Characterization of Human Glycoprotein VI Gene 5′ Regulatory and Promoter Regions

Kenichi Furihata, Thomas J. Kunicki

Objective—Platelet glycoprotein VI is a collagen receptor belonging to the immunoglobulin-like protein family that is essential for platelet interactions with collagen and is exclusively expressed in the megakaryocytic lineage. The objective of this study was to characterize the human glycoprotein VI gene (GP6) 5′ regulatory and promoter regions.

Methods and Results—We first used 5′ RACE to establish experimentally that the major transcription start site lies 28 bp upstream from the start codon. We next subcloned the 5′ regulatory region of GP6 into pGL3-basic [pGL3(−1576)] and used deletion mutagenesis to identify important regulatory regions, comparing the activity of transiently expressed promoter-luciferase constructs in Dami and HeLa cells. We found that megakaryocyte lineage-specific transcription is largely controlled within the segment −191/−39. By site-directed mutagenesis, we confirmed that a GATA-1 site at −176 and an Ets-1 site at −45 play important roles in the regulation of GP6 transcriptional activity.

Conclusions—We have determined that the GP6 sequence −191 to −39 represents the core promoter and that transcription is driven largely by GATA-1 (−176) and c-Ets-1 (−45) sites within this segment. (Arterioscler Thromb Vasc Biol. 2002;22:1733-1739.)

Key Words: gene regulation ▪ platelets ▪ signal transduction

Platelet glycoprotein VI (GPVI) is a member of the immunoglobulin-like protein family that is an essential platelet receptor for collagens and is exclusively expressed in the megakaryocytic lineage.2,3 We recently reported a variation in platelet GPVI content among normal healthy individuals for predisposition to thromboembolic disorders. From a clinical standpoint, it would be important to ascertain the risk for hemorrhagic or thromboembolic disorders. From a genetic perspective, the observed variation in platelet GPVI content may influence the number of common characteristics. In each case, the core regulatory region and promoter properties. Considering the critical role of GPVI in collagen-initiated signal transduction and platelet procoagulant activity, the observed variation in GPVI content may influence risk for hemorrhagic or thromboembolic disorders. From a clinical standpoint, it would be important to ascertain the mechanism(s) that controls GPVI content and to screen individuals for predisposition to thromboembolic disorders.

The promoter regions of megakaryocytic-specific genes, such as the integrin subunit αmβ3,6 platelet factor-4,7 platelet glycoprotein V,8 and platelet glycoprotein Ibα,9 share a number of common characteristics. In each case, the core promoter is generally short (≤500 bp), it lacks CCAAT or TATA consensus sequences, and it has functional GATA, Sp-1, and Ets cis-acting elements.10,11 GATA and Ets factors are not exclusive to the megakaryocytic lineage but are also critical for expression of erythroid-restricted and other unrelated genes, such as platelet and endothelial cell adhesion molecule-112 and P-selectin.13

Among megakaryocyte-specific genes, the αmβ3 promoter has been the best studied. The human αmβ3 promoter has a positive regulatory element in its 5′ region (from −598 to −400), wherein there are an Ets binding motif at −515 and a GATA-1 binding motif at −463. Both of these elements seem to play important roles in the expression of αmβ3.6 Three groups have postulated that there is a silencing element in the αmβ3 promoter that suppresses expression of the αmβ3 promoter in all cells except those of the megakaryocyte lineage.5,14,15 However, Era et al,6 using in vitro differentiation of murine D3 embryonic stem cells, recently refuted this observation and observed that solely a positive regulatory element in the region from −598 to −400 is sufficient for megakaryocyte lineage-specific expression of the αmβ3 gene.6 Likewise, Albanese et al16 reported that the GATA element at −463 and a flanking region of the human αmβ3 promoter were sufficient for restoring megakaryocyte lineage-specific expression of the murine αmβ3 gene in human cell lines. On the basis of these observations, it seems likely that positive regulatory elements play a prominent role in defining the lineage specificity of the human αmβ3 promoter.

To elucidate the mechanism of megakaryocyte lineage-specific expression of GPVI, we first excised a 1.5-kb...
fragment of the GPVI gene (GP6) 5' regulatory region from a Bac clone harboring the human gene (Clone 211(12 Incyte Genomics) and inserted this segment into pGL3-basic, upstream from a luciferase reporter gene. By luciferase assays and DNase I footprinting, we determined that a region of GP6 spanning from −191 to −39, which contains 1 GATA-1 and 1 Ets-1 binding site, is necessary for the transcriptional activity of GP6 in transfected cell lines. From a comparison of genomic DNA of 17 normal volunteers, 3 dimorphisms in the GP6 5' regulatory region were found to be in linkage disequilibrium, defining 2 major haplotypes.

