Prostacyclin Analogues Inhibit Tissue Factor Expression in the Human Monocytic Cell Line THP-1 Via a Cyclic AMP-Dependent Mechanism

David J. Crutchley, Denis E. Solomon, and Lobelia B. Conanan

Increased expression of tissue factor procoagulant by peripheral blood monocytes has been implicated in a number of thrombotic disorders. The present studies were undertaken to determine whether stable analogues of prostacyclin, a potent endothelium-derived platelet inhibitor and vasodilator, could inhibit tissue factor expression by human monocytic cells. Exposure of monocytic tumor THP-1 cells to 100 ng/ml endotoxin, 2 units/ml interleukin-1β, or 5 ng/ml tumor necrosis factor-α for 4 hours led to increased tissue factor procoagulant activity. Preincubation for 30 minutes with iloprost, ciprostene, and carbacyclin led to a dose-dependent inhibition of tissue factor expression induced by all three challenging agents. Iloprost was the most potent: 50% inhibition occurred at 5 nM, a concentration close to the reported dissociation constant for iloprost binding to the platelet prostacyclin receptor. An orally active analogue, cicaprost, was equally effective against endotoxin-induced tissue factor expression. Carbacyclin and ciprostene were 100 times less potent. Iloprost prevented the endotoxin-induced expression of tissue factor antigen on the surface of THP-1 cells, as determined by flow cytometry. Iloprost (500 pM–50 nM) increased intracellular levels of cyclic AMP. This effect was potentiated by isobutylmethylxanthine, an inhibitor of phosphodiesterase. The inhibitory effects of iloprost on tissue factor expression were also potentiated by isobutylmethylxanthine and mimicked by forskolin and dibutyryl cyclic AMP but not dibutyryl cyclic GMP. These results suggest that prostacyclin may play a role in downregulating tissue factor expression in monocytes, at least in part via elevation of intracellular levels of cyclic AMP. (Arteriosclerosis and Thrombosis 1992;12:664–670)

KEY WORDS • iloprost • interleukin-1β • tumor necrosis factor-α • cytokines • thromboplastin • cyclic AMP

Exposure of peripheral blood monocytes to a variety of agents leads to the expression of tissue factor, a membrane-bound glycoprotein that plays a critical role in blood coagulation as the cellular receptor and cofactor for factor VIIa.1,2 The agents include bacterial endotoxin;3 antigen–antibody complexes4; activated complement fragments5-6; and cytokines, such as the T-cell–derived monocyte procoagulant–inducing factor,7 interleukin-1 (IL-1),8-10 and tumor necrosis factor (TNF),8,9,11 although the efficacy of the latter two has been questioned.7 Monocyte tissue factor expression is thought to be a result of gene transcription and mRNA stabilization.12,13 At present, the physiological factors that downregulate these processes are not well identified.

We have previously shown that iloprost, a stable analogue of prostacyclin,14 is a potent inhibitor of endotoxin-induced tissue factor expression in human peripheral blood monocytes and a monocytic tumor cell line, THP-1.15 These preliminary studies suggest a novel role for prostacyclin in addition to its well-established role as a vasodilator and a platelet inhibitor (for review, see Reference 16). The present studies were undertaken to determine whether the effects of iloprost are 1) shared by other prostacyclin analogues and 2) expressed against tissue factor induction by agents other than endotoxin. In addition, because both iloprost and prostacyclin stimulate adenylate cyclase,17-20 we have explored the role of adenosine 3',5'-cyclic monophosphate (cAMP) in mediating the inhibitory effects of iloprost on monocytic cell tissue factor expression.

