medium. After 24 hours, the cells were treated with 2 μmol/L CsA or 30 ng/mL EGF for another 24 hours. Then the cells were harvested and the luciferase and β-galactosidase activities were measured as described in our previous report.35 The luciferase activity was normalized by β-galactosidase activity.

**Reverse Transcriptase–PCR**

Procedures for reverse transcriptase (RT)–PCR were described in our previous reports.33 The primers for CXCR-1 and CXCR-2 were designed according to Hayes et al.,34 as follows: CXCR-1: 5'-GGGGCCACACAACTCTTC-3' and 5'-AGTGCTTGGCC-CTAATGCCTC-3'; CXCR-2: 5'-CCGGCGCTGTTGAGTGTAG-3' and 5'-TCTGGTTCGGTCTGTAATA-3'.

**DNA Synthesis**

DNA synthesis in HASMCs was assessed by measurement of incorporation of [3H]-thymidine into the cells. HASMCs were grown in 12-well culture plates to 70% confluence and then made quiescent with differentiation medium for 4 days. The cells were then treated with 2 μmol/L CsA or with 40 or 400 ng/mL of human recombinant IL-8 (Sigma) for 24 hours. [3H]-thymidine (1 μCi per well) was added to the media, followed by an additional 4-hour incubation. Incorporation of [3H]-thymidine was assessed by measurement of the radioactivity in the trichloroacetic acid–insoluble fraction of the cells, as described in our previous report.35
CsA but not FK506 increases IL-8 mRNA levels. HASMCs maintained in the differentiation medium for 4 days were treated with 2 μmol/L CsA (A) or 2 μmol/L FK506 (B) for 30 minutes followed by 30 ng/mL EGF or 40 ng/mL TNF-α for 6 hours. Northern blot analysis was performed using IL-8 and GAPDH cDNA as probes. Representative images by BAS 2000 system are shown. One experiment was done in duplicate. IL-8 mRNA levels were normalized by those of GAPDH mRNA. Quantitative results from 2 separate experiments were expressed as mean±SD (n=4). *P<0.05, **P<0.01 vs unstimulated cells. #P<0.01.

Figure 1. CsA but not FK506 increases IL-8 mRNA levels. HASMCs maintained in the differentiation medium for 4 days were treated with 2 μmol/L CsA (A) or 2 μmol/L FK506 (B) for 30 minutes followed by 30 ng/mL EGF or 40 ng/mL TNF-α for 6 hours. Northern blot analysis was performed using IL-8 and GAPDH cDNA as probes. Representative images by BAS 2000 system are shown. One experiment was done in duplicate. IL-8 mRNA levels were normalized by those of GAPDH mRNA. Quantitative results from 2 separate experiments were expressed as mean±SD (n=4). *P<0.05, **P<0.01 vs unstimulated cells. #P<0.01.

Statistical Analysis
Statistical analysis was carried out using 1-way fractional ANOVA, followed by Fisher’s protected least-significant difference analysis.

Results

CsA But Not FK506 Increases IL-8 mRNA Levels
Primary HASMCs cultured in the differentiation medium for 4 days were treated with EGF or TNF-α for 6 hours in the presence or absence of CsA. As shown in Figure 1A, Northern blot analysis revealed that the basal level of IL-8 mRNA was undetectable. Interestingly, IL-8 mRNA level increased significantly in the cells cultured in the presence of CsA. Treatment with either EGF or TNF-α also significantly increased the mRNA level. This EGF-dependent or TNF-α-dependent increase was significantly enhanced in the presence of CsA.

In contrast to CsA, no significant increase in IL-8 mRNA was observed in the cells cultured in the presence of FK506, as shown in Figure 1B. Also, no additive effect of FK506 on EGF-dependent or TNF-α-dependent increase in IL-8 mRNA was observed.

To study whether CsA-dependent increase in IL-8 mRNA requires de novo protein synthesis, the cells were pretreated with 10 μg/mL of a protein synthesis inhibitor, CHX, for 15 minutes, and then 2 μmol/L CsA was added to the media, followed by 6 hours of incubation. CHX alone markedly increased IL-8 mRNA levels. However, no additive increase was observed by combined treatment with CHX and CsA (data not shown), suggesting that de novo protein synthesis is necessary for the induction of IL-8 mRNA by CsA.