**Methods**

Cell Lines and Blood Samples

The human cell line HeLa was purchased from the American Type Culture Collection (Manassas, Va). A Dami cell clone was received as a gift from Dr David Wilcox (Medical College of Wisconsin, Milwaukee). These cell lines were grown in DMEM supplemented with 10% FBS (HeLa) or Iscove's medium supplemented with 10% horse serum (Dami). Blood samples were obtained by medical personnel of The Scripps Clinic General Clinical Research Center from normal human volunteers, as described previously, with informed consent.

5' RACE

Online Table I (please see http://www.ahajournals.org) summarizes the GP6-specific oligonucleotide primers that are referred to in the present study. Total RNA was extracted from washed platelets with the use of an RNAqueous RNA extraction kit (Ambion) according to the manufacturer's instructions. First-strand cDNA was synthesized by using a RETROscript reverse transcription–polymerase chain reaction (PCR) first-strand synthesis kit (Ambion) and primer GSP1 (5'-CCATGATCCCTCTTGATGAT-3'), the remaining RNA was digested with a combination of RNase H and RNase T1 (GIBCO-BRL), and then the GP6-specific first-strand cDNA was purified with the use of a GeneClean III kit (BIO 101) and tailed with dCTP by use of terminal deoxynucleotidyl transferase (Roche Molecular Biochemicals). The tailed cDNA was amplified by PCR with the use of 5' RACE anchor primer (GIBCO-BRL, 5'-GGCCACGCGTCGACTAGTACGGGIGGGIIGGGIIG-3') and primer GSP2 (5'-CGCGTAGTGGACGTCGAGC-3'), followed by a second round of PCR with the use of Abridged Universal Amplification Primer (GIBCO-BRL, 5'-GGCCACGCGTGCAACTAGTACGGGIGGGIIGGGIIG-3') and primer GSP3 (5'-GGAGGGTCCCTGCTGCAGAAGC-3'). The PCR product was gel-purified, subcloned into pGEM-T Easy vector (Promega), and sequenced.

Comparison of Human and Murine GP6 Sequence

The human and murine GP6 sequences were retrieved from the GenBank high-throughput gene sequence (htgs) database (human clone RP11-700BS5 and mouse clone RP23-106A10, respectively) by using the appropriate CDNA as query sequence and the BLAST algorithm (available at http://www.ncbi.nlm.nih.gov/BLAST/blas t_t_references.html). The sequences of these Bac clones are not yet fully refined, and contigs within the obtained sequences are unordered. On the basis of the corresponding cDNA sequence, we established the correct order of the genomic contigs before the corresponding sequences from each species were compared with one another with use of the Vista software package (available at http://www-gsl.lbl.gov/VISTA/).