Methods

Monocytic tumor THP-1 cells21 were purchased from the American Type Culture Collection, Rockville Pike, Md. Antibiotics and materials for the preparation of cell-culture media were obtained from GIBCO, Grand Island, N.Y. Fetal bovine serum was obtained from HyClone, Logan, Utah. Forskolin, 3-isobutyl-1-methylxanthine (IBMX), dibutyryl cAMP, and dibutyryl cGMP were obtained from Sigma Chemical Co., St. Louis, Mo. Bacterial endotoxin (lipopolysaccharide B; Escherichia coli 0111:B4) was obtained from DIFCO, Detroit, Mich. Human recombinant TNF-α and IL-1β were obtained from Boehringer, Indianapolis, Ind. Rabbit brain thromboplastin standard was obtained from Ortho, Raritan, N.J. Human plasma deficient in factor VII was obtained from Helena Labs, Beaumont, Tex.
Cell Culture and Incubations

The cells were grown in RPMI-1640 medium containing 100 μg/ml streptomycin, 100 units/ml penicillin, 10% fetal bovine serum, and 10 mM N-2-hydroxyethylpiperazine-N-2’-ethanesulfonic acid (HEPES), pH 7.4. Serum was heated at 56°C for 30 minutes to inactivate complement. For the experiments, cells were seeded into 24-well dishes at a density of 5 × 10^5 cells/ml and incubated for 30 minutes at 37°C with growth medium containing varying concentrations of prostacyclin analogues or other inhibitors. Iloprost and cicaprost were supplied by Schering AG, Berlin, FRG. Ciprostene was provided by Berlex, Cedar Knolls, N.J. Cicaprost and carbacyclin were provided by Upjohn Diagnostics, Kalamazoo, Mich.

Flow Cytometry

Fluorescent flow cytometry was carried out essentially according to the method of Carson et al.2 Briefly, THP-1 cells were incubated with or without 100 nM iloprost for 30 minutes and then incubated for 4 hours longer with 100 ng/ml endotoxin. After washing, a sample of the cell suspension was tested for surface procoagulant activity. The remaining cells were incubated with normal human serum for 15 minutes to block the Fe binding sites. The serum contained 10 mM EDTA to block factor VII binding. Cells were then washed twice with Tris-saline buffer and incubated with 20 μg/ml of a fluorescein isothiocyanate (FITC)—conjugated monoclonal antibody to human tissue factor (4508CI, generously provided by Dr. Richard Hart, American Diagnostica) or FITC-conjugated nonimmune immunoglobulin G (IgG). Incubations were carried out at 4°C for 30 minutes. The cells were then washed twice, and the fluorescence intensity of 5–10 × 10^4 cells was measured at 488 nm in a FACScan flow cytometer (Becton Dickinson, Rutherford, N.J.) equipped with an air-cooled argon ion laser. The instrument was gated to the appropriate cell size by comparing forward-angle versus 90° light scatter.

Data Analysis

Statistical analysis of data was performed by Student's t test. Differences were considered significant at the 95% confidence level or higher. I_50 (the concentration causing 50% inhibition) values were calculated from a plot of percent inhibition versus log of inhibitor concentration. Percent inhibition was calculated as

\[
100 \times \left[\frac{(\text{challenged} - \text{control}) + \text{inhibitor}}{(\text{challenged} - \text{control}) - \text{inhibitor}}\right]
\]

Results

Exposure of THP-1 cells for 4 hours to 5 ng/ml TNF-α produced significant (p<0.005) twofold increases in surface tissue factor activity. Similarly, exposure to 2 units/ml IL-1β produced three- to fivefold increases. Preliminary experiments established that these were submaximal concentrations for both agents. As shown in Figure 1, preincubation of the cells for 30 minutes with 100 pM iloprost slightly enhanced IL-1β- and TNF-α-induced activity, although only the latter...
enhancement was statistically significant. Higher concentrations of iloprost strongly inhibited cytokine-induced tissue factor expression; at 100 nM the stimulatory effects of both cytokines were essentially abolished. An \( I_50 \) of approximately 5 nM was obtained against each agent, identical to that observed with endotoxin-induced tissue factor expression.15

The ability of three other prostacyclin analogues to inhibit monocyctic cell tissue factor induction was investigated. As shown in Figure 2, an orally active analogue, cicaprost,24 was equipotent with iloprost in inhibiting endotoxin-induced tissue factor expression. Carbacacyclin and ciprostene were approximately two orders of magnitude less potent. Thus, ciprostene at concentrations of 100 nM or higher inhibited tissue factor activity in the absence of endotoxin, whereas iloprost essentially abolished both responses. Lesser changes in monoclonal antibody binding relative to those obtained from cells treated with cytokines alone, as follows: *\( p < 0.05 \), **\( p < 0.005 \).