It should be noted that the CsA effect observed in HASMCs was not reproduced in human T cell–derived Jurkat T cells. Jurkat T cells were pretreated with 2 μmol/L CsA for 30 minutes, followed by 1 μmol/L calcium ionophore and 30 nmol/L PMA for 6 hours. The basal level of IL-8 mRNA was not increased by CsA. In accordance with a previous report,22 CsA inhibited the increase in IL-8 mRNA by combined treatment with calcium ionophore and PMA (data not shown).

CsA Stimulates Synthesis of IL-8
Concentrations of IL-8 in culture media produced from HASMCs were determined by ELISA. As shown in Figure 2, treatment of the cells with CsA for 12 hours resulted in a significant increase in IL-8 accumulation in culture media. Treatment with EGF or TNF-α also significantly increased IL-8 accumulation, and CsA additionally increased the levels. The IL-8 concentrations in culture media well reflect its mRNA levels shown in Figure 1A.

CsA Does Not Affect IL-8 mRNA Stability
Expression of human IL-8 gene is regulated at the transcriptional16 and posttranscriptional levels.19 We first evaluated effects of CsA on IL-8 mRNA stability, as shown in Figure 3. Expression of IL-8 mRNA was induced by EGF. Then a RNA synthesis inhibitor ActD was added and the cells were cultured in the presence or absence of CsA. Northern blot analysis revealed the long half-life of IL-8 mRNA in HASMCs; >75% of mRNA was detected after 6 hours of incubation with ActD in the absence of CsA. Similar levels of the mRNA were detected in the presence of CsA, demonstrating that CsA does not increase IL-8 mRNA stability. It was also found that stability of the mRNA induced by calcium ionophore and PMA was similar to that induced by EGF and that CsA had no effect on the stability (data not shown).

CsA Enhances DNA-Binding Activity of AP-1
It has been shown that the human IL-8 promoter sequence spanning nucleotides −133 to −1 containing AP-1, NF-κB,
CsA does not affect stability of IL-8 mRNA. The differentiated HASMCs were pretreated with 30 ng/mL EGF for 6 hours, followed by EGF plus 1 μg/mL ActD for 1 hour. The cells were then divided into 2 groups. One was cultured in the presence of 2 μmol/L CsA (○) and the other was cultured in its absence (□) for indicated periods. IL-8 and GAPDH mRNA levels were determined by Northern blot analysis, and the IL-8 mRNA levels were normalized by those of GAPDH mRNA. The experiment was done in duplicate. Similar results were obtained from a separate experiment.

and C/EBPβ binding sites is essential and sufficient for transcriptional regulation of the gene. We thus examined effect of CsA on DNA-binding activities of these 3 transcription factors by EMSA. Binding activity to the AP-1 site (−126 to −120) in human IL-8 promoter was shown in Figure 4A. Treatment of the cells with CsA substantially increased the AP-1 binding activity (lane 2) above the basal level (lane 1). EGF also increased the binding activity (lane 3), and CsA additionally increased the activity (lane 4). Although TNF-α alone had only marginal effect on AP-1 binding (lane 5), TNF-α together with CsA augmented the binding activity (lane 6). When supershift analysis was performed using the extracts obtained from the cells treated with EGF and CsA, the AP-1 band was supershifted by anti-c-Fos antibody (lane 7) and its density was markedly reduced by anti-c-Jun antibody (lane 8) compared with the density of lane 4. Anti-Fra1, anti-Fra2, and anti-FosB antibodies had no effect on the AP-1 binding (data not shown). These results indicate that the binding activity of AP-1 consisting of cFos/c-Jun heterodimer is increased by CsA.

Binding activity to the NF-κB site (−81 to −72) was shown in Figure 4B. Two retarded bands were evident in all lanes. The lower band with the asterisk represents nonspecific binding. When HASMCs were treated with TNF-α, the density of the upper band increased (lane 5) above the basal level (lane 1). This band was supershifted by anti-p56 antibody (lane 7) but not by anti-p50 (lane 8), p52, and c-Rel (data not shown) antibodies, suggesting that this NF-κB consists of p65 homodimer. The result is consistent with the previous report. On the other hand, EGF did not activate NF-κB (lane 3). In contrast to the AP-1 binding, CsA did not exchange NF-κB binding in the basal condition (lane 2) nor in the cells treated with EGF (lane 4) and TNF-α (lane 6).

The binding activity to the C/EBPβ site (−93 to −84) was also investigated by EMSA (data not shown). A single band was detected in unstimulated cells, and its density was markedly reduced by anti-C/EBPβ antibody. The density of the band was not altered by CsA, EGF, and TNF-α, indicating that C/EBPβ binding is not altered by treatment with CsA.