Generation of Human GP6 Promoter Constructs

Online Table II (please see http://www.ahajournals.org) summarizes the GP6 promoter constructs created for the present study. A Bac clone harboring human GP6 was identified at Incyte Genomics, Inc, by PCR-based screening of a human genomic library with the use of primer 6.1 (5'-GAGAAGCAGCTGACCCCTCGGTG-3') and primer GSP3, generated in our laboratory. The proximal 5' region of GP6 (−1576 to 75) was then amplified from this Bac clone DNA by PCR with the use of primers GP6p1F (5'-CCGGTACCTGGAGACAGAATCTGGTCCTG) and GP6pR (5'-CTCGAAGCCGTCAGGTCGAAGC-3'). The amplified PCR product was subcloned into pGEM-T easy vector (Promega) and sequenced. A 1651-bp KpnI-Xhol fragment with the correct sequence was then excised from this vector and subcloned into the luciferase reporter plasmid pGL3-basic (Promega), creating pGL3(−1576). A shorter 1577-bp XhoI-HindIII DNA fragment spanning −1502 to 75 was obtained from pGL3(−1576) and cloned into pGL3-basic, creating pGL3(−1502). This construct was subsequently used as a template for generation of a series of 5'-end unidirectional truncation mutants. In brief, pGL3(−1502) was first digested with a combination of KpnI and XmnI, and then its 5' end was unidirectionally truncated by using the Erase-A-Base Kit (Promega), creating pGL3(−1325), pGL3(−937), and pGL3(−539). An internal deletion mutant was constructed by PCR-based 2-step mutagenesis from pGL3(−1502) such that this mutant, pGL3(del(−191)), bears a deletion of the segment between −191 and −39. To more precisely locate the core promoter of GP6, 4 overlapping DNA fragments were amplified by PCR from the genomic DNA of each of 1 high-content and 1 low-content donor by using 1 of the 4 forward primers: −322 (5'-CCCCGGGAGGACCTTCAAGAGTGTGTA-3'), −191 (5'-CCCCCCGGAGGCTGTCAGAAGGAGAATCTGGTA-3'), −159 (5'-CCCCGGGGAGGCTTCGGAGAAAGTAGC-3'), and −38 (5'-CCCCGGGGCTCAGGTCAGAAGGAGAAC-3') together with the common reverse primer, pGL3R. After nucleotide sequencing of the DNA fragments subcloned in pGEM-T easy, the 4 GP6 promoter constructs were each subcloned into pGL3-basic, and the resultant constructs were designated pGL3(−322), pGL3(−191), pGL3(−159), and pGL3(−38). The substitution mutant pGL3(ΔGATA) was made by PCR-based mutagenesis from pGL3(−322), whereby the consensus GATA motif at −176 was mutated to GAGC. Similarly, in the mutant pGL3(ΔEts), the Ets-1 site at −45 was mutated from ACATCTCTG to ACACTCTG, whereas in the combined mutant pGL3(ΔGATA/ΔEts), the GATA-1 and Ets-1 sites were mutated. The sequences of all mutants were confirmed before use.

The murine counterparts of the core promoter regions −193 to 75 and −159 to 75 were amplified from mouse genomic DNA by PCR with the use of corresponding primers designed from the sequence of Mus musculus clone RP23-106A10 (GenBank accession No. AC087129). After verifying sequences, these were also subcloned into pGL3-basic to generate pGL3(m(−193) and pGL3(m(−159).

**Transient Expression Analysis of GP6 Promoter Activities by Luciferase Assay**

Dami cells were transfected with 1 of the pGL3-GP6 promoter constructs together with the Renilla luciferase reporter vector pRL-TK (Promega) by using the Effectene transfection reagent (Qiagen). In brief, 5×10⁴ Dami cells were transfected with a combination of 0.1 μg of pRL-TK and 0.2 μg of 1 of the pGL3-GP6 promoter constructs. Additional cells were transfected with the pGL3-control vector (Promega) and pRL-TK in the same medium. The cells were cultured for 48 hours in the presence of the DNA mixture and then harvested. Firefly and Renilla luciferase activities were measured on a luminometer by using the Dual-Luciferase Reporter Assay System (Promega) according to the manufacturer's instructions. To normalize for transfection efficiency, the promoter activity of each construct was expressed as the ratio of firefly luciferase activity to Renilla luciferase activity. When experiments in different cell lines were compared, the results were expressed as the ratio of GP6 activity to pGL3-control activity.

DNase I Footprinting

To confirm the occupancy of putative transcription factor binding elements in the GP6 core promoter region, we performed DNase I–based footprinting with the use of nuclear extracts from Dami and HeLa cells. The nuclear extract of HeLa cell (HeLaScribe Nuclear
Extract) was purchased from Promega, and that of Dami was prepared by the method of Dignam et al. The footprinting reaction was performed by using the Core Footprinting System (Promega) according to the manufacturer’s instructions with minor modifications. Briefly, 5 μg purpl3 (−322) was digested with XhoI and then dephosphorylated at its 5’ ends with calf intestine alkaline phosphatase. After 3P-labelling of both 5’ ends with the use of T4 polynucleotide kinase, the DNA fragment was further digested with XhoI, and the resultant 397-nucleotide DNA fragment (probe) was gel-purified. This probe was incubated either with 20 or 40 μg HeLa nuclear extract, or 10 μg Dami nuclear extract, or 40 μg HeLa nuclear extract for 30 minutes at ambient temperature; a control was incubated in the absence of nuclear extract. The DNA–protein mixture was subjected to RQ1 RNase-free DNase digestion for 2 minutes at ambient temperature. Each reaction mixture was extracted with phenol–chloroform and subjected to electrophoresis on a 6% denaturing sequencing acrylamide gel to obtain the footprinting reaction products. Briefly, 5 mmol/L EDTA. After electrophoresis, the gel was dried and exposed to x-ray film with an intensifying screen at −70°C.