indicating increased expression of tissue factor antigen on the cell surface (Figure 4B). Pretreatment of the cells for 30 minutes with 100 nM iloprost abolished the endotoxin effect (Figure 4C). Specific cellular fluorescence intensity agreed reasonably well with the procoagulant activities of these cells, which were as follows: control, 56 units/\( 10^6 \) cells; endotoxin treated, 336 units/\( 10^6 \) cells; and iloprost followed by endotoxin, 61 units/\( 10^6 \) cells. Thus, procoagulant activity increased sixfold and fluorescence intensity increased threefold after endotoxin exposure, whereas iloprost essentially abolished both responses. Lesser changes in monoclonal antibody binding relative to those in procoagulant activity have also been noted by Carson et al.23 The role of cAMP in the inhibition of tissue factor expression by iloprost was then explored. Measurements of cAMP in THP-1 cells were made after a 30-minute incubation, the time point at which the challenging agents would routinely be added. Basal levels of cAMP were low (0.9±0.2 pmol/10^6 cells) and increased modestly to 2.8±0.8 pmol/10^6 cells after exposure to 500 pM iloprost (Figure 5). At this concentration iloprost had no observable effect on procoagulant activity. Higher concentrations induced dramatic increases in cAMP; 5 nM iloprost induced a 27-fold increase, and 50 nM induced a 95-fold increase, to 23.3±7.0 and 83.3±14.9 pmol/10^6 cells, respectively. Iloprost therefore increased cAMP levels in THP-1 cells over the same concentration range at which it inhibited tissue factor expression.

As expected, the stimulation of cAMP by iloprost was significantly enhanced in the presence of 200 \( \mu \)M IBMX, an inhibitor of phosphodiesterase. Thus, 500 pM, 5 nM, and 50 nM iloprost induced ninefold, 69-fold, and 234-fold increases in cAMP, respectively, relative to untreated cells (Figure 5). Importantly,
IBMX also strongly potentiated the inhibitory effects of iloprost on monocyctic cell tissue factor expression. Thus, although iloprost alone had no effect at 50 and 500 pM, it produced approximately 30% inhibition at 50 pM (not shown) and completely inhibited tissue factor expression at 500 pM when combined with IBMX (Figure 6). Indeed, iloprost combined with IBMX was more potent than would be predicted from its effects on cAMP levels. Thus, 500 pM iloprost plus IBMX raised cAMP levels to 8.0±2.5 pmol/10^6 cells and abolished tissue factor expression, whereas 5 nM iloprost alone increased cAMP levels to 23.3±7.0 pmol/10^6 cells and produced only 70% inhibition. This apparent discrep-
Further experiments showed that the effects of iloprost could be mimicked by agents that elevate intracellular cAMP by receptor-independent mechanisms. These included forskolin, which is thought to directly activate the catalytic subunit of adenylate cyclase, and dibutyryl cAMP, a membrane-penetrable analogue of cAMP. As shown in Figure 7, both endotoxin- and TNF-α-induced tissue factor expression in THP-1 cells was almost completely inhibited by 100 μM forskolin or 1 mM dibutyryl cAMP; in contrast, 1 mM dibutyryl cGMP had no effect. Sodium butyrate at 1 mM was similarly inactive (not shown).

Discussion

Our previous studies have shown that iloprost potently inhibits endotoxin-induced expression of tissue factor procoagulant activity by human peripheral blood monocytes and THP-1 monocytic cells. Iloprost also blunted further increases in procoagulant activity in cells already exposed to endotoxin, although higher concentrations were required under these conditions. Iloprost was approximately one order of magnitude more potent than prostaglandin E2, which has also been shown to inhibit monocyte tissue factor expression.