Taken together, these results suggest that TNF-α stimulates expression of IL-8 gene by activating NF-κB, whereas EGF does by increasing AP-1. Under both conditions as well as in unstimulated cells, CsA increases AP-1 binding to IL-8 promoter.

Mutation of the AP-1 Site in IL-8 Promoter Inhibits CsA-Dependent Promoter Activation

To confirm the role of AP-1 in CsA-dependent IL-8 gene expression, the plasmids containing a luciferase reporter gene driven by the minimum IL-8 promoter (p-133) or its mutant for AP-1 site (p-133-mutAP1) were constructed and a reporter gene assay was performed in HASMCs. As shown in Figure 5A, when p-133 was transfected, its basal activity was detected, and treatment of the transfected cells with CsA significantly increased the promoter activity by 4-fold. EGF also significantly increased the activity, and CsA increased it additionally. These results correlate well with those of Northern blot analysis (Figure 1A). In contrast, when p-133-mutAP1 was transfected, the stimulatory effects of CsA and EGF were abolished. These data indicate that the increased AP-1 binding to the AP-1 site located in −126 to −120 in human IL-8 promoter is responsible for CsA-dependent expression of IL-8 gene in HASMCs.
CsA Increases c-Fos Protein Synthesis

Western blot analysis was performed to evaluate effect of CsA on c-Fos and c-Jun protein levels in HASMCs. As shown in Figure 5B, a single band of 60 kDa was detected by anti-c-Fos antibody. The size of this band was compatible with that of the human c-Fos protein and different from those of other Fos family members such as Fra-1, Fra-2, and FosB. Treatment of the cells with CsA increased the c-Fos protein level. Treatment with EGF also increased the level, and combined treatment with CsA resulted in additional increase. Anti-c-Jun antibody detected a single band of 40 kDa protein. In contrast to c-Fos, c-Jun protein levels were not altered by CsA nor EGF. These results indicate that the AP-1 binding activities increased by CsA and EGF are attributable, at least in part, to the augmented c-Fos synthesis.

IL-8 Stimulates DNA Synthesis in HASMCs

IL-8 belongs to CXC chemokine family, and its action is exerted through the membrane receptors CXCR1 and CXCR2. Thus, expression of these receptors was examined in HASMCs. As shown in Figure 6A, when RT products from HASMCs cultured in the growth medium for 4 days were used, a single band was amplified by RT-PCR using CXCR1 primers and CXCR2 primers. The sizes of the bands were consistent with the expected lengths of the amplified products, 363 bp for CXCR1 and 385 bp for CXCR2, demonstrating expression of these receptors in HASMCs. Interestingly, when the cells were cultured in the differentiation medium, the densities of amplified bands were increased, suggesting that differentiation of HASMCs may stimulate expression of CXCR1 and CXCR2. On the other hand, no bands were amplified from the samples in which RT was omitted from reverse transcription reaction (data not shown), indicating that contamination of our RNA samples with genomic DNA was, if any, below the detectable level.

Effects of CsA and IL-8 on proliferation of HASMCs were then assessed by ³H-thymidine incorporation into the cells. As shown in Figure 6B, treatment of the cells with 2 μmol/L CsA or with 40 ng/mL IL-8 did not increase thymidine incorporation. However, 400 ng/mL IL-8 significantly stimulated thymidine incorporation of HASMCs.

Discussion

The present study for the first time demonstrates upregulation of IL-8 expression by CsA in HASMCs. Furthermore, EGF-dependent and TNF-α-dependent IL-8 expression in the cells is also enhanced by CsA. Although several reports have demonstrated inhibitory effect of CsA on IL-8 expression in various cell types, such as T cells, eosinophils, neutrophils, monocytes, keratinocytes, cardiac myxoma...
cells,27 and colon epithelial cells,28 and also in our experiment using Jurkat T cells, few reports have shown its stimulatory effect. In umbilical vein endothelial cells,39 airway epithelial cells,40 dermal fibroblasts,41 and synovocytes of patients with rheumatoid arthritis,42 CsA has been shown to increase the abundance of IL-8 mRNA or protein. However, none of these reports addressed the mechanisms by which CsA regulates the IL-8 expression.