EMSA for GATA-1 at −176 and Ets-1 at −45

Double-stranded DNA probes were obtained by annealing the following individually synthesized single-stranded oligonucleotides: for GATA −176 (nucleotides −185 to −166, 5′-GCCAGAGGAGATAAGGC CGG-3’) for Ets −45 (nucleotides −51 to −32, 5′-CATCATACACCTCTGAGCCCTGTT-3’). The probes were 5’-end-labeled with 3P before electrophoretic mobility shift assay (EMSA). The following double-stranded competitors were similarly synthesized: GATA mutant (5′-GCCAGAGGAGGAGCC GCGG-3’), GATA consensus (5′-CAGGAGGATACGAAATGCAGA-3’), GATA Gel Shift Oligonucleotides, Santa Cruz Biotechnology), Ets mutant (5′-CATCAACACCTCGAGCCCTGTT-3’), Ets consensus (5′-GATCATGACGGAGGAAAGTTCGA-3’), Ets-1/PEA3 Gel Shift Oligonucleotides, Santa Cruz Biotechnology), and PUL1 consensus (5′-GGGCTGTGGAGGAGAAATGCAGA-3’). The same HeLa and Dami nuclear extracts prepared for DNase I footprinting were used in this assay.

Each 3P-labeled probe (50 fmol) was mixed with 10 μg nuclear extract, 2 μg poly(dl-dC) (Amersham Pharmacia Biotech), and 10-fold or 100-fold molar excess of competitor in the binding buffer (25 mmol/L HEPES, pH 7.9, containing 10% [vol/vol] glycerol, 50 mmol/L KCl, 0.5 mmol/L EDTA, 0.5 mmol/L phenylmethylsulfonyl fluoride, and 0.5 mmol/L dithiothreitol) and incubated at ambient temperature for 30 minutes. Samples were then analyzed in a 4% or 5% polyacrylamide gel with the use of a running buffer of 50 mmol/L Tris and 0.38 mol/L glycine, pH 8.3, containing 5 mmol/L EDTA. After electrophoresis, the gel was dried and exposed to x-ray film.

Nucleotide Sequence Variation in the 5′ Regulatory Region of GP6

Genomic DNA was isolated from peripheral blood leukocytes of each of 1 high-content and 1 low-content GPVI donor1 by using the Puregene DNA isolation kit (Blood kit, Gentra Systems). A DNA fragment of 1.5 kb corresponding to the GP6 5′ regulatory region was amplified by PCR by using primers Kpnl-GP6P1F (5′-CGGCTACCTGAGGACGAAATCTCGCTGTTGTT-3’) and BglII-GP6P1R (5′-CGAGATCTACACAGGATCGATGTTGTAAG-3’) and then directly sequenced.

We identified a T/C dimorphism at −126 and 2 nearby dimorphisms of G/G at −1353 and T/C at −1350 between the high- and low-content GP6 donor sequences. To obtain genotypic frequencies of these dimorphisms, we similarly extracted genomic DNA from 15 more subjects. A genomic DNA fragment encompassing the 2 nearby dimorphisms was amplified by seminested PCR with the use of primers GP6P1F and GP6P1R in a primary PCR reaction and then GP6P1F and GP6P4R (5′-AGCCTGGGCAACACAGTAAA-3’) in a secondary, seminested PCR reaction. The 196-bp DNA product was directly sequenced to identify the −1536 G/A and −1530 T/C dimorphisms. Another T/C dimorphism at −126 was not genotyped by NaeI digestion of a 444-bp PCR product obtained with primers GP6P2F (5′-TGGCGAAAAAGAGACGTACT-3’) and GP6P4R. NaeI cuts the DNA fragment bearing −126C (GCCGGC), yielding 200-bp and 244-bp fragments in individuals homozygous for −126C. In individuals homozygous for −126T, a single 444-bp fragment is obtained. In individuals heterozygous for this dimorphism, 3 fragments are obtained: at 444, 244, and 200 bp.