We have now extended these observations to show that iloprost inhibits tissue factor induction by two cytokines, TNF-α and IL-1β, with the same potency observed with induction by endotoxin. Furthermore, the inhibitory properties were shared by three other prostacyclin analogues, cicaprost, ciprostene, and carbacyclin. Thus, inhibition of tissue factor expression by monocytes in response to various stimuli appears to be a general property of this class of drugs.

The IC50 values of 5 and 500 nM obtained for iloprost and carbacyclin, respectively, are very close to the reported dissociation constants (8 nM and 528 nM, respectively) for the binding of these agents to the prostacyclin receptor on platelet membranes. These observations suggest that prostacyclin itself is likely to...
be the endogenous ligand and may, therefore, be a physiological regulator of monocyte tissue factor expression. In view of the suggested relation between tissue factor and the receptors for several growth factors and cytokines, it is reasonable to ask whether prostacyclin may have other effects on monocyte function and whether the monocyte, like the platelet, may be a primary target of this agent.

The present studies also provide additional insight into the mechanism of action of iloprost. Three lines of evidence suggest that iloprost, like prostacyclin, exerts its effects at least in part via stimulation of adenylate cyclase. First, iloprost increased cAMP levels in THP-1 cells over the same concentration range at which it inhibited tissue factor expression. Second, the effects of iloprost on both cAMP and tissue factor were potentiated by an inhibitor of phosphodiesterase. Third, both endotoxin- and TNF-α-induced tissue factor expression were effectively inhibited by other agents known to elevate cAMP. The latter observation confirms earlier reports of variable inhibition by such agents on tissue factor activity in human peripheral blood monocytes. The ability of iloprost to block the expression of tissue factor antigen on the surface of THP-1 cells (Figure 4) would be consistent with a role of the inhibition of tissue factor synthesis, perhaps by interference with the process of gene transcription and/or stabilization of tissue factor mRNA. Further studies are required to clarify this point.

Increased tissue factor expression by monocytes may contribute to the disseminated intravascular coagulation associated with disorders such as allograft rejection, bacterial infection, carcinoma, and immune disease (for reviews, see References 1 and 30). More recently, it has been suggested that increased monocyte tissue factor activity may also contribute to the postoperative thrombotic state. Other investigators have shown that the development of fulminant hepatitis is accompanied by increased monocyte procoagulant activity in a virally infected mouse model. Furthermore, both the hepatitis and the monocyte procoagulant activity are inhibited by the administration of prostaglandin E2 and its 16,16-dimethyl analogue. The potency of iloprost and its effectiveness against multiple challenging agents suggest that it may also be valuable in the treatment of monocyte-driven coagulopathies, particularly if used in conjunction with a clinically effective phosphodiesterase inhibitor.

Acknowledgments

We wish to thank Mr. Andy Toledo for his help in culturing the THP-1 cells and Ms. Ana Maria Manz and Mr. Franco Arias for their excellent technical assistance. We also thank Mr. Art Alamo of the University of Miami School of Medicine, Miami, Fla., for his help in performing the flow cytometry experiments. Finally, we wish to thank Dr. Richard Hart of American Diagnostica, Greenwich, Conn., and Dr. Steven Carson of the University of Nebraska Medical Center, Omaha, Neb., for their generous gifts of monoclonal antibodies to human tissue factor.

References

Prostacyclin analogues inhibit tissue factor expression in the human monocytic cell line THP-1 via a cyclic AMP-dependent mechanism.

D J Crutchley, D E Solomon and L B Conanan

doi: 10.1161/01.ATV.12.6.664
Arteriosclerosis, Thrombosis, and Vascular Biology is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 1992 American Heart Association, Inc. All rights reserved.
Print ISSN: 1079-5642. Online ISSN: 1524-4636

The online version of this article, along with updated information and services, is located on the World Wide Web at:
http://atvb.ahajournals.org/content/12/6/664

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Arteriosclerosis, Thrombosis, and Vascular Biology can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Arteriosclerosis, Thrombosis, and Vascular Biology is online at:
http://atvb.ahajournals.org//subscriptions/