Expression of human IL-8 gene is regulated at the transcriptional and posttranscriptional levels. Rapid degradation of certain transcripts has been ascribed to AU-rich sequences in their 3′-untranslated regions and to distinct proteins interacting and degrading the transcripts.43 AU-rich sequences are present in the 3′-untranslated region of human IL-8 mRNA, and its degradation has been shown to be subject to such modulation.19–21 We thus studied stability of IL-8 mRNA in the presence and absence of CsA using RNA synthesis inhibitor ActD. However, we found that CsA does not affect IL-8 mRNA stability in HASMCs. Unexpectedly, we found that IL-8 mRNA induced by EGF has a long half-life, compared with a report that the half-life of the mRNA induced by IL-1α is approximately 40 minutes in human endometrial stromal cells.20 To elucidate whether the long half-life of IL-8 mRNA depends on external stimuli used, stability of the mRNA induced by calcium ionophore and PMA was examined in HASMCs. It was found that the stability is similar to that induced by EGF (data not shown), indicating that the half-life of IL-8 mRNA in HASMCs is longer than that in endometrial stromal cells, which does not depend on stimuli.

The lack of effect of CsA on the mRNA stability suggests its stimulatory effect on IL-8 gene transcription. Indeed, it was demonstrated that CsA augmented IL-8 promoter activity. Introduction of a mutation into the AP-1 site in the promoter abrogated the CsA-dependent promoter activation. Consistent with this result, EMSA revealed the increase in binding activity of c-Fos/c-Jun heterodimer AP-1 by CsA in the presence and absence of EGF or TNF-α. These results demonstrate the essential role of AP-1 in transcriptional activation of IL-8 gene by CsA in HASMCs.

It was also demonstrated that the increased AP-1 binding activity was accompanied by c-Fos synthesis, indicating that CsA-dependent IL-8 gene expression is, at least in part, mediated by de novo c-Fos synthesis. This notion is compatible with the result that pretreatment with protein synthesis inhibitor CHX did not result in additive increase in IL-8 mRNA by CsA.

How does CsA increase expression of c-Fos? Elk-1, one of the Ets family of transcription factors, is activated on various external stimuli, such as serum and growth factors, and induces c-Fos gene expression by binding to the serum-responsive element in its promoter cooperatively with another transcription factor, serum response factor.44 The activity of Elk-1 is stimulated by extracellular signal–regulated kinase through phosphorylation of serine-383 in the C-terminal activation domain.45 Recently, it was shown that calcineurin specifically dephosphorylates the phospho-serine-383 in Elk-1 and suppresses the activities of Elk-1 and thereby c-Fos promoter.46 This report raises the possibility that the CsA-dependent increase in c-Fos expression might be attributable to inhibition of calcineurin activity by CsA. However, this mechanism is unlikely in HASMCs, because FK506, another calcineurin inhibitor, failed to increase IL-8 mRNA level in the cells. Additional studies are required to elucidate the mechanisms of CsA action on c-Fos.

Yue et al11 reported that IL-8 stimulates proliferation of rat and human SMCs. Also, Simonini et al47 showed that neutralization of IL-8 in culture media by its antibody inhibits growth of human coronary SMCs. However, IL-8 action on SMCs is ambiguous, because some reports indicated that IL-8 binding and its receptor expression were not detected in human SMCs.36,48 In contrast to these observations, it was clearly demonstrated that HASMCs express CXCR1 and CXCR2 and that 400 ng/mL IL-8 increased DNA synthesis. These results indicate the presence of functional receptor for IL-8 in HASMCs. The controversy of expression of CXCR1 and CXCR2 may be attributable to culture conditions for SMCs, because our result indicated that the presence of 5% FBS, EGF, and FGF-B in culture media tended to reduce the expression of CXCR1 and CXCR2. On the other hand, CsA had no effect on DNA synthesis. This may be attributable to the low level of IL-8 production by CsA or antiproliferative effects of CsA on HASMCs.

The IL-8 concentration required for the increase in DNA synthesis is 400 ng/mL, which is much higher than the concentration in culture medium observed after the combined treatment with CsA and TNF-α for 12 hours (44 ng/mL in Figure 2). However, it is possible that CsA-dependent IL-8 production in the microenvironment in vivo could result in the local concentation much higher than that in cell culture system. Additional studies are thus required to confirm the stimulatory effect of CsA on proliferation of HASMCs.

In conclusion, the present study demonstrates that CsA increases IL-8 gene expression by stimulating AP-1 DNA binding after c-Fos synthesis in HASMCs. These findings provide a novel aspect of biological effects of CsA on the cells. In addition, considering the role of IL-8 in development of atherosclerosis,7,9,10 the enhanced IL-8 expression by CsA in HASMCs could be one of the mechanisms for acceleration of posttransplant atherosclerosis.

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