Results

5′ RACE

A 330-bp DNA fragment was obtained after the final round of PCR. Three of the 5 sequenced clones began at an adenine located 28 bases upstream from the translation start codon ATG (Figure 1); the remaining 2 clones began at a cytosine located adjacent to and 1 base downstream from the aforementioned adenine. Consequently, we assigned the major transcription start site for GP6 to the adenine located 28 bases upstream from the ATG. This experimental finding confirms the transcription start site predicted in the report of Ezumi.18 This transcription start site is only 3 bases upstream from the original GP6 cDNA sequence obtained by primer extension.2 Throughout the present article, all of the nucleotides in the
Firefly luciferase to Renilla luciferase Ratio

Figure 2. Transient expression analysis of the GP6 promoter in Dami and HeLa cells. A series of GP6-luciferase constructs were generated by 5’ end unidirectional truncation. Each construct is identified regarding the base pair number at the 5’ end relative to the transcription start site. Each bar represents normalized relative luciferase activity in Dami cells (solid bar) or HeLa cells (open bar) and is the mean of 3 independent experiments. Error bars represent 1 SD. The activity of each GP6 promoter construct is expressed as a ratio of that to SV40 promoter-driven pGL3-control.

GP6 5’ regulatory region are numbered relative to the adenine base transcription start site, now designated 1.

Vista Analysis of Homologous Regions Between Human and Murine GP6
Both the GP6 sequences of the Mus musculus clone RP23-106A10 (GenBank accession No. AC087129) and Homo sapiens chromosome 19 clone RP11-700B5 (GenBank accession No. AC019238) were reordered regarding the corresponding cDNA sequences. By comparing the murine and human GP6 sequences with use of the Vista software package, we were able to identify highly homologous sequences in the region immediately 5’ to exon 1. Within the proximal portion of the GP6 5’ regulatory region, there is 1 conserved noncoding sequence (CNS) in which the human and murine sequences exhibit 77% identity over a 257-bp segment that spans from −182 to 75 (human sequence numbering; online Figure I, available at http://www.ahajournals.org). The high degree of homology within this sequence between these evolutionarily disparate species implies an important function for this region of the GP6 gene.

Characterization of the 5’ Regulatory Region of GP6 by Luciferase Reporter Assay
The plasmid pGL3(−1576) contains a segment of the GP6 5’ regulatory region DNA from nucleotide −1576 to 75 that includes the 5’ regulatory region and exon 1, which encodes the first 11 amino acids of the signal peptide, as well as a part of intron I. A second plasmid, pGL3(−1502), contains the same GP6 DNA but spans from −1502 to 75. A series of 5’ unidirectional deletion mutants (online Table II) were derived from pGL3(−1502), as described in Methods, and tested for their ability to drive the luciferase reporter gene in transiently transfected Dami cells. Figure 2 summarizes the results of these transfection studies.

Relative to pGL3(−1576), pGL3(−1325) showed a 50% decrease in promoter activity, whereas pGL3(−937) exhibi-

Figure 3. Determination of GP6 core promoter sequence. An internal deletion mutant, pGL3-del(−191 to −39), and 5’ truncation mutants were generated from a parent GP6-luciferase construct, pGL3(−1502), as described in Methods. Dami cells were transiently transfected with each of the promoter–luciferase reporter gene constructs together with pRL-TK, and firefly and Renilla luciferase activities were determined. Mean (solid bars) and 1 SD are shown.

uted a 24% increase in activity. These findings suggest that negative regulatory elements are located in a region between −1325 and −937. Relative to pGL3(−937), pGL3(−191) exhibited an ≈50% decrease in activity, and pGL3(−159) showed a >75% decrease in activity. These results indicate that the GP6 core promoter elements are located downstream from −191 and that there is an essential positive control element within the sequence spanning from −191 to −159. Qualitatively similar findings were obtained in K562 cells transfected with the same reporter constructs (not shown).

To evaluate cell-type specificity of the GP6 promoter, the same promoter constructs were transfected into HeLa cells, and luciferase activities were similarly measured. Although pGL3(−937), pGL3(−539), and pGL3(−322) showed very weak promoter activities in HeLa cells, pGL3(−191) and pGL3(−159) showed little if any activity (Figure 2). Mutation of GATA at −176 decreased the promoter activity of pGL3(ΔGATA) in Dami cells compared with its parent construct, pGL3(−322), but showed no effect on the promoter activity in HeLa cells.

Contribution of GATA and Ets Elements to GP6 Core Promoter Activity
The deletion of the sequence −191 to −39 from pGL3(−1502) resulted in a 62% decrease in the activity of pGL3-del(−191) in transfected Dami cells, confirming that a major positive regulatory element resides within this segment (Figure 3). Within the −191/−39 region, there is 1 GATA-1 binding site at −176 and 1 Ets-1 binding site at −45 (Figure 1). To measure the contribution of these GATA-1 and Ets sites to promoter activity, we generated and tested 3 shorter promoter constructs, pGL3(−191), pGL3(−159), and pGL3(−38), in Dami cells. Although pGL3(−191) retained ≈80% of the activity of pGL3(−1502), deletion of a segment from −191 to −160, which contains the GATA-1 site at −176, decreased the promoter activity of pGL3(−191) by 4-fold. A further removal of the sequence from −159 to −39 decreased the promoter activity of pGL3(−38) to background
The murine GP6 sequence corresponding to the human sequence −191 to −160 is also a positive regulatory element (online Figure II, available at http://www.ahajournals.org). This segment also contains a T/C dimorphism at −126 that is in linkage disequilibrium with 2 dimorphisms, −1536 G/A and −1530 T/C. The −126T and the −126C haplotypes were transfected into Dami cells in the context of pGL3(−322) and pGL3(−159). In either case, no significant differences were observed between promoter activities of the 2 haplotypes (data not shown).

Site-directed mutagenesis confirmed the positive regulatory role of the GATA-1 site at −176 and the Ets-1 site at −45. As illustrated in Figure 4, substitution mutation of the GATA-1 site decreased the promoter activity by 30%, whereas that of the c-Ets-1 site decreased the promoter activity by 40%. The combined mutation of the GATA-1 and c-Ets-1 sites decreased the promoter activity by 3-fold, suggesting an additive effect of these transcription factors.

Transcription Factor Binding Sites Within the 191 Nucleotides Preceding the Transcription Start Site
A number of transcription factor binding motifs are present in the putative core promoter sequence of GP6 (Figure 1). To investigate whether any of these motifs can be bound by nuclear proteins from megakaryocytic lineage cells, we performed DNase I footprinting (Figure 5). The GATA-1 site at −176 was clearly occupied by a Dami nuclear protein(s) but not by HeLa nuclear proteins, suggesting in vivo involvement of the GATA-1 site for megakaryocyte lineage–specific expression of GPVI. However, other predicted transcription factor binding motifs, including c-Rel and PU.1, were apparently not occupied by Dami nuclear proteins in the DNase I footprinting. The c-Ets-1 motif at −45 might be protected, inasmuch as 2 bands indicated by arrowheads in Figure 5B were present in lane 4 but absent in lanes 2 and 3. However, absence of the bands in lane 1(control) makes it difficult to conclude that the c-Ets-1 site is actually protected by Dami nuclear protein.

Discussion
GPVI is a platelet-specific collagen receptor that belongs to the killer cell receptor subgroup of the immunoglobulin gene family and cooperates with integrin αβ3 in vivo to effect optimal collagen-induced platelet adhesion and activation.

In many respects, the GP6 5′ regulatory and promoter regions resemble those of other genes encoding megakaryocyte lineage–specific proteins. Most notably, the GP6 promoter does not have a TATA box but contains functional GATA-1 and Ets-1 binding motifs. In luciferase reporter assays using 5′ unidirectional deletion mutants of the GP6 5′ regulatory region, we determined that the core promoter of GP6 lies within the sequence from −191 to −39. This region...
is highly conserved between human and mouse GP6, as revealed by Vista analysis, and there is roughly 77% identity within a 259-bp segment from position 182 to 75 (human sequence numbering).

One GATA-1 site at −176 and 1 c-Ets-1 site at −45 were identified in the core promoter. Deletion of the sequence −191 to −160 significantly decreased activity in transfected Dami cells, indicating that an essential positive regulatory element exists within this sequence. Several lines of evidence confirmed that GATA-1 at −176 is the essential element. First, mutation of the GATA-1 site decreased the promoter activity in Dami cells by 30%. Second, this GATA-1 motif is fully conserved between the human and murine sequence. Third, binding of Dami cell nuclear protein to this sequence has no effect. To further evaluate a possible contribution of these haplotypes to the control of platelet GP6 promoter levels, we compared haplotype frequencies in a group of 17 randomly selected donors. There is no statistically significant correlation between these haplotypes and platelet GPVI content (data not shown). Because platelet GPVI levels are correlated with integrin αbβ3 densities and because the appearance of GPVI during human megakaryocyte maturation coincides temporally with the appearance of integrin αbβ3, it is possible that there is a common mechanism involved in the coordinated expression of these important collagen receptors.

The characterization of the GP6 5' regulatory region, as described in the present study, represents an important preliminary step in the elucidation of the precise molecular mechanisms responsible for the control of platelet GPVI levels.